# TO PROCESS AGILENT DATA GENERATED WITH FEATURE EXTRACTION:

To process data, we first need to load data into Nexus. You can load data directly by loading input data files via the file chooser (see DATA LOADING VIA THE ONE-CLICK METHOD below) or you can load your data by creating and loading a sample descriptor (see DATA LOADING USING A SAMPLE DESCRIPTOR below). The one-click data loading method is the best way to load data for most cases. Only in rare situations a user will need to use the sample descriptor method to load data (e.g. if using replicate data, you will need to use the sample descriptor method). With the one-click method, you will select input files via a file chooser and will assign any factors (clinical annotations) after the data is loaded. With the sample descriptor method, you will specify the input file location and assign factors to your samples by specifying them in the sample descriptor. When data is loaded factors will already be associated and will appear in the **Data Set** tab. Choose one of the two methods below to load your data.

### DATA LOADING VIA THE ONE-CLICK METHOD

- 1. Run Nexus and click the **Create a New Project** or **Open Existing Project** button. If creating a new project, in the resulting dialog, specify a name for your project and choose a genome and then the appropriate genome build based on the mapping information for your probes. If adding additional samples to an existing project (of the same genome build), select your project folder in the resulting dialog.
- 1. Click on the Load button and select Load Data in the Data Set tab. Select Agilent FE from the drop down in the resulting Add Sample Data window. A number of options appear below the checkbox Set default processing settings based on the following. This "guide" assists in adjusting the parameters in the Settings and is based on the type of data you are loading. If you do not want to use this guide, uncheck the box (all the options will be grayed out) and *skip to Step 6*. If you use the "guide", making selections here adjusts parameters in the "Analysis" section as well as "Systematic Correction" section of the Settings window. The Help buttons to the left of the options bring up pop ups with additional information on what changes the selection will make to the settings parameters.
- 2. Select one of the radio buttons depending on the type of data you are loading: For mosaic samples such as cancer (possible increased false-positive rate) or For non-mosaic samples. The selection here adjusts the thresholds used to make calls (e.g. High Gain, Gain, Loss, Big Loss).
- 3. Adjust the **Stringency** slider to one of 3 positions: **Lenient**, **Average**, or **Stringent**. This adjusts the sensitivity of the algorithm in making calls and sets the appropriate value for the **Significance Threshold** setting. "Lenient" will generate more calls and "Stringent", less calls (more specificity).
- 2. Choose if you want to apply systematic correction by checking or unchecking the checkbox for Perform Systematic Correction. This corrects the wavy pattern often present in the probe distribution. Since the correction is based on probes used, the specific array is needed. Select the array from the Array Types dropdown menu. Arrays listed here are based on the files and folders available in the SystematicCorrection directory of the Organism->genome build folders. If your array type is not listed in the dropdown, please contact BioDiscovery Support (support@biodiscovery.com) to obtain the necessary files.

- 3. Click on Select Files to select your input files via the file chooser and click Done. Your samples will be listed in the Data Set tab. If you want to assign any factors to your samples, you can do so with the selection in the Add Factors button dropdown. With Add Factor, you will manually enter factor values for each sample by typing them individually or by selecting column cells and copying and pasting values from a spreadsheet (see the user manual for "Allow column selection"). With Load Factors, you will select a tab delimited text file containing sample names and any factors assigned to them. A column called Sample Name containing names matching the sample names in the project is required and any number of factor columns with factor names as column headers (DO NOT prefix the column header with Factor:). See Loading Factors in the Data Set Tab section of the user's manual.
- 4. If you used the guide, files are automatically loaded and processed after you selected files and clicked **Done** in the previous step. If you did not use the guide, click **View** to process the data and see the results.

## DATA LOADING USING A SAMPLE DESCRIPTOR

- The sample descriptor file is a tab delimited text file used to assign factors to samples and load data into Nexus. In the **Templates** folder of the Nexus installation directory, you will find several arrays types that Nexus is able to process. Open the folder titled **Agilent** and select the appropriate template file based on your input data. If you have technical replicates (e.g. dye-swap) refer to the document called *Processing Replicate Data* for further details on processing such data. If you have dye-swap data, open **AgilentFE-DyeSwap-Template.txt** file, otherwise open **AgilentFE-Template.txt**. It may be easiest to edit the sample descriptor template file by opening it in a spreadsheet application such as Excel.
- 2. In the first column, Sample Name, specify a unique name for each sample that you are loading (e.g. sample1, sample2, etc.). Note that you cannot use the following special characters /\:\*?"<>| within the sample name. In the second column, File, specify the location of your input data file. You can use either the full path (e.g. C:\Program Files\BioDiscovery\Nexus\My Projects\sample1.txt) or a relative path (relative to where you will save this sample descriptor file). If the descriptor is in the same folder as the sample data files then you can specify just the name (e.g. sample1.txt) without any path qualifiers. Those are the only required columns. You can add an unlimited number of additional columns to specify Factors (clinical annotations) for each sample. The column header is Factor: followed by a name for the Factor. Save this file with a new name and make sure to save it as a tab-delimited text file.

Example of a sample descriptor:

Data Type:	Agilent FE		
Sample Name	File	Factor:Gender	Factor:Tumor Type
sample 1	C:\Nexus Projects\Brain Tumor\sample1.txt	M	GBM
sample 2	C:\Nexus Projects\Brain Tumor\sample2.txt	M	GBM
sample 3	C:\Nexus Projects\Brain Tumor\sample3.txt	F	AOA
sample 4	C:\Nexus Projects\Brain Tumor\sample4.txt	M	AO

### Example of a sample descriptor with technical replicates:

Data Type:	Agilent FE			
Sample Name	File	File2	File3	Factor:Gender
sample 1	C:\BrainTumorProject\array1.txt	C:\BrainTumorProject\array11.txt	C:\BrainTumorProject\array31.txt	M
sample 2	C:\BrainTumorProject\array2.txt	C:\BrainTumorProject\array12.txt		F
sample 3	C:\BrainTumorProject\array3.txt	C:\BrainTumorProject\array13.txt		M

#### Example of a sample descriptor with technical replicates from dye-swap experiments:

Data Type:	Agilent FE			
Sample Name	File	-File2	Factor:Gender	Factor:Tumor Type
sample 1	C:\BrainTumorProject\array1.txt	C:\BrainTumorProject\array1R.txt	M	GBM
sample 2	C:\BrainTumorProject\array2.txt	C:\BrainTumorProject\array2R.txt	F	AOA
sample 3	C:\BrainTumorProject\array3.txt	C:\BrainTumorProject\array3R.txt	M	AOA

- 3. Run Nexus and click the **Create a New Project** or **Open Existing Project** button. If creating a new project, in the resulting dialog, specify a name for your project and choose a genome and then the appropriate genome build based on the mapping information for your probes. If adding additional samples to an existing project (of the same genome build), select your project folder in the resulting dialog.
- 4. On the next screen, click the **Load** button, select **Load Descriptor** from the dropdown and choose the sample descriptor file you saved in step 2 above. You will see all your samples loaded into Nexus.
- 5. Click **View** to process the data and see the results.