

Tumor Type Lung adenocarcinoma

| Date of Birth | Medical Facility | | |
|------------------|-----------------------|--------------------|-------|
| Sex | Ordering Physician | Specimen Received | |
| FMI Case # | Additional Recipient | Date of Collection | |
| Medical Record # | Medical Facility ID # | Specimen Type | Blood |
| Specimen ID | Pathologist | | |

ABOUT THE TEST:

FoundationACT[™] (Assay for Circulating Tumor DNA) is a next-generation sequencing (NGS) assay that identifies clinically relevant genomic alterations in circulating tumor DNA and matches them to targeted therapies and clinical trials.

PATIENT RESULTS

TUMOR TYPE: LUNG ADENOCARCINOMA

5 genomic alterations

3 therapies associated with potential clinical benefit

2 therapies associated with lack of response

4 clinical trials

Genomic Alterations Identified[†] *EGFR* L858R, T790M *TP53* H193Y, H214R – equivocal[#], R248Q

For a complete list of the genes assayed and performance specifications, please refer to the Appendix See Appendix for details

THERAPEUTICIMPLICATIONS

| Genomic Alterations | Allele Frequency | FDA-Approved Therapies (in patient's tumor type) | FDA-Approved Therapies (in another tumor type) | Potential Clinical Trials |
|---|-----------------------|---|---|----------------------------------|
| EGFR L858R, T790M | 16.3%, 11.2% | (-) Erlotinib ‡ (-) Gefitinib ‡ Osimertinib | Cetuximab Panitumumab | Yes, see clinical trials section |
| TP53 H193Y, H214R - equivocal, R248Q | 3.8%, 0.12%, 0.54% | None | None | None |

‡ (-) Patient may be resistant to therapy

Note: Genomic alterations detected may be associated with activity of certain FDA-approved drugs; however, the agents listed in this report may have little or no evidence in the patient's tumor type. Neither the therapeutic agents nor the trials identified are ranked in order of potential or predicted efficacy for this patient, nor are they ranked in order of level of evidence for this patient's tumor type.

N/A= Not Applicable; Allele Frequency is not applicable for copy number amplifications or rearrangements.



Tumor Type Lung adenocarcinoma

GENOMICALTERATIONS

GENE ALTERATION

EGFR

L858R. T790M

INTERPRETATION

Gene and Alteration: EGFR encodes the epidermal growth factor receptor, which belongs to a class of proteins called receptor tyrosine kinases. In response to signals from the environment, EGFR passes biochemical messages to the cell that stimulate it to grow and divide ¹. EGFR L858R is a mutation within the kinase domain and has been shown to activate kinase activity and confer sensitivity to EGFR tyrosine kinase inhibitors, such as erlotinib and gefitinib^{2,3,4}. The EGFR T790M resistance mutation suggests that this tumor may be resistant to the first-generation EGFR inhibitors gefitinib and erlotinib⁵ and may be less responsive to the second-generation EGFR inhibitor afatinib⁶. The amplification of EGFR with the T790M mutation has also been linked to resistance to the irreversible EGFR inhibitor dacomitinib⁷.

Frequency and Prognosis: EGFR alterations have been reported in 13-35% of lung adenocarcinomas^{8,9}. EGFR amplification has been documented in up to 62% of non-small cell lung cancer (NSCLC), and has been correlated with EGFR protein expression as measured by immunohistochemistry, although this correlation is not consistent for low level gene amplification^{10,11,12,13}. EGFR protein expression or overexpression has been reported in up to 70% of NSCLC tumors¹⁴. EGFR mutations have been reported to predict improved survival in patients with resected Stage 1-3 lung adenocarcinoma¹⁵ or resected Stage 1 NSCLC¹⁶.

Potential Treatment Strategies: EGFR activating mutations or amplification may predict sensitivity to EGFR inhibitors, including osimertinib, afatinib, erlotinib, gefitinib, cetuximab, panitumumab, and lapatinib^{17,18,19,20,21}. However, strong clinical evidence indicates that the EGFR T790M mutation confers resistance to gefitinib and erlotinib⁵, and preclinical studies indicate that cells expressing EGFR T790M are resistant to lapatinib^{22,23}. T790M has also been reported in 48% (20/42) of patients with acquired afatinib resistance⁶, suggesting patients with T790M may be less responsive to this therapy 6,24,25 ; however, disease control rates of more than 50% have been reported for patients with erlotinib- or gefitinib-resistant NSCLC treated with afatinib²⁶, including T790M-positive patients²⁷. A combination of afatinib and cetuximab has shown clinical efficacy for T790M-positive NSCLC^{28,29}, although careful dosing may be required^{29,30}. Third-generation EGFR inhibitors, such as osimertinib and rociletinib, selectively target mutated forms of EGFR including EGFR T790M; osimertinib is FDA approved to treat patients with EGFR T790M-positive advanced NSCLC who have progressed on EGFR inhibitor therapy¹⁷. Osimertinib and rociletinib, respectively, achieved objective response rates (ORR) of 61% and 59% in T790M-positive cases and 21% and 29% in T790M-negative cases^{17,31}. Resistance to EGFR inhibition may arise by reactivation of the MAPK pathway, and preclinical evidence suggests that co-targeting EGFR and MAPK signaling may retard the development of acquired resistance to third-generation EGFR inhibitors^{32,33,34}. Necitumumab is an anti-EGFR antibody that is FDA approved to treat metastatic squamous NSCLC in combination with gemcitabine and cisplatin. Addition of necitumumab increased overall and progression-free survival in patients with squamous NSCLC relative to chemotherapy alone; however, it exhibited a poor tolerability profile in non-squamous NSCLC, and EGFR expression has not been demonstrated to be predictive of clinical benefit in NSCLC^{35,36}. Preclinical studies have reported that EGFR-mutant cells are sensitive to HSP90 inhibitors^{37,38,39,40}. Clinical studies of HSP90 inhibitors, alone and in combination with EGFR inhibitors, have reported response rates ranging from 0% to 18% in patients with NSCLC harboring EGFR mutations (Garon et al., 2012; ASCO Abstract 7543)^{41,42,43,44}, although combination treatment was deemed too toxic⁴⁴. The reovirus Reolysin, which targets cells that harbor activated RAS signaling due to alterations in RAS genes or upstream activators such as EGFR^{45,46,47}, is also in clinical trials in some tumor types. A trial of Reolysin in combination with paclitaxel and carboplatin in patients with NSCLC harboring activating KRAS or EGFR alterations reported significantly improved response and survival rates compared to assumed historical data for paclitaxel and carboplatin alone⁴⁸.



GENE ALTERATION

TP53

H193Y, H214R -

equivocal, R248Q

INTERPRETATION

Gene and Alteration: Functional loss of the tumor suppressor p53, which is encoded by the TP53 gene, is common in aggressive advanced cancers ⁴⁹. Mutations affecting the DNA binding domain (aa 100-292), the tetramerization domain (aa 325-356), or the C-terminal regulatory domain (aa 356-393), such as observed here, are thought to disrupt the transactivation of p53-dependent genes and are predicted to promote tumorigenesis^{50,51,52,53}. Germline mutations in TP53 are associated with the very rare disorder Li-Fraumeni syndrome and the early onset of many cancers^{54,55,56,57,58,59}. Estimates for the prevalence of germline TP53 mutations in the general population range from 1:5,000⁶⁰ to 1:20,000⁵⁹, and in the appropriate clinical context, germline testing of TP53 is recommended.

Frequency and Prognosis: TP53 is one of the most commonly mutated genes in lung cancer, and mutations in this gene have been reported in 43-80% of non-small cell lung cancers (NSCLCs) ^{61,62,63,64,65} and specifically in 45% of lung adenocarcinoma samples^{66,67}. Mutations in TP53 have been associated with lymph node metastasis in patients with lung adenocarcinoma⁶⁸.

Potential Treatment Strategies: There are no approved therapies to address TP53 mutation or loss. However, tumors with TP53 loss of function alterations may be sensitive to the WEE1 inhibitor AZD1775^{69,70,71,72}, therapies that reactivate mutant p53 such as APR-246⁷³, or p53 gene therapy and immunotherapeutics such as SGT-53^{74,75,76,77} and ALT-801 (Hajdenberg et al., 2012; ASCO Abstract e15010). Combination of AZD1775 with paclitaxel and carboplatin achieved significantly longer progression-free survival than paclitaxel and carboplatin alone in patients with TP53-mutant ovarian cancer (Oza et al., 2015; ASCO Abstract 5506). Furthermore, AZD1775 in combination with carboplatin achieved a 27% (6/22) response rate and 41% (9/22) stable disease rate in patients with TP53-mutant ovarian cancer refractory or resistant to carboplatin plus paclitaxel (Leijen et al., 2015; ASCO Abstract 2507). In a Phase 1 clinical trial, 8 of 11 evaluable patients receiving SGT-53 as a single agent exhibited stable disease⁷⁸. Clinical trials of SGT-53 in combination with chemotherapy are underway. Additionally, the combination of a CHK1 inhibitor and irinotecan reportedly reduced tumor growth and prolonged survival in a TP53 mutant, but not TP53 wild-type, breast cancer xenotransplant mouse model⁷⁹. Kevetrin has also been reported to activate p53 in preclinical studies and might be relevant in the context of mutant p53 (Kumar et al., 2012; AACR Abstract 2874). Clinical trials of these agents are under way for some tumor types for patients with a TP53 mutation.



Tumor Type Lung adenocarcinoma

THERAPIES

| FDA-APPROVED THERAPIES IN PATIENT TUMOR TYPE | | | | | |
|--|---|--|--|--|--|
| THERAPY | SUMMARY OF DATA IN PATIENT TUMOR TYPE | | | | |
| Osimertinib | Approved Indications: Osimertinib is an irreversible EGFR tyrosine kinase inhibitor (TKI) that is selective for EGFR TKI-sensitizing mutations and the EGFR T790M mutation. It is FDA approved to treat patients with metastatic EGFR T790M-positive non-small cell lung cancer (NSCLC) and disease progression on or after EGFR TKI therapy. | | | | |
| | Gene Association: EGFR TKI-sensitizing mutations and/or the EGFR T790M mutation may predict sensitivity to osimertinib ^{17,80} . T790M-positive patients showed higher response rates than T790M-negative cases in a Phase 1 study for patients with acquired EGFR TKI resistance (61% vs. 21%) ¹⁷ . | | | | |
| | Supporting Data: Osimertinib has been studied primarily for the treatment of EGFR-mutant NSCLC. Phase 2 studies of osimertinib demonstrated objective response rates (ORR) of 57-61% and disease control rates (DCR) of 90-92% for patients with T790M-positive advanced NSCLC who had progressed on prior EGFR TKI therapy; most objective responses (96%) were ongoing at the median 4-month follow-up (Yang et. al., 2015; WCLC Abstract 943, Mitsudomi et al., 2015; WCLC Abstract 1406). In the Phase 1 expansion cohort with the approved dose of osimertinib (80 mg), the ORR was 54% (32/59), the median duration of response was 12.4 months, and the median progression-free survival (PFS) was 13.5 months for patients with T790M-positive NSCLC (Janne et al., 2015; DOI: 10.1093/annonc/mdv128.05). This trial reported an ORR of 21% and median PFS of 2.8 months for T790M-negative cases with acquired EGFR TKI resistance ¹⁷ . Treatment-naïve patients with EGFR- mutant NSCLC achieved an ORR of 60% (18/30) and a DCR of 93% (28/30) (Ramalingnam et al., 2015; ASCO Abstract 8000). A Phase 1b study combined osimertinib with the investigational immunotherapy durvalumab, MEK inhibitor selumetinib, or MET inhibitor savolitinib, and observed partial responses (PR) for each of the combinations (9/14 PR with durvalumab, 9/23 PR with selumetinib, 6/11 PR with savolitinib) (Ramalingam et al., 2015; ASCO Abstract 2509). Osimertinib is being compared with erlotinib or gefitinib as first-line treatment for EGFR-mutant NSCLC (NCT02296125). | | | | |
| | · · | | | | |
| THERAPIES ASSOCIATE | ED WITH LACK OF RESPONSE | | | | |
| THERAPY | SUMMARY OF DATA IN PATIENT TUMOR TYPE | | | | |
| Erlotinib | Approved Indications: Erlotinib is a small molecule inhibitor of EGFR. It is FDA approved for the treatment of non-small cell lung cancer (NSCLC) and pancreatic cancer. | | | | |

Gene Association: EGFR activating mutations or amplification may predict sensitivity to erlotinib. However, the EGFR T790M mutation has been associated with resistance to erlotinib, leading to the suggestion that these drugs will not be effective a tumor that contains the T790M mutation, particularly in patients who have already received erlotinib or gefitinib 5 .

Supporting Data: The approval of erlotinib in NSCLC is based on a Phase 3 randomized trial demonstrating prolonged overall survival for unselected patients with NSCLC treated with erlotinib compared to standard chemotherapy ⁸¹. Furthermore, several randomized Phase 3 trials have shown a significant improvement in response and progression-free survival for this class of medications compared with combination chemotherapy in patients with known EGFR mutations. This includes the EURTAC trial of erlotinib vs. platinum-based chemotherapy¹⁸.

Gefitinib

Approved Indications: Gefitinib targets the tyrosine kinase EGFR and is FDA approved to treat nonsmall cell lung cancer (NSCLC) harboring exon 19 deletions or exon 21 (L858R) substitution mutations in EGFR.

For more comprehensive information please log on to the Interactive Cancer Explorer™ To set up your Interactive Cancer Explorer account, contact your sales representative or call (888) 988-3639.

Electronically Signed by Shakti Ramkissoon, M.D. | Jeffrey S. Ross, M.D., Medical Director | CLIA Number: 22D2027531 | 25 May 2016 Foundation Medicine, Inc., 150 2nd Street, 1st Floor, Cambridge, MA 02141 | 1.888.988.3639 **Gene Association:** EGFR activating mutations or amplification may predict sensitivity to therapies such as gefitinib. Clinical studies have consistently shown significant improvement in response rates and progression-free survival for patients with EGFR-mutated NSCLC treated with gefitinib, compared to chemotherapy ^{82,83,84,85,86,87}. However, the EGFR T790M mutation has been associated with resistance to gefitinib, leading to the suggestion that these drugs will not be effective in a tumor that contains the T790M mutation, particularly in patients who have already received erlotinib or gefitinib⁵.

Supporting Data: Gefitinib achieved an objective response rate of 69.8% and an overall survival of 19.2 months as first-line treatment of Caucasian patients with NSCLC and EGFR sensitizing mutations, which were mostly EGFR exon 19 deletions and EGFR L858R¹⁹. In the retrospective analysis of a Phase 3 study in Asia, gefitinib increased progression-free survival in a subgroup of patients with EGFR mutation-positive NSCLC as compared with carboplatin/paclitaxel doublet chemotherapy (hazard ratio for progression 0.48)^{85,88}.

| ADDITIONAL THER | APIES – FDA-APPROVED IN OTHER TUMOR TYPES |
|-----------------|---|
| THERAPY | SUMMARY OF DATA IN OTHER TUMOR TYPE |
| Cetuximab | Approved Indications: Cetuximab is a monoclonal antibody that targets EGFR. It is FDA approved for the treatment of head and neck squamous cell carcinoma (HNSCC) and KRAS wild-type metastatic colorectal cancer (CRC). Gene Association: EGFR activating mutations or amplification may confer sensitivity to EGFR inhibitory antibodies such as cetuximab. |
| | Supporting Data: In previously untreated patients with non-small cell lung cancer (NSCLC), the FLEX study demonstrated that in NSCLC tumors with high expression of EGFR, treatment with cetuximab plus chemotherapy resulted in longer overall survival compared to chemotherapy alone ²¹ . There was no clear association between cetuximab response and EGFR mutations in the FLEX trial ²¹ . In a Phase 2 study of 31 patients with Stage 3 NSCLC, addition of cetuximab to radiotherapy and chemotherapy produced an overall response rate of 67%; EGFR gene copy number was not predictive of efficacy outcome ⁸⁹ . A Phase 3 study of 938 patients with progressive NSCLC after platinum-based therapy concluded that, in unselected patients, the addition of cetuximab to chemotherapy was not recommended in this second-line setting ⁹⁰ . Cetuximab is also being studied as part of a therapeutic regimen for patients with EGFR mutations who develop secondary resistance to erlotinib or gefitinib. A Phase 1b study combining afatinib and the anti-EGFR antibody cetuximab in patients with advanced EGFR-mutant lung cancer with acquired resistance to erlotinib/gefitinib observed an overall objective response rate of 29%, and comparable response rates in both T790M-positive and T790M-negative tumors (32% vs. 25%) ²⁸ . A Phase 1 study of combination erlotinib and cetuximab treatment in patients with NSCLC, including those with squamous tumors, inhibitor-resistant EGFR mutations, and wild-type EGFR, as well as those who had progressed on prior erlotinib treatment, reported partial responses in two of 20 patients and stable disease lasting at least 6 months in three of 20 patients ⁹¹ ; however, in this study a patient identified with an exon 19 deletion and T790M progressed rapidly on cetuximab and erlotinib, consistent with predictions based on computational analysis of T790M ⁹² . |
| Panitumumab | Approved Indications: Panitumumab is a monoclonal antibody that targets EGFR. It is FDA approved for the treatment of KRAS wild-type metastatic colorectal cancer (CRC). |
| | Gene Association: EGFR activating mutations or amplification may confer sensitivity to EGFR inhibitory antibodies such as panitumumab. |



Report Date

Supporting Data: In a Phase 2 trial of advanced non-small cell lung cancer (NSCLC), the addition of panitumumab to paclitaxel/carboplatin did not result in improved clinical benefit ⁹³, and subsequent studies investigating the addition of panitumumab to pemetrexed/cisplatin reported no benefit for patients with wild-type KRAS lung adenocarcinoma⁹⁴. The combination of afatinib and panitumumab has been explored for 2 patients with EGFR T790M NSCLC, with 1 partial response reported³⁰.

Genomic alterations detected may be associated with activity of certain FDA-approved drugs; however, the agents listed in this report may have little or no evidence in the patient's tumor type.



CLINICAL TRIALS TO CONSIDER

IMPORTANT: While every effort is made to ensure the accuracy of the information contained below, the information available in the public domain is continually updated and should be investigated by the physician or research staff. This is not meant to be a complete list of available trials. In order to conduct a more thorough search, please go to www.clinicaltrials.gov and use the search terms provided below. For more information about a specific clinical trial, type the NCT ID of the trial indicated below into the search bar.

GENE RATIONALE FOR POTENTIAL CLINICAL TRIALS

Activating mutations in EGFR have been shown to confer sensitivity to EGFR inhibitors.

However, the presence of the T790M resistance mutation suggests that some inhibitors will be ineffective. Other agents, including third generation EGFR inhibitors and HSP90 inhibitors, may be relevant.

```
L858R, T790M
```

EGFR

Examples of clinical trials that may be appropriate for this patient are listed below. These trials were identified through a search of the trial website clinicaltrials.gov using keyword terms such as "EGFR", "cetuximab", "panitumumab", "afatinib", "osimertinib", "BIBW 2992", "CO-1686", "AZD9291", "PF-00299804", "HSP90", "reolysin", "NSCLC", "lung", "solid tumor", and/or "advanced cancer".

| TITLE | PHASE | TARGETS | LOCATIONS | NCT ID |
|--|---------|--------------|-------------------------------|-------------|
| A Phase I/II, Multicenter, Open-label Study of | Phase | EGFR | Massachusetts, New York, | NCT02108964 |
| EGFRmut-TKI EGF816, Administered Orally in | 1/Phase | | multiple ex-US locations | |
| Adult Patients With EGFRmut Solid | 2 | | | |
| Malignancies | | | | |
| TIGER-3: A Phase 3, Open-label, Multicenter, | Phase 3 | EGFR | California, Florida, Georgia, | NCT02322281 |
| Randomized Study of Oral Rociletinib (CO- | | | Illinois, Maryland, Michigan, | |
| 1686) Monotherapy Versus Single-agent | | | Minnesota, New Jersey, New | |
| Cytotoxic Chemotherapy in Patients With | | | York, Ohio, Oregon, | |
| Mutant EGFR Non-small Cell Lung Cancer | | | Pennsylvania, Texas, Utah, | |
| (NSCLC) After Failure of at Least 1 Previous | | | Virginia, multiple ex-US | |
| EGFR-directed Tyrosine Kinase Inhibitor (TKI) | | | locations | |
| and Platinum-doublet Chemotherapy | | | | |
| A Multi-arm, Phase Ib, Open-Label, Multicentre | Phase 1 | EGFR, PD-L1, | Georgia, Massachusetts, New | NCT02143466 |
| Study to Assess the Safety, Tolerability, | | MEK | York, Tennessee, multiple ex- | |
| Pharmacokinetics and Preliminary Anti-tumour | | | US locations | |
| Activity of AZD9291 in Combination With | | | | |
| Ascending Doses of Novel Therapeutics in | | | | |
| Patients With EGFRm+ Advanced NSCLC Who | | | | |
| Have Progressed Following Therapy With an | | | | |
| EGFR TKI | | | | |
| Phase 1/2 Open-Label Study Of PF-06747775 | Phase | EGFR | California, Connecticut, | NCT02349633 |
| (Epidermal Growth Factor Receptor T790M | 1/Phase | | Pennsylvania, Washington, | |
| Inhibitor) In Patients With Advanced Epidermal | 2 | | Seoul (Korea, Republic of) | |
| Growth Factor Receptor (EGFR) Mutant (Del 19 | | | | |
| Or L858R +/- T790M) Advanced Non-Small Cell | | | | |
| Lung Cancer | | | | |



VARIANTS OF UNKNOWN SIGNIFICANCE

Note: One or more variants of unknown significance (VUS) were detected in this patient's tumor. These variants may not have been adequately characterized in the scientific literature at the time this report was issued and/or the genomic context of these alterations makes their significance unclear. We choose to include them here in the event that they become clinically meaningful in the future.

BTK E41V



GENES ASSAYED IN FOUNDATIONACT™

FoundationACT interrogates the complete exonic sequence of 27 genes, introns of 6 genes involved in rearrangements, and select exons of an additional 34 genes. The assay will be updated periodically to reflect new knowledge about cancer biology.

| DNA Gene List: Entire Exonic Sequence for the Detection of Base Substitutions, Insertions/Deletions, and Copy Number Alterations | | | | | |
|--|----------------------------|---------------------------|----------------------------|--------------------------|----------|
| BRCA1 | CDK4 | ERBB2 | KRAS | MYCN | PTPN11 |
| BRCA2 | CDK6 | ERRFI1 | MDM2 | NF1 | SMO |
| CCND1 | CDKN2A | FGFR1 | MET | PDCD1LG2 (PD-L2) | ТР53 |
| CD274 (PD-L1) | CRKL | FGFR2 | МҮС | PTEN | VEGFA |
| CDH1 | EGFR | FOXL2 | | | |
| | | | | | |
| DNA Gene List: For the I | Detection of Select Rearra | ingements | | | |
| ALK | EGFR | FGFR3 | PDGFRA | RET | ROS1 |
| | | | | | |
| DNA Gene List: Select Ex | onic Sequence for the De | tection of Base Substitut | tions, Insertions/Deletion | ns, and Copy Number Alte | erations |

| DNA Gene List: Select Exonic Sequence for the Detection of Base | Substitutions, | Insertions/Dele | tions, and Copy Number Altera | ations |
|---|----------------|-----------------|-------------------------------|--------|
|---|----------------|-----------------|-------------------------------|--------|

| ABL1 Exons 4-9 | CTNNB1 Exon 3 | GNA11 Exons 4,5 | JAK2 Exon 14 | MTOR Exons 19,30,39,40, 43-45,47,48,53,56 | <i>PIK3CA</i> Exons 2,3,5-8,10,14, 19,21 |
|-------------------------------------|-----------------------|--------------------|-----------------------------|---|--|
| AKT1 Exon 3 | DDR2 Exons 5,17,18 | GNAQ Exons 4,5 | JAK3 Exons 5,11-13,15,16 | MYD88 Exon 4 | RAF1 Exons 3,4,6,7,10,14, 15,17 |
| ALK | ESR1 | GNAS | <i>KIT</i> | NPM1 | <i>RET</i> |
| Exons 20-29 | Exons 4-8 | Exon 1 | Exons 8,11,12,17 | Exons 4-6,8,10 | Exons 11,13-16 |
| ARAF Exons 4,5,7,11,13, 15,16 | EZH2 Exon 16 | HRAS Exons 2,3 | MAP2K1 (MEK1) Exons 2,3 | NRAS Exons 2,3 | <i>TERT</i> (Promoter only) |
| BRAF | FGFR3 | IDH1 | MAP2K2 (MEK2) | PDGFRA | |
| Exons 11-18 | Exons 7,9,14 | Exon 4 | Exons 2-4,6,7 | Exons 12,18 | |
| BTK | <i>FLT3</i> | <i>IDH2</i> | <i>MPL</i> | PDGFRB | |
| Exons 2,15 | Exons 14,15,20 | Exon 4 | Exon 10 | Exons 12-21,23 | |

For more comprehensive information please log on to the Interactive Cancer Explorer™ To set up your Interactive Cancer Explorer account, contact your sales representative or call (888) 988-3639.

Electronically Signed by Shakti Ramkissoon, M.D. | Jeffrey S. Ross, M.D., Medical Director | CLIA Number: 22D2027531 | 25 May 2016 Foundation Medicine, Inc., 150 2nd Street, 1st Floor, Cambridge, MA 02141 | 1.888.988.3639



FOUNDATIONACT[™] PERFORMANCE SPECIFICATIONS

| | Mutant Allele Frequency (MAF) / Tumor Fraction [†] | Sensitivity | Positive Predictive Value (PPV) | |
|---|--|---|------------------------------------|--|
| Pasa Substitutions | ≥0.5% | >98.9% (99.6%-100%)* | > 99.9% (99.6%-100%)* | |
| base substitutions | 0.1% - 0.5% | 67.3% (61.7%-72.5%)* | 93.6% (89.2%-96.3%)* | |
| Insertions/Deletions (1-40 bp) | ≥1% | >99% (97.2%-100%)* | 98.8% (95.3%-99.8%)* | |
| Rearrangements** | ≥1% | >99% (90.8%-100%)* | 98.0% (87.8%-99.9%)* | |
| | <1% | 86.8% (71.1%-95.1%)* | >99% (87.0%-100%)* | |
| Copy Number Amplifications [‡] | ≥20% | 95.3% (82.9%-99.2%)* | 97.6% (85.9%-99.9%)* | |
| REPRODUCIBILITY (average concordance between replicates) | | 96.8% inter-batch precision 100% intra-batch precision | | |

*95% Confidence Interval

** Performance for gene fusions within targeted introns only. Sensitivity for gene fusions occurring outside targeted introns or in highly repetitive intronic sequence contexts is reduced.

⁺ Copy Number Amplifications were calculated using Tumor Fraction.

^{\dagger} Copy-number \geq 8 in genes with at least four targets.

Assay specifications are based on a minimum unique median exon coverage of 5,000x. For cell-free DNA input of \geq 50 ng, the unique median exon coverage is typically 6,000-10,000x.

For additional information specific to the performance of this specimen, please contact Foundation Medicine, Inc. at 1-888-988-3639.

For more comprehensive information please log on to the Interactive Cancer Explorer™ To set up your Interactive Cancer Explorer account, contact your sales representative or call (888) 988-3639.



REFERENCES

- ¹ Ciardiello F, Tortora G (2008) EGFR antagonists in cancer treatment. N Engl J Med 358(11):1160-74.
- ² Lynch TJ, Bell DW, Sordella R, et al. (2004) Activating mutations in the epidermal growth factor receptor underlying responsiveness of non-small-cell lung cancer to gefitinib. N Engl J Med 350(21):2129-39.
- ³ Paez JG, Jänne PA, Lee JC, et al. (2004) EGFR mutations in lung cancer: correlation with clinical response to gefitinib therapy. Science 304(5676):1497-500.
- ⁴ Pao W, Miller V, Zakowski M, et al. (2004) EGF receptor gene mutations are common in lung cancers from "never smokers" and are associated with sensitivity of tumors to gefitinib and erlotinib. Proc Natl Acad Sci USA 101(36):13306-11.
- ⁵ Sequist LV, Waltman BA, Dias-Santagata D, et al. (2011) Genotypic and histological evolution of lung cancers acquiring resistance to EGFR inhibitors. Sci Transl Med 3(75):75ra26.
- ⁶ Wu SG, Liu YN, Tsai MF, et al. (2016) The mechanism of acquired resistance to irreversible EGFR tyrosine kinase inhibitor-afatinib in lung adenocarcinoma patients. Oncotarget ePub Feb 2016.
- ⁷ Ercan D, Zejnullahu K, Yonesaka K, et al. (2010) Amplification of EGFR T790M causes resistance to an irreversible EGFR inhibitor. Oncogene 29(16):2346-56.
- ⁸ Vallee A, Sagan C, Le Loupp AG, et al. (2013) Detection of EGFR gene mutations in non-small cell lung cancer: lessons from a single-institution routine analysis of 1,403 tumor samples. Int J Oncol 43(4):1045-51.
- ⁹ Imielinski M, Berger AH, Hammerman PS, et al. (2012) Mapping the hallmarks of lung adenocarcinoma with massively parallel sequencing. Cell 150(6):1107-20.
- ¹⁰ Hemmings C, Broomfield A, Bean E, et al. (2009) Immunohistochemical expression of EGFR in colorectal carcinoma correlates with high but not low level gene amplification, as demonstrated by CISH. Pathology 41(4):356-60.
- ¹¹ Park S, Choi YL, Sung CO, et al. (2012) High MET copy number and MET overexpression: poor outcome in non-small cell lung cancer patients. Histol Histopathol 27(2):197-207.
- ¹² Oakley GJ, Chiosea SI (2011) Higher dosage of the epidermal growth factor receptor mutant allele in lung adenocarcinoma correlates with younger age, stage IV at presentation, and poorer survival. J Thorac Oncol 6(8):1407-12.
- ¹³ Liang Z, Zhang J, Zeng X, et al. (2010) Relationship between EGFR expression, copy number and mutation in lung adenocarcinomas. BMC Cancer 10:376.
- ¹⁴ Watzka SB, Rauscher-Pötsch I, Nierlich P, et al. (2010) Concordance between epidermal growth factor receptor status in primary non-small-cell lung cancer and metastases: a post-mortem study. Eur J Cardiothorac Surg 38(1):34-7.
- ¹⁵ Marks JL, Broderick S, Zhou Q, et al. (2008) Prognostic and therapeutic implications of EGFR and KRAS mutations in resected lung adenocarcinoma. J Thorac Oncol 3(2):111-6.
- ¹⁶ Izar B, Sequist L, Lee M, et al. (2013) The impact of EGFR mutation status on outcomes in patients with resected stage I non-small cell lung cancers. Ann Thorac Surg 96(3):962-8.
- ¹⁷ Jänne PA, Yang JC, Kim DW, et al. (2015) AZD9291 in EGFR inhibitor-resistant non-small-cell lung cancer. N Engl J Med 372(18):1689-99.
- ¹⁸ Rosell R, Carcereny E, Gervais R, et al. (2012) Erlotinib versus standard chemotherapy as first-line treatment for European patients with advanced EGFR mutation-positive non-small-cell lung cancer (EURTAC): a multicentre, open-label, randomised phase 3 trial. Lancet Oncol 13(3):239-46.



REFERENCES

- ¹⁹ Douillard JY, Ostoros G, Cobo M, et al. (2014) First-line gefitinib in Caucasian EGFR mutation-positive NSCLC patients: a phase-IV, open-label, single-arm study. Br J Cancer 110(1):55-62.
- ²⁰ Sequist LV, Yang JC, Yamamoto N, et al. (2013) Phase III study of afatinib or cisplatin plus pemetrexed in patients with metastatic lung adenocarcinoma with EGFR mutations. J Clin Oncol 31(27):3327-34.
- ²¹ Pirker R, Pereira JR, von Pawel J, et al. (2012) EGFR expression as a predictor of survival for first-line chemotherapy plus cetuximab in patients with advanced non-small-cell lung cancer: analysis of data from the phase 3 FLEX study. Lancet Oncol 13(1):33-42.
- ²² Avizienyte E, Ward RA, Garner AP (2008) Comparison of the EGFR resistance mutation profiles generated by EGFR-targeted tyrosine kinase inhibitors and the impact of drug combinations. Biochem J 415(2):197-206.
- ²³ Gilmer TM, Cable L, Alligood K, et al. (2008) Impact of common epidermal growth factor receptor and HER2 variants on receptor activity and inhibition by lapatinib. Cancer Res 68(2):571-9.
- ²⁴ Landi L, Tiseo M, Chiari R, et al. (2014) Activity of the EGFR-HER2 Dual Inhibitor Afatinib in EGFR-Mutant Lung Cancer Patients With Acquired Resistance to Reversible EGFR Tyrosine Kinase Inhibitors. Clin Lung Cancer 15(6):411-417.e4.
- ²⁵ Kim Y, Ko J, Cui Z, et al. (2012) The EGFR T790M mutation in acquired resistance to an irreversible second-generation EGFR inhibitor. Mol Cancer Ther 11(3):784-91.
- ²⁶ De Grève J, Moran T, Graas MP, et al. (2015) Phase II study of afatinib, an irreversible ErbB family blocker, in demographically and genotypically defined lung adenocarcinoma. Lung Cancer 88(1):63-9.
- ²⁷ Yang JC, Sequist LV, Geater SL, et al. (2015) Clinical activity of afatinib in patients with advanced non-small-cell lung cancer harbouring uncommon EGFR mutations: a combined post-hoc analysis of LUX-Lung 2, LUX-Lung 3, and LUX-Lung 6. Lancet Oncol ePub Jun 2015.
- ²⁸ Janjigian YY, Smit EF, Groen HJ, et al. (2014) Dual Inhibition of EGFR with Afatinib and Cetuximab in Kinase Inhibitor-Resistant EGFR-Mutant Lung Cancer with and without T790M Mutations. Cancer Discov ePub Jul 2014.
- ²⁹ Ribeiro Gomes J, Cruz MR (2015) Combination of afatinib with cetuximab in patients with EGFR-mutant non-small-cell lung cancer resistant to EGFR inhibitors. Onco Targets Ther 8:1137-42.
- ³⁰ Castellanos EH, Rivera G, Wakelee H, et al. (2015) Overcoming Resistance Without the Risk of Reaction: Use of Afatinib and Panitumumab in Two Cases of Epidermal Growth Factor Receptor-Mutated Non-Small Cell Lung Cancer With T790M Mutations. Clin Lung Cancer ePub Mar 2015.
- ³¹ Sequist LV, Soria JC, Goldman JW, et al. (2015) Rociletinib in EGFR-mutated non-small-cell lung cancer. N Engl J Med 372(18):1700-9.
- ³² Ercan D, Xu C, Yanagita M, et al. (2012) Reactivation of ERK signaling causes resistance to EGFR kinase inhibitors. Cancer Discov 2(10):934-47.
- ³³ Eberlein CA, Stetson D, Markovets AA, et al. (2015) Acquired Resistance to the Mutant-Selective EGFR Inhibitor AZD9291 Is Associated with Increased Dependence on RAS Signaling in Preclinical Models. Cancer Res 75(12):2489-500.
- ³⁴ Tricker EM, Xu C, Uddin S, et al. (2015) Combined EGFR/MEK Inhibition Prevents the Emergence of Resistance in EGFR-Mutant Lung Cancer. Cancer Discov 5(9):960-71.
- ³⁵ Thatcher N, Hirsch FR, Luft AV, et al. (2015) Necitumumab plus gemcitabine and cisplatin versus gemcitabine and cisplatin alone as first-line therapy in patients with stage IV squamous non-small-cell lung cancer (SQUIRE): an open-label, randomised, controlled phase 3 trial. Lancet Oncol 16(7):763-74.



REFERENCES

- ³⁶ Paz-Ares L, Mezger J, Ciuleanu TE, et al. (2015) Necitumumab plus pemetrexed and cisplatin as first-line therapy in patients with stage IV non-squamous non-small-cell lung cancer (INSPIRE): an open-label, randomised, controlled phase 3 study. Lancet Oncol 16(3):328-37.
- ³⁷ Shimamura T, Lowell AM, Engelman JA, et al. (2005) Epidermal growth factor receptors harboring kinase domain mutations associate with the heat shock protein 90 chaperone and are destabilized following exposure to geldanamycins. Cancer Res 65(14):6401-8.
- ³⁸ Shimamura T, Li D, Ji H, et al. (2008) Hsp90 inhibition suppresses mutant EGFR-T790M signaling and overcomes kinase inhibitor resistance. Cancer Res 68(14):5827-38.
- ³⁹ Sawai A, Chandarlapaty S, Greulich H, et al. (2008) Inhibition of Hsp90 down-regulates mutant epidermal growth factor receptor (EGFR) expression and sensitizes EGFR mutant tumors to paclitaxel. Cancer Res 68(2):589-96.
- ⁴⁰ Xu W, Soga S, Beebe K, et al. (2007) Sensitivity of epidermal growth factor receptor and ErbB2 exon 20 insertion mutants to Hsp90 inhibition. Br J Cancer 97(6):741-4.
- ⁴¹ Sequist LV, Gettinger S, Senzer NN, et al. (2010) Activity of IPI-504, a novel heat-shock protein 90 inhibitor, in patients with molecularly defined non-small-cell lung cancer. J Clin Oncol 28(33):4953-60.
- ⁴² Socinski MA, Goldman J, El-Hariry I, et al. (2013) A multicenter phase II study of ganetespib monotherapy in patients with genotypically defined advanced non-small cell lung cancer. Clin Cancer Res 19(11):3068-77.
- ⁴³ Isambert N, Delord JP, Soria JC, et al. (2015) Debio0932, a second-generation oral heat shock protein (HSP) inhibitor, in patients with advanced cancer-results of a first-in-man dose-escalation study with a fixed-dose extension phase. Ann Oncol ePub Feb 2015.
- ⁴⁴ Johnson ML, Yu HA, Hart EM, et al. (2015) Phase I/II Study of HSP90 Inhibitor AUY922 and Erlotinib for EGFR-Mutant Lung Cancer With Acquired Resistance to Epidermal Growth Factor Receptor Tyrosine Kinase Inhibitors. J Clin Oncol ePub Apr 2015.
- ⁴⁵ Strong JE, Coffey MC, Tang D, et al. (1998) The molecular basis of viral oncolysis: usurpation of the Ras signaling pathway by reovirus. EMBO J 17(12):3351-62.
- ⁴⁶ Coffey MC, Strong JE, Forsyth PA, et al. (1998) Reovirus therapy of tumors with activated Ras pathway. Science 282(5392):1332-4.
- ⁴⁷ Gong J, Mita MM (2014) Activated ras signaling pathways and reovirus oncolysis: an update on the mechanism of preferential reovirus replication in cancer cells. Front Oncol 4:167.
- ⁴⁸ Villalona-Calero MA, Lam E, Otterson GA, et al. (2015) Oncolytic reovirus in combination with chemotherapy in metastatic or recurrent non-small cell lung cancer patients with KRAS-activated tumors. Cancer ePub Dec 2015.
- ⁴⁹ Brown CJ, Lain S, Verma CS, et al. (2009) Awakening guardian angels: drugging the p53 pathway. Nat Rev Cancer 9(12):862-73.
- ⁵⁰ Kato S, Han SY, Liu W, et al. (2003) Understanding the function-structure and function-mutation relationships of p53 tumor suppressor protein by high-resolution missense mutation analysis. Proc Natl Acad Sci USA 100(14):8424-9.
- ⁵¹ Joerger AC, Fersht AR (2008) Structural biology of the tumor suppressor p53. Annu Rev Biochem 77:557-82.
- ⁵² Kamada R, Nomura T, Anderson CW, et al. (2011) Cancer-associated p53 tetramerization domain mutants: quantitative analysis reveals a low threshold for tumor suppressor inactivation. J Biol Chem 286(1):252-8.
- ⁵³ Kim H, Kim K, Choi J, et al. (2012) p53 requires an intact C-terminal domain for DNA binding and transactivation. J Mol Biol 415(5):843-54.
- ⁵⁴ Bougeard G, Renaux-Petel M, Flaman JM, et al. (2015) Revisiting Li-Fraumeni Syndrome From TP53 Mutation Carriers. J Clin Oncol 33(21):2345-52.



REFERENCES

- ⁵⁵ Sorrell AD, Espenschied CR, Culver JO, et al. (2013) Tumor protein p53 (TP53) testing and Li-Fraumeni syndrome : current status of clinical applications and future directions. Mol Diagn Ther 17(1):31-47.
- ⁵⁶ Nichols KE, Malkin D, Garber JE, et al. (2001) Germ-line p53 mutations predispose to a wide spectrum of early-onset cancers. Cancer Epidemiol Biomarkers Prev 10(2):83-7.
- ⁵⁷ Taubert H, Meye A, Würl P (1998) Soft tissue sarcomas and p53 mutations. Mol Med 4(6):365-72.
- ⁵⁸ Kleihues P, Schäuble B, zur Hausen A, et al. (1997) Tumors associated with p53 germline mutations: a synopsis of 91 families. Am J Pathol 150(1):1-13.
- ⁵⁹ Gonzalez KD, Noltner KA, Buzin CH, et al. (2009) Beyond Li Fraumeni Syndrome: clinical characteristics of families with p53 germline mutations. J Clin Oncol 27(8):1250-6.
- ⁶⁰ Lalloo F, Varley J, Ellis D, et al. (2003) Prediction of pathogenic mutations in patients with early-onset breast cancer by family history. Lancet 361(9363):1101-2.
- ⁶¹ Cancer Genome Atlas Research Network (2014) Comprehensive molecular profiling of lung adenocarcinoma. Nature 511(7511):543-50.
- ⁶² Mogi A, Kuwano H (2011) TP53 mutations in nonsmall cell lung cancer. J Biomed Biotechnol 2011:583929.
- ⁶³ Tekpli X, Landvik NE, Skaug V, et al. (2013) Functional effect of polymorphisms in 15q25 locus on CHRNA5 mRNA, bulky DNA adducts and TP53 mutations. Int J Cancer 132(8):1811-20.
- ⁶⁴ Vignot S, Frampton GM, Soria JC, et al. (2013) Next-generation sequencing reveals high concordance of recurrent somatic alterations between primary tumor and metastases from patients with non-small-cell lung cancer. J Clin Oncol 31(17):2167-72.
- ⁶⁵ Maeng CH, Lee HY, Kim YW, et al. (2013) High-throughput molecular genotyping for small biopsy samples in advanced non-small cell lung cancer patients. Anticancer Res 33(11):5127-33.
- ⁶⁶ Cortot AB, Younes M, Martel-Planche G, et al. (2014) Mutation of TP53 and alteration of p14(arf) expression in EGFR- and KRASmutated lung adenocarcinomas. Clin Lung Cancer 15(2):124-30.
- ⁶⁷ Itakura M, Terashima Y, Shingyoji M, et al. (2013) High CC chemokine receptor 7 expression improves postoperative prognosis of lung adenocarcinoma patients. Br J Cancer 109(5):1100-8.
- ⁶⁸ Seo JS, Ju YS, Lee WC, et al. (2012) The transcriptional landscape and mutational profile of lung adenocarcinoma. Genome Res 22(11):2109-19.
- ⁶⁹ Hirai H, Arai T, Okada M, et al. (2010) MK-1775, a small molecule Wee1 inhibitor, enhances anti-tumor efficacy of various DNAdamaging agents, including 5-fluorouracil. Cancer Biol Ther 9(7):514-22.
- ⁷⁰ Bridges KA, Hirai H, Buser CA, et al. (2011) MK-1775, a novel Wee1 kinase inhibitor, radiosensitizes p53-defective human tumor cells. Clin Cancer Res 17(17):5638-48.
- ⁷¹ Rajeshkumar NV, De Oliveira E, Ottenhof N, et al. (2011) MK-1775, a potent Wee1 inhibitor, synergizes with gemcitabine to achieve tumor regressions, selectively in p53-deficient pancreatic cancer xenografts. Clin Cancer Res 17(9):2799-806.
- ⁷² Osman AA, Monroe MM, Ortega Alves MV, et al. (2015) Wee-1 kinase inhibition overcomes cisplatin resistance associated with high-risk TP53 mutations in head and neck cancer through mitotic arrest followed by senescence. Mol Cancer Ther 14(2):608-19.
- ⁷³ Lehmann S, Bykov VJ, Ali D, et al. (2012) Targeting p53 in vivo: a first-in-human study with p53-targeting compound APR-246 in refractory hematologic malignancies and prostate cancer. J Clin Oncol 30(29):3633-9.



REFERENCES

- ⁷⁴ Xu L, Huang CC, Huang W, et al. (2002) Systemic tumor-targeted gene delivery by anti-transferrin receptor scFvimmunoliposomes. Mol Cancer Ther 1(5):337-46.
- ⁷⁵ Xu L, Tang WH, Huang CC, et al. (2001) Systemic p53 gene therapy of cancer with immunolipoplexes targeted by anti-transferrin receptor scFv. Mol Med 7(10):723-34.
- ⁷⁶ Camp ER, Wang C, Little EC, et al. (2013) Transferrin receptor targeting nanomedicine delivering wild-type p53 gene sensitizes pancreatic cancer to gemcitabine therapy. Cancer Gene Ther 20(4):222-8.
- ⁷⁷ Kim SS, Rait A, Kim E, et al. (2015) A tumor-targeting p53 nanodelivery system limits chemoresistance to temozolomide prolonging survival in a mouse model of glioblastoma multiforme. Nanomedicine 11(2):301-11.
- ⁷⁸ Senzer N, Nemunaitis J, Nemunaitis D, et al. (2013) Phase I study of a systemically delivered p53 nanoparticle in advanced solid tumors. Mol Ther 21(5):1096-103.
- ⁷⁹ Ma CX, Cai S, Li S, et al. (2012) Targeting Chk1 in p53-deficient triple-negative breast cancer is therapeutically beneficial in human-in-mouse tumor models. J Clin Invest 122(4):1541-52.
- ⁸⁰ Cross DA, Ashton SE, Ghiorghiu S, et al. (2014) AZD9291, an Irreversible EGFR TKI, Overcomes T790M-Mediated Resistance to EGFR Inhibitors in Lung Cancer. Cancer Discov 4(9):1046-1061.
- ⁸¹ Shepherd FA, Rodrigues Pereira J, Ciuleanu T, et al. (2005) Erlotinib in previously treated non-small-cell lung cancer. N Engl J Med 353(2):123-32.
- ⁸² Han JY, Park K, Kim SW, et al. (2012) First-SIGNAL: first-line single-agent iressa versus gemcitabine and cisplatin trial in neversmokers with adenocarcinoma of the lung. J Clin Oncol 30(10):1122-8.
- ⁸³ Maemondo M, Inoue A, Kobayashi K, et al. (2010) Gefitinib or chemotherapy for non-small-cell lung cancer with mutated EGFR. N Engl J Med 362(25):2380-8.
- ⁸⁴ Mitsudomi T, Morita S, Yatabe Y, et al. (2010) Gefitinib versus cisplatin plus docetaxel in patients with non-small-cell lung cancer harbouring mutations of the epidermal growth factor receptor (WJTOG3405): an open label, randomised phase 3 trial. Lancet Oncol 11(2):121-8.
- ⁸⁵ Mok TS, Wu YL, Thongprasert S, et al. (2009) Gefitinib or carboplatin-paclitaxel in pulmonary adenocarcinoma. N Engl J Med 361(10):947-57.
- ⁸⁶ Qi WX, Fu S, Zhang Q, et al. (2015) Anti-epidermal-growth-factor-receptor agents and complete responses in the treatment of advanced non-small-cell lung cancer: a meta-analysis of 17 phase III randomized controlled trials. Curr Med Res Opin 31(1):25-33.
- ⁸⁷ Zhao H, Fan Y, Ma S, et al. (2015) Final overall survival results from a phase III, randomized, placebo-controlled, parallel-group study of gefitinib versus placebo as maintenance therapy in patients with locally advanced or metastatic non-small-cell lung cancer (INFORM; C-TONG 0804). J Thorac Oncol 10(4):655-64.
- ⁸⁸ Fukuoka M, Wu YL, Thongprasert S, et al. (2011) Biomarker analyses and final overall survival results from a phase III, randomized, open-label, first-line study of gefitinib versus carboplatin/paclitaxel in clinically selected patients with advanced non-small-cell lung cancer in Asia (IPASS). J Clin Oncol 29(21):2866-74.
- ⁸⁹ Ramalingam SS, Kotsakis A, Tarhini AA, et al. (2013) A multicenter phase II study of cetuximab in combination with chest radiotherapy and consolidation chemotherapy in patients with stage III non-small cell lung cancer. Lung Cancer 81(3):416-21.
- ⁹⁰ Kim ES, Neubauer M, Cohn A, et al. (2013) Docetaxel or pemetrexed with or without cetuximab in recurrent or progressive nonsmall-cell lung cancer after platinum-based therapy: a phase 3, open-label, randomised trial. Lancet Oncol 14(13):1326-36.



REFERENCES

- ⁹¹ Wheler JJ, Tsimberidou AM, Falchook GS, et al. (2013) Combining erlotinib and cetuximab is associated with activity in patients with non-small cell lung cancer (including squamous cell carcinomas) and wild-type EGFR or resistant mutations. Mol Cancer Ther 12(10):2167-75.
- ⁹² Tsigelny IF, Wheler JJ, Greenberg JP, et al. (2015) Molecular determinants of drug-specific sensitivity for epidermal growth factor receptor (EGFR) exon 19 and 20 mutants in non-small cell lung cancer. Oncotarget 6(8):6029-39.
- ⁹³ Crawford J, Swanson P, Schwarzenberger P, et al. (2013) A phase 2 randomized trial of paclitaxel and carboplatin with or without panitumumab for first-line treatment of advanced non-small-cell lung cancer. J Thorac Oncol 8(12):1510-8.
- ⁹⁴ Schuette W, Behringer D, Stoehlmacher J, et al. (2015) CHAMP: A Phase II Study of Panitumumab With Pemetrexed and Cisplatin Versus Pemetrexed and Cisplatin in the Treatment of Patients With Advanced-Stage Primary Nonsquamous Non-Small-Cell Lung Cancer With Particular Regard to the KRAS Status. Clin Lung Cancer ePub Jun 2015.



ABOUT FOUNDATIONACT™

FoundationACTTM: FoundationACT was developed and its performance characteristics determined by Foundation Medicine, Inc. (Foundation Medicine). FoundationACT has not been cleared or approved by the United States Food and Drug Administration (FDA). The FDA has determined that such clearance or approval is not necessary. FoundationACT may be used for clinical purposes and should not be regarded as purely investigational or for research only. Foundation Medicine's clinical reference laboratory is certified under the Clinical Laboratory Improvement Amendments of 1988 (CLIA) as qualified to perform high-complexity clinical testing.

Diagnostic Significance: FoundationACT identifies alterations to select cancer-associated genes or portions of genes (biomarkers).

Qualified Alteration Calls (equivocal): All equivocal calls, regardless of alteration type, imply that there is adequate evidence to call the alteration with confidence. However, the repeatability of equivocal calls may be lower than non-equivocal calls. The threshold used in FoundationACT for identifying a copy number amplification is five (5) for ERBB2 and six (6) for all other genes. For copy number amplifications, the equivocal status may be applied to calls in samples with calculated tumor fraction <30% but above the noise threshold. In addition, copy number amplifications in genes with three (3) baited exons are also marked as equivocal. For substitutions, the equivocal status is applied to calls with allele frequency between 0.1% and 0.5%.

The Report incorporates analyses of peer-reviewed studies and other publicly available information identified by Foundation Medicine; these analyses and information may include associations between a molecular alteration (or lack of alteration) and one or more drugs with potential clinical benefit (or potential lack of clinical benefit), including drug candidates that are being studied in clinical research.

NOTE: A finding of biomarker alteration does not necessarily indicate pharmacologic effectiveness (or lack thereof) of any drug or treatment regimen; a finding of no biomarker alteration does not necessarily indicate lack of pharmacologic effectiveness (or effectiveness) of any drug or treatment regimen.

Alterations and Drugs Not Presented in Ranked Order: In this Report, neither any biomarker alteration, nor any drug associated with potential clinical benefit (or potential lack of clinical benefit), are ranked in order of potential or predicted efficacy.

Level of Evidence Not Provided: Drugs with potential clinical benefit (or potential lack of clinical benefit) are not evaluated for source or level of published evidence.

No Guarantee of Clinical Benefit: This Report makes no promises or guarantees that a particular drug will be effective in the treatment of disease in any patient. This Report also makes no promises or guarantees that a drug with potential lack of clinical benefit will in fact provide no clinical benefit.

No Guarantee of Reimbursement: Foundation Medicine makes no promises or guarantees that a healthcare provider, insurer or other third party payor, whether private or governmental, will reimburse a patient for the cost of FoundationACT.

Treatment Decisions are Responsibility of Physician: Drugs referenced in this Report may not be suitable for a particular patient. The selection of any, all or none of the drugs associated with potential clinical benefit (or potential lack of clinical benefit) resides entirely within the discretion of the treating physician. Indeed, the information in this Report must be considered in conjunction with all other relevant information regarding a particular patient, before the patient's treating physician recommends a course of treatment.

Decisions on patient care and treatment must be based on the independent medical judgment of the treating physician, taking into consideration all applicable information concerning the patient's condition, such as patient and family history, physical examinations, information from other diagnostic tests, and patient preferences, in accordance with the standard of care in a given community. A treating physician's decisions should not be based on a single test, such as this Test, or the information contained in this Report.

Certain sample of variant characteristics may result in reduced sensitivity. These include: low sample quality, deletions and insertions >40bp, or repetitive/high homology sequences. FoundationACT is performed using cell-free DNA, and as such germline events may not be reported. The following target typically has low coverage resulting in a reduction in sensitivity: *TP53* exon 1 and *PDGFRA* exon 12.