

Safety

Primary tumor cells should be contained in a Class II biological safety cabinet and handled using Biosafety Level 2 (BSL-2) work practices and facilities (2).

Cell Culture Media and Antibiotics

- Recommended cell culture media (3):
 - DMEM-F12 medium with L-glutamine
 - 5 mM HEPES
 - 0.1% sodium bicarbonate
 - 0.4% BSA or 10% FBS
- Antibiotics
 - Normocin (Invivogen; 100 µg/mL)
- Supplements
 - 20 ng/ml EGF
 - 10 ng/ml bFGF
 - Supplements recommended for unthawing samples
 - DNase I (1 mg/mL), resuspended in PBS without calcium or magnesium and filter-sterilized.

Unthawing Instructions

- On the day of the thaw, pre-warm 19 mls of recommended cell culture media in a 37°C incubator. Divide the media into two 15-mL conical tubes (1 tube with 9 mLs of media; 1 tube with 10 mLs of media).
- Quickly thaw the DTC sample in a 37-40°C water bath until 2mm crystals remain. Move sample through the water to speed thawing.
- Slowly add the 1-mL DTC sample to 9 mls of pre-warmed media. Gently mix by inversion (do not vortex).
- Centrifuge the conical tube at 300 x g for 5-10 minutes (no brake). Carefully remove the supernatant.
- After the initial wash to remove DMSO from the sample, gently resuspend the pellet in the remaining 10 mLs of pre-warmed cell culture media.
- If the cells clump together, perform the following:
 - Add DNase I (1 mg/mL) to the cell culture media.
 - Incubate at 37°C for 5-10 minutes to facilitate digestion of released DNA.
 - Centrifuge the conical tube at 300 x g for 5-10 minutes (no brake). Carefully remove supernatant containing DNase I.
 - Gently resuspend the pellet in 10 mLs of pre-warmed cell culture media. Go to next step.
- Allow cells to rest for at least 1 hour in the media at 37°C before checking the viability and plating.
- Check the viability and cell number of the sample.
- Plate the cells according to assay requirements.

Culturing Primary Cells

- For sphere formation assays, using untreated tissue culture flasks is recommended to reduce cell adherence (4).
- The media recipe listed above is a base media for thawing and acclimating lung tumor cells, in preparation for cell culture.
- Additional supplementation will be required to expand lung tumor primary cells *in vitro* (3), depending on the end-goal of the assay. Please see the literature for supplementation recommendations, depending on the specific needs of the cell types targeted for expansion.

References

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2. Centers for Disease Control and Prevention. 2009. Biosafety in Microbiological and Biomedical Laboratories (BMBL). U.S. Dept. of Health and Human Services. 5th ed.
3. Eramo, A.; F. Lotti; G. Sette; E. Pillozzi; M. Biffoni; A. Di Virgilio; C. Conticello; L. Ruco; C. Peschle; R. De Maria. 2008. Identification and expansion of the tumorigenic lung cancer stem cell population. *Cell Death and Differentiation*. 15:504-514.
4. Pan, J.; Q. Zhang; Y. Wang; M. You. 2010. 26S proteasome activity is down-regulated in lung cancer stem-like cells propagated *in vitro*. *PLoS One*. 5:e13298