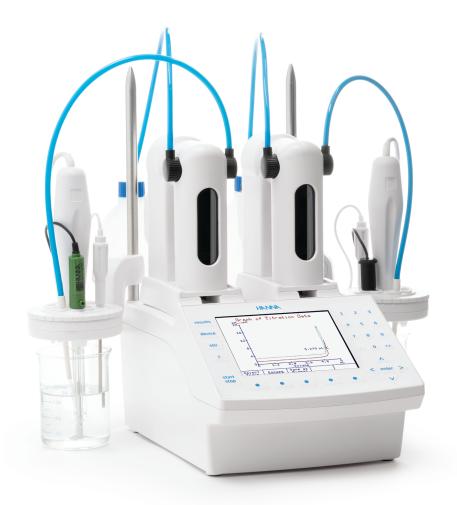


AUTOMATIC POTENTIOMETRIC TITRATOR





HI932 AUTOMATIC POTENTIOMETRIC TITRATOR

Congratulations on choosing your new Hanna titrator. It is a powerful and versatile instrument capable of accurate and fast analysis of a wide range of samples. In this manual, you'll find:

QUICK START GUIDE

This guide will help you quickly setup, operate, and introduce you to your new titrator. It covers basic connections, user interface, how to perform calibrations, and how to run a titration.

INSTRUCTION MANUAL

The manual provides a comprehensive description of the operating principles user interface, general options, methods, titration/ direct reading mode, pH, mV and ISE mode, maintenance, etc.

APPLICATIONS BROCHURE

This brochure contains complete instructions for commonly-used analyses. Additional methods and method packs are available; contact your local Hanna office for more details.

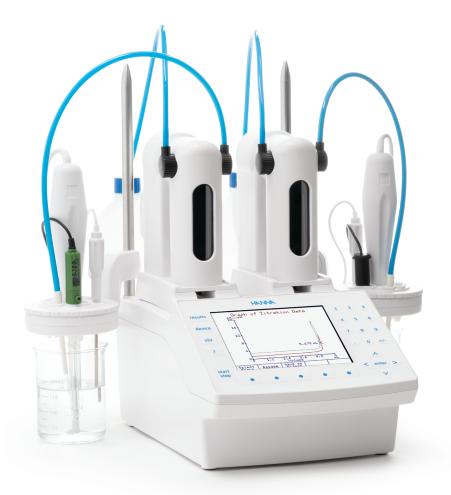
TITRATION THEORY

This guide outlines the principles of operation of the titrator. It covers the chemistry of titrations, titration types, and result calculations.

If you need additional technical information, do not hesitate to e-mail us at tech@hannainst.com or view our worldwide contact list for a Hanna Instruments representative near you at www.hannainst.com.

HI932

AUTOMATIC POTENTIOMETRIC TITRATOR





Dear

Congratulations on choosing a Hanna Instruments product.

Customer, Please read this Quick Start Guide carefully before using the instrument. This guide will provide you with the necessary information for the correct use of the instrument.

The purpose of this guide is to present a quick overview of setting up and using the instrument.

For detailed information illustrating the extensive capabilities of your titrator, please refer to the Instruction Manual.

INTRODUCTION	4
SAFETY MEASURES	4
TITRATOR CONNECTIONS	5
USER INTERFACE	6
HOW TO SELECT YOUR LANGUAGE	7
HOW TO USE THE CONTEXTUAL HELP	7
METHODS	7
HOW TO CALIBRATE A pH ELECTRODE	7
HOW TO PERFORM A TITRATION	

QUICK START GUIDE

INTRODUCTION

The H1932 automatic titrator is designed to perform a wide variety of potentiometric titrations with high accuracy, flexibility and reproducibility, allowing the user to obtain both accurate results and high-speed analysis.

The titrator can perform fixed endpoint or equivalence point titrations and direct measurements by measuring the pH/mV and temperature of the sample.

Reports and methods can be transferred to a PC via a USB interface, saved to a USB storage device or printed directly from the titrator. An external keyboard can also be attached for added convenience.

The H1922 Autosampler can be connected for sample automation.

How can I find certain information?

- The Quick Start Guide will help the user learn how to operate the titrator within a short period of time.
- The Instruction Manual provides a complete description of the operating principles (user interface, general options, methods, titration/direct reading mode, pH, mV and ISE mode, maintenance, etc.).
- The Titration Theory outlines the basic concepts of titration.
- The contextual **Help** screens contain detailed explanations of every screen.

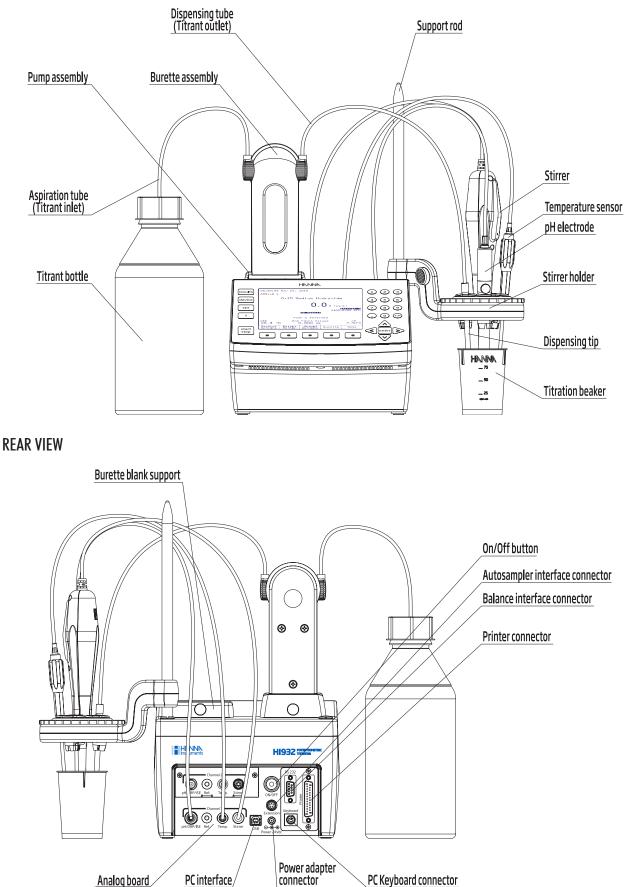
SAFETY MEASURES

The following safety measures must be followed:

- 1. Never connect or disconnect the pump assembly or other peripheral with the titrator turned on.
- 2. Verify that the burette and the attached tubing are assembled correctly.
- 3. Always check that the titrant bottle and the titration beaker are placed on a flat, stable surface.
- 4. Always wipe up spills and splashes immediately.
- 5. Avoid the following environmental working conditions:
 - Severe vibrations
 - Direct sunlight
 - Atmospheric relative humidity above 95% non-condensing
 - Environment temperatures below 10°C and above 40°C
 - Explosion hazards
- 6. Have the titrator serviced by qualified service personnel only.

TITRATOR CONNECTIONS

FRONT VIEW

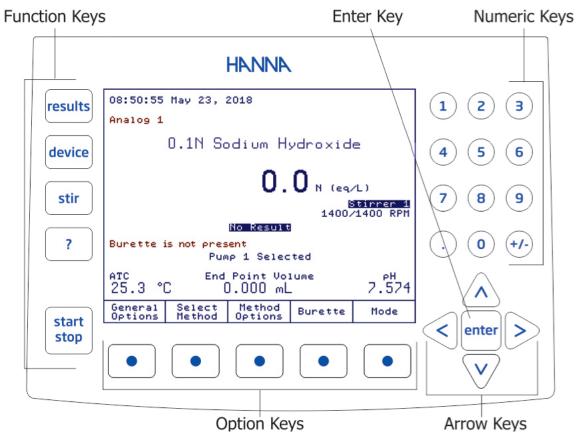


USER INTERFACE

Keypad

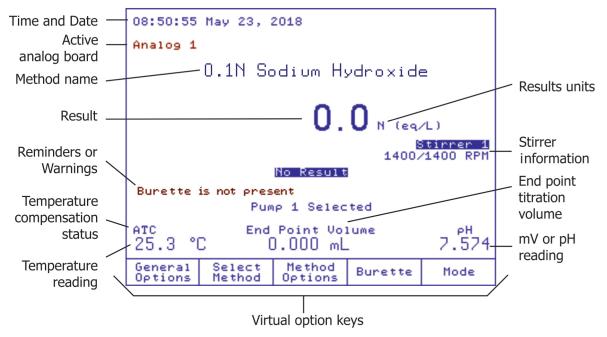
The titrator's keypad has 27 keys grouped in five categories, as follows:

Display



The user interface contains several screens. In each screen, many information fields are present at the same time. The information is displayed in an easy-to-read manner.

Virtual option keys describe the function performed when the corresponding option key is pressed.



HOW TO SELECT YOUR LANGUAGE

To change the language, press $G_{Options}^{General}$ from the main screen. Highlight the *Language* option. Using the Λ and ∇ keys, select the language and press S_{elect} .

Restart the titrator in order to apply the new language setting.

General Options	
Select the option to be modified.	
Save to USB Restore from USB Administration: Disabled Temperature: °C, ATC Date and Time Setting Display Settings Beeper: Off Stirrer: Enabled Language: English	
Titrant 1 Volume Alert: Titrant 1 Age Reminder: Titrant 2 Volume Alert: Titrant 2 Age Reminder: Espanol	
Select Escape	

HOW TO USE THE CONTEXTUAL HELP

Information about the titrator can be easily accessed by pressing (?). The contextual help can be accessed at any time and it provides useful information about the current screen.

METHODS

The H1932 titrator can store up to 100 methods (standard and user) and 30 autosampler sequences.

Standard Methods

Each titrator is supplied with a package of standard methods. Standard method packs are developed at Hanna Instruments to meet analysis requirements of specific industries (e.g., water treatment, wine, dairy, etc.).

User-Defined Methods

User defined methods allow the user to create and save their own methods. Each new method is based on an existing method which is altered to suit a specific application.

HOW TO CALIBRATE A pH ELECTRODE

To enter pH calibration mode, press Mode, then PH, then Calibration PH, then Calibration PH.

PREPARATION

Pour small quantities pH 4.01, pH 7.01 and pH 10.01 buffer solutions into clean beakers. If possible, use plastic beakers to minimize any EMC interferences.

For accurate calibration and to minimize cross-contamination, use two beakers for each buffer solution: one for rinsing the electrode and one for calibration.

CALIBRATION PROCEDURE

- If the instrument has been previously calibrated and calibration was not cleared, the old calibration can be cleared by pressing
- **Note:** It is very important to clear calibration history when a new electrode is used. Most errors and warning messages that appear during calibration depend on calibration history.

- **QUICK START GUIDE**
- Use the Next Buffer or Buffer to select pH 4.01 buffer solution.
- Use the second beaker of pH 4.01 buffer solution to rinse the pH electrode, temperature probe and propeller stirrer.
- Immerse the pH electrode, temperature probe and propeller stirrer in the pH 4.01 buffer solution. The pH electrode's bulb must be completely immersed in the buffer solution and the reference junction needs to be 5-6 mm below the surface. Add additional buffer if necessary.
- Press stir to turn on the propeller stirrer.
- Once the reading has stabilized, press Accept to update the calibration.
- Repeat this procedure for pH 7.01 and 10.01 buffer solutions.
- Press Escape to accept and exit pH calibration mode.

HOW TO PERFORM A TITRATION

Required Solutions

- Titrant 500 mL of 0.1 M (mol/L) Sodium Hydroxide (NaOH) in a titrant bottle.
- Sample 0.1 mol/L Hydrochloric Acid (HCl).
- Distilled or deionized water.

Note: Analytical grade reagents and water should be used for accurate results.

Priming the Burette

- Insert the aspiration tube in the titrant bottle and the dispensing tube in a waste beaker.
- From the main screen press
 - Highlight the *Prime Burette* option and then press Select
 - Enter the number of burette rinses. At least 3 rinses are recommended.
 - Press Accept to start.
 - The message "Executing..." will be displayed.

Note: Make sure you have continuous liquid flow inside the burette. For accurate results, the aspiration tube, the dispensing tube and the syringe must be free of air bubbles.

Method Selection

For this analysis, we will use the H11009 Neutralization w/ NaOH.

To select this method:

- Press Select Method
 Use the A and V keys to highlight H11009 Neutralization w/ NaOH.
- Press Select

Setting Method Parameters

To display the method parameters, press Method Options

The View/Modify Method screen will be displayed.

Only certain parameters can be changed.

For this titration, the NaOH titrant concentration and the size of the HCI sample need to be entered.

To accomplish this:

- Highlight *Titrant Conc.* option, then press Select. The **Titrant Concentration** screen will be displayed.
- Enter the correct value, then press Accept
- Highlight *Analyte Size* option, then press Select
- Enter the volume of the sample (e.g.: 5 mL), then press Accept
- Press Escape, highlight Save Method option and then press Select

	Titi	rant1 Co	onc.			Sar	nele Vol	ume	
Enter the titrant 1 concentration.			Enter millil		al sample	volume i	n		
		0.106	2 <mark>6 M</mark> (mol	7L)			1.0000	mL	
						olume wil size is	l be used selected.	when fix	ed
Accept	Escape	Delete Digit		Exponent	Accept	Escape	Delete Digit		Exponent

Setup Titration Report

Users can select the information that is stored for each titration.

To setup the titration report, follow the procedure below:

- From the main screen, press results. The Data Parameters screen will be displayed.
- Highlight Setup Titration Report and press Select
- Mark the fields to be included in the titration report with the "*" symbol. Use the 🛆 and 👽 keys to highlight a field and select / Unselect to toggle the field.
- Press save the customized report .

Preparing the Sample

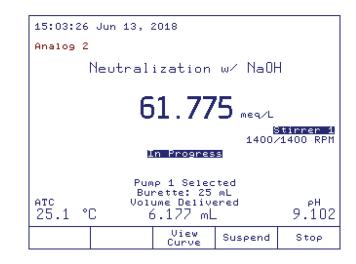
- Add 50 to 65 mL of distilled/deionized water to the titration beaker.
- Use a pipette or burette to add 5.0 mL of the sample (0.1M Hydrochloric Acid (HCl)) into the same beaker.
- Slide the stirrer assembly up.
- Place the beaker under the stirrer assembly.
- Lower the stirrer assembly until the electrodes are submersed and the stirrer is close to the bottom of the beaker.
- Adjust the level of the sample solution with distilled/deionized water so that the pH electrode bulb is completely immersed in the sample solution and the reference junction of the electrode is 5-6 mm below the surface.

Performing a Titration

- From the main screen, press stop. You will be prompted to enter the analyte size. Enter 5 mL and press enter. The titrator will start the analysis.
- At the end of the titration, the message "Titration Completed" will appear on the display with the final concentration of the analyte in the sample and the equivalence endpoint volume.

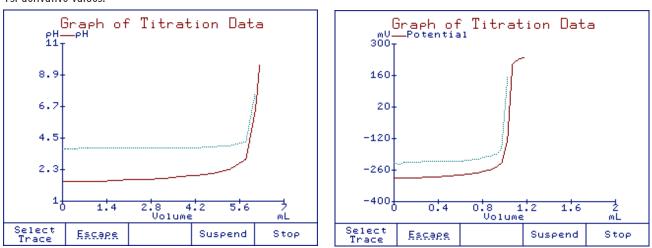
Understanding the Displayed Information

During a titration the following screen is displayed:



Viewing Graph During Titration

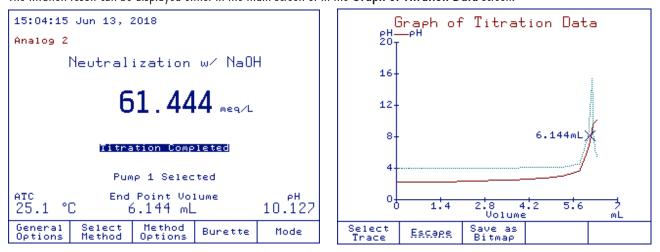
After a few doses are dispensed, View Curve will become active. Press View Curve to display the real-time titration graph. The curves displayed are plots of the pH and the 1st derivative versus Titrant Volume (for details, see the Instruction Manual). The two graphs are scaled to fit in the same screen window. Press Select Trace to change the y-axis scale to either the pH values or the 1st derivative values.



Titration Termination

The titration is normally terminated when the first equivalence endpoint is detected according to the selected algorithm. To ensure the correct detection and interpolation of the equivalence endpoint, the titrator will dispense a few additional doses after the endpoint was reached.

The titration result can be displayed either in the main screen or in the Graph of Titration Data screen:



When the titration has ended, the titrator will display the equivalence endpoint volume and the final concentration of the analyte together with the "Titration Completed" message.

To view the titration graph and/or results, press results.

When the titration ends, an "x" will mark the endpoint on the pH versus titrant volume curve in the **Graph of Titration Data** screen. The value of the endpoint volume is also displayed next to the endpoint.

Results

The results obtained from a titration are stored in a report file that can be viewed, transferred to a USB Storage Device or PC and printed.

Viewing the last titration data

- From the main screen, press results. The Data Parameters screen will be displayed.
- From the **Data Parameters** screen highlight the *Review Last Analysis Report* option and press Select. The **Review Result** screen will be displayed.
- Use the Page Up and Page Down keys to display information related to the last titration performed.

See Titration Report on next page.

Printing the titration report

Connect a DOS / Windows-compatible parallel printer directly to the DB 25-pin connector located on the back of the titrator.

Note: When connecting the printer, please turn off the titrator and the printer.

Printing out the report:

- From the **Review Report** screen, press
- During the information transfer to the printer, the message "Printing" will be displayed on the screen.
- Press Escape to return to the Data Parameters screen.
- Press Escape again to return to the main screen.

Saving data to USB Storage Device

This feature allows saving the results of titrations or pH / mV / ISE logging sessions on a USB storage device.

- From the main screen, press General Options screen will be displayed.
- Highlight the Save Files to USB Storage Device option using the \bigwedge and \bigvee keys.
- Insert the USB storage device into the USB socket.
- Press Select , the List of Files on Titrator screen will be displayed.

Li	ist of F	iles on	Titrat	or		
Use <-/-> arrow keys to select file type 2 tray report files						
	WRAY0002.RPT TRAY0001.RPT					
Escape	Copy file	Сору А11	Delete File	Delete All		

- Use the \lt and \gt keys to select the report files.
- Press Copy All to transfer all available reports to the USB storage device, or highlight the name of the report file to be transferred and press Copy File.

• Transferring a report file will automatically transfer the corresponding log file and titration graph (*.BMP file if applicable).

- Press Escape to return to the General Options screen.
- Press Escape again to return to the main screen.

Titration report

While scrolling with the $\begin{bmatrix} Page \\ Up \end{bmatrix}$ and $\begin{bmatrix} Page \\ Down \end{bmatrix}$ keys, the fields below can be seen on the titrator display or printed. The same information is available on the saved report file (Ti_00007.rpt in this example).

HI932 - Titration Report

Method Name: New	
Time & Date:	15:01 Jun 13, 2018
Report ID:	Ti_00011
Calibra	ation Data
	al Efficiency Temp. and Date
4.010pH 169.3mV	
7.010pH -5.8mV	in 13, 2018 98.7% 23.9°C A
11:42 Ju 10.010pн -180.7mV	n 13, 2018 98.7% 24.0°C A
	in 13, 2018
GLP & Mete	er Information
Sample Name:	
Company Name:	
Operator Name:	
Electrode Name: Field 1:	
Field 2:	
Field 3:	
Titrator Software Ve	ersion: v1.00
Base Board Software	Version: v1.00
Pump 1 Software Vers	sion: v1.00
Pump 2 Software Vers	sion: v1.00
Stirrer 1 Software V	
Titrator Serial Numb	per: TT180525011
Analog Board1 Serial	
Analog Board2 Serial	
Pump 1 Serial Number	DP180525004
Pump 2 Serial Number	
Stirrer 1 Serial Nur	
Analog 1 Calibration	
Analog 2 Calibration	1 Date: May 25, 2018

Meth	nod Parameters
Name:	Neutralization w/ NaOH
Method Revision:	3.0
Analysis Type:	Standard Titration

Analog Board:	Analog	2
Stirrer Configuration:		
Stirrer:	Stirrer	1
Stirring Speed:	1400 R	PM
Pump Configuration:		
Titrant pump:	Pump	1
Reagent Addition 1:	Disabl	
Reagent Addition 2:	Disabl	ed
Dosing Type:	Dynam	ic
Min Vol:	0.050	
Max Vol:	0.500	
delta E:	20.000	
End Point Mode: pH 1		
Recognition Options	11g point,100 p	
Threshold:	50 mV/1	mT.
Range:		NO
Filtered Derivative		NO
Pre-Titration Volume:	0.000 1	
Pre-Titration Stir Time		
Measurement Mode:		
delta E:	1.0	-
delta t: Min wait:	2 s	
	2 s	
Max wait:	15 s	
Electrode Type:		pH
Blank Option:	No Bla	
Calculations: Sample	=	
Dilution Option:	Disabl	
Titrant Name:	0.1N Ha	
Titrant Conc.:	0.1000 N (eq/	
Analyte Size:	10.0000	
Analyte Entry:	Fix	
Maximum Titrant Volume:		
Potential Range: -200		
Volume/Flow Rate: 25		
Signal Averaging:	1 Readi	ng
Significant Figures:	XXX	XX
N (eq/L)> meq/L		
V eq 1000meq		
_**		
L eq		
mL L		
*		
1000mL		
V = volume dispensed in	n liters.	
0.100 eq/L \rightarrow titrant of	conc.	
10.000 mL -> sample vol	Lume	
Nr Volume[mL] mV		hic Temp.[°C] Time
0 0.000 274.4		0.0 24.9 A 00:00:00
1 0.050 274.4		1.0 25.0 A 00:00:07
2 0.100 274.4		0.0 25.0 A 00:00:10
2 0 200 274 2	2 2 2 2 2	0 0 2 0 7 0 7 00.00.12

3

4

5

6

7

8

0.200

0.400

0.800

1.300

1.800

2.300

274.3

274.0

273.2

271.5

269.5

267.2

2.222

2.227

2.241

2.271

2.304

2.344

25.1 A 00:00:30

-4.7 25.1 A 00:00:37

A 00:00:12 A 00:00:15

A 00:00:18

A 00:00:24

25.0

25.0

25.0

25.0

-0.8

-1.6

-2.0

-3.4

-3.9

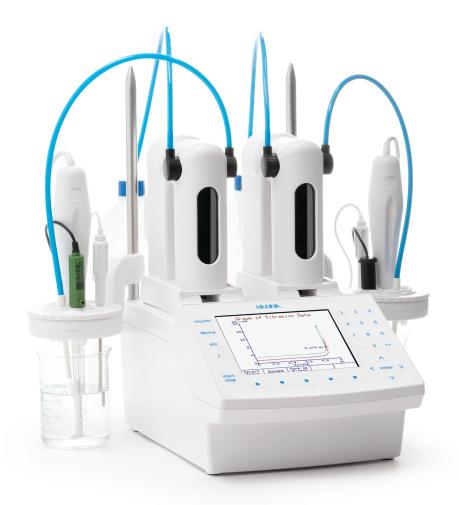
9 10 11	2.800 3.300 3.800	264.4 260.8 256.1	2.393 2.455 2.535	-5.7 -7.2 -9.3	25.1 25.1 25.1	A 00:00:43 A 00:00:50 A 00:00:58
12	4.300	250.3	2.635	-11.7	25.1	A 00:01:05
13	4.800	241.9	2.779	-16.8	25.1	A 00:01:14
14	5.300	228.3	3.011	-27.2	25.1	A 00:01:23
15	5.800	193.0	3.614	-70.5	25.1	A 00:01:31
16	6.077	21.0	6.556	-620.0	25.1	A 00:01:48
17	6.128	-38.2	7.568	-1183.2	25.1	A 00:02:03
18	6.177	-123.6	9.031	-1708.0	25.1	A 00:02:19
19	6.227	-157.7	9.616	-682.8	25.1	A 00:02:28
20	6.278	-174.5	9.903	-335.8	25.1	A 00:02:35
21	6.339	-187.8	10.130	-215.9	25.1	A 00:02:42

Titration ResultsMethod Name:Neutralization w/ NaOHTime & Date:15:01 Jun 13, 2018Analyte Size:10.0000 mLEnd Point Volume:6.144 mLpH Equivalence Point:8.063Result:61.444 meq/LInitial & Final pH:2.219 to 10.130Titration Duration:2:42 [mm:ss]Titration went to Completion

Analyst Signature: ____

HI932

AUTOMATIC POTENTIOMETRIC TITRATOR





Dear | Thank you for choosing a Hanna Instruments product.

Customer, Please read this instruction manual carefully before using this instrument. This manual will provide you with the necessary information for the correct use of this instrument, as well as a precise idea of its versatility.

If you need additional technical information, do not hesitate to e-mail us at tech@hannainst.com or view our worldwide contact list for a Hanna Instruments representative near you at www.hannainst.com.

CHAPTER 1. INTRODUCTION CHAPTER 2. SETUP CHAPTER 3. USER INTERFACE CHAPTER 4. GENERAL OPTIONS CHAPTER 5. TITRATION METHODS CHAPTER 6. TITRATION / DIRECT READING MODE CHAPTER 7. pH MODE CHAPTER 8. mV MODE CHAPTER 9. ISE MODE CHAPTER 10. AUXILIARY FUNCTIONS CHAPTER 11. MAINTENANCE, PERIPHERALS CHAPTER 12. AUTOSAMPLER APPENDIX 1. TECHNICAL SPECIFICATIONS APPENDIX 2. ACCESSORIES CERTIFICATION **RECOMMENDATIONS FOR USERS** WARRANTY

INTRODUCTION

1. INTRODUCTION

H1932 is an automatic potentiometric titrator with high accuracy, great flexibility and repeatability.

The titrator is designed to perform a variety of potentiometric titrations, allowing the user to obtain both good results and high-speed analysis.

The main attributes of the HI932 titrator are:

- Small footprint, requires minimal bench space
- Casing made with strong, chemically resistant plastic
- Flexible electrode holder supports up to 3 electrodes, 4 dispensing tubes, 1 temperature sensor and removable stirrer
- Electrode holder positions electrodes in the center of beaker, allowing for smaller sample sizes
- Integrated Peristaltic Pump available for reagent addition
- Support for 100 titration methods and 30 autosampler sequences
- User-customizable reports
- Integrated research grade pH/mV/ISE meter
- Clearly displayed warning and error messages

This manual provides information regarding installation and functionality of the titrator and refined operation suggestions. Before using the titrator, it is recommended you become familiar with its various features and functionality.

CHAPTER 2. SETUP

2.1. UNPACKING	2-3
2.2. SAFETY MEASURES	2-4
2.3. INSTALLATION	2-4
2.3.1. TITRATOR FRONT VIEW	2-4
2.3.2. TITRATOR REAR VIEW	2-5
2.3.3. TITRATOR REAR VIEW WITH PERISTALTIC PUMP	2-5
2.3.4. TITRATOR RIGHT-SIDE VIEW	2-6
2.3.5. TITRATOR ASSEMBLY	2-6
2.3.5.1. ASSEMBLING STIRRER AND ELECTRODES HOLDER	2-6
2.3.5.2. ATTACHING STIRRER AND ELECTRODES	2-7
2.3.5.3. CONNECTING THE PUMP	2-8
2.3.5.4. ATTACHING BURETTE BLANK SUPPORT	2-8
2.3.5.5. ATTACHING THE BURETTE	2-9
2.3.5.6. CONNECTING PERISTALTIC PUMP TUBING	2-10
2.3.5.7. ELECTRICAL CONNECTIONS	2-11

2.1. UNPACKING

ITEM

Remove the titrator and accessories from the packaging and examine it carefully to make sure that no damage has occurred during shipping. Notify your nearest Hanna Service Center if damage is observed. Each H1932 potentiometric titrator is supplied with:

SETUP

QUANTITY

Titrator	1 рс
Pump Assembly	
Burette Assembly	
Burette (with 25 mL syringe)	
Aspiration Tube with Fitting and Protection Tube	
 Dispensing Tube with Normal Dispensing Tip, Fitting, Protection Tube and Tube Guide 	
Tube Locks	
Tool for Burette Cap Removal	
Light Spectrum Protection Screen	
Electrodes Holder and Stirrer	1 рс
Stirrer Holder	
Overhead Stirrer	
Propellers (3 pcs)	
Support Rod	
Burette Blank Support	1 рс
Pump and Burette Locking Screws with Plastic Head	2 pcs
Temperature Sensor	1 рс
Shorting Cap	1 рс
Power Adapter	1 рс
USB Cable	1 рс
Instruction Manual	1 рс
USB Memory Stick	1 рс
HI900 PC Application (Installation Kit on USB Stick)	1 рс
Quality Certificate	1 рс

See **APPENDIX 2**, titrator components section for pictures.

If any of the items are missing or damaged, please contact your sales representative.

Note: Save all packing materials until you are sure that the instrument functions correctly. Any damaged or defective items must be returned in their original packing materials together with the supplied accessories.

SETUP

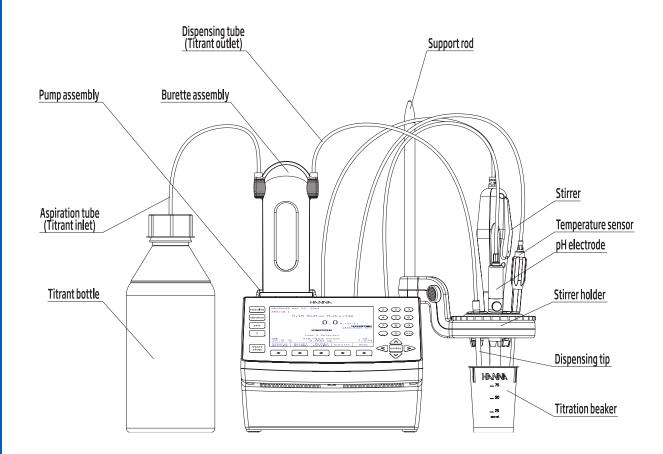
2.2. SAFETY MEASURES

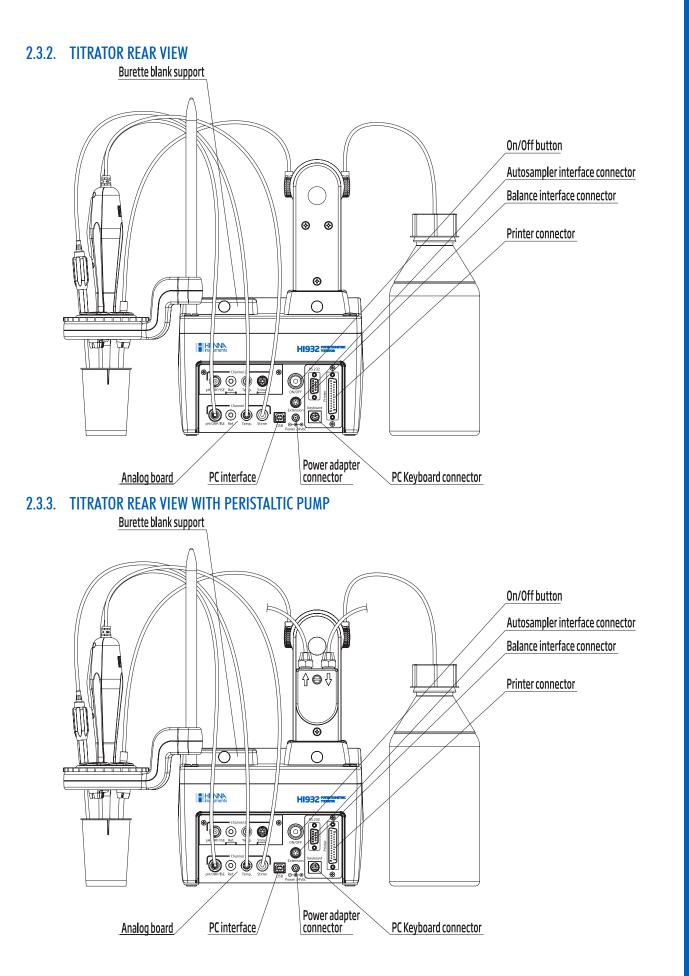
The following safety measures must be followed:

- 1. Never connect or disconnect the pump assembly with the titrator turned on.
- 2. Verify that the burette and the attached tubing are assembled correctly (see Maintenance, Peripherals, Burette Maintenance for more details).
- 3. Always check that the titrant bottle and the titration beaker are on a flat surface.
- 4. Always wipe up spills and splashes immediately.
- 5. Avoid the following environmental working conditions:
 - Severe vibrations
 - Direct sunlight
 - Atmospheric relative humidity above 95% non-condensing
 - Environment temperatures below 10°C and above 40°C
 - Explosion hazards
- 6. Have the titrator serviced only by qualified service personnel.

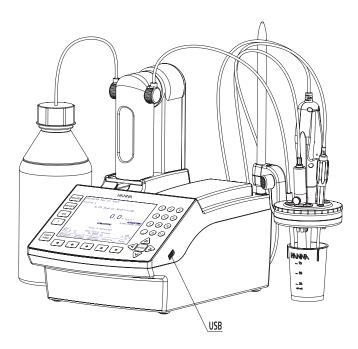
2.3. INSTALLATION

2.3.1. TITRATOR FRONT VIEW





2.3.4. TITRATOR RIGHT-SIDE VIEW



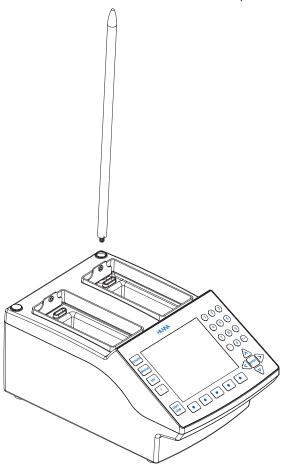
2.3.5. TITRATOR ASSEMBLY

Note: Assembly operations must be completed before connecting the Titrator to the power supply!

2.3.5.1. ASSEMBLING STIRRER AND ELECTRODES HOLDER

To assemble the electrode holder and support rod:

- Remove protective cap from titrator case
- Insert the support rod into the titrator case and turn it clock-wise to secure it in place

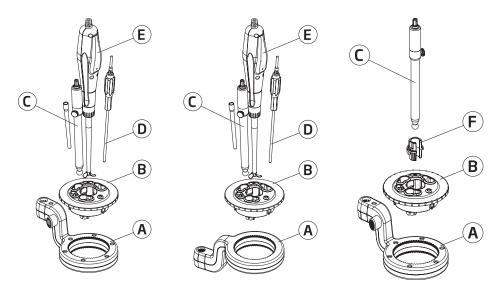


2.3.5.2. ATTACHING STIRRER AND ELECTRODES

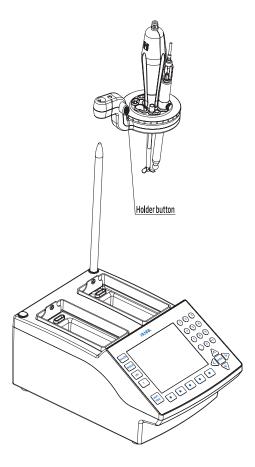
To attach the stirrer to the titrator, follow these steps:

- Place the electrodes holder (B) in the stirrer support housing (A). Stirrer support housing can be inverted if necessary.
- Insert electrode (C), temperature sensor (D) and stirrer (E) into the dedicated holes in the electrode holder. Push them until they are in stable position.

Note: For small sample sizes, use the electrode adapter (F) in the center of the holder.



• Slide the electrode holder into the support rod and set the desired height using the holder button.

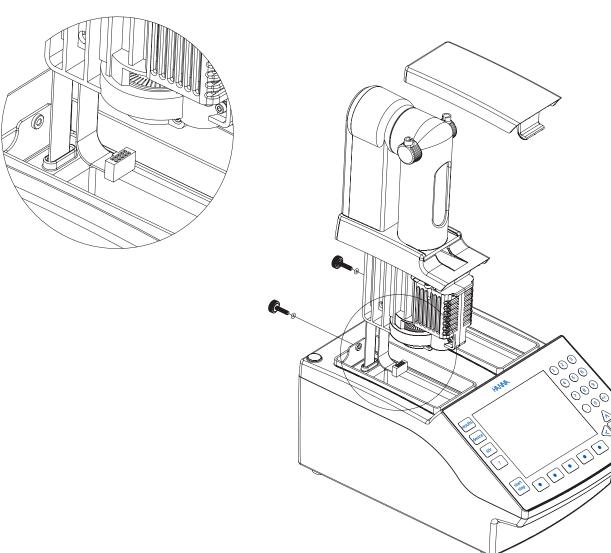


2.3.5.3. CONNECTING THE PUMP

To connect the pump, follow these steps:

- Retrieve the pump cable from inside the bay. The pump 1 connector is located in the left bay and pump 2 in the right bay.
- Connect the cable to the pump as shown below. The pump connector is located on the bottom of the pump.
- Lower the pump into the titrator, then slide it towards the front of the titrator case until it is firmly latched.
- Secure the pump with the locking screw.

This procedure can be repeated to connect a second pump.



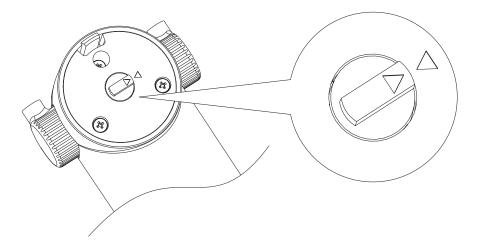
2.3.5.4. ATTACHING BURETTE BLANK SUPPORT

To attach the burette blank support, follow these steps:

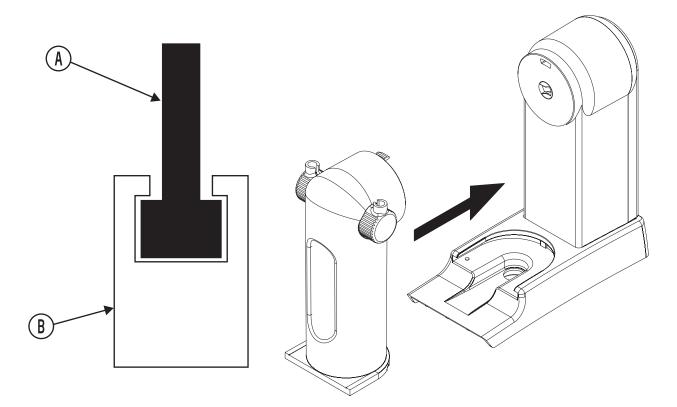
- Insert burette blank support into the bay. Lower the burette blank support into the titrator, then slide it towards the front of the titrator case until it is firmly latched.
- Secure the burette blank support with the locking screw.

2.3.5.5. ATTACHING THE BURETTE

Make sure that the mark from the valve actuating cap and from the burette body are aligned.



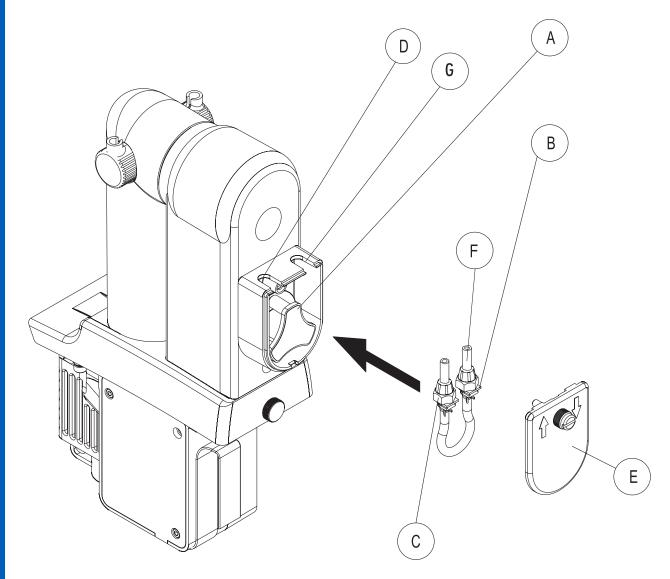
While ensuring the correct coupling between the syringe plunger (A) and the pump piston (B), slide the burette into the support on the burette pump.



2.3.5.6. CONNECTING PERISTALTIC PUMP TUBING

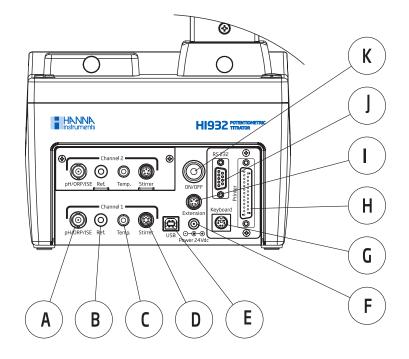
To attach the pump tubing to the burette pump with the built-in peristaltic pump:

- Use a screw driver to remove the plastic cover (E) from the pump.
- Remove the blue tube connectors (F).
- Insert the roller tube (C) into the left side of the holder (D). The fitting on the top of the roller tube will sit in the top of the housing.
- Manually rotate the pump (A) counter-clock wise until the tubing it mounted on the pump.
- Insert the roller tube (B) into the right side of the holder (G). The fitting on the top of the roller tube will sit in the top of the housing.
- Attach aspiration and dispensing tubing to the roller tubing and replace the blue tube connectors (F).
- Replace the plastic cover (E).



2.3.5.7. ELECTRICAL CONNECTIONS

- Connect the electrode to the BNC connector (A).
- Connect the temperature sensor to the RCA connector (C).
- Connect the stirrer to the MINI-DIN connector (D).
- Connect the power adapter cable to the power input connector (F).



Nr	Function	Type of Connector
A	Connection for pH, ORP, ISE half-cell and ISE combination electrodes	BNC Socket
В	Reference electrode	Ø 4 mm Banana Socket
С	Temperature sensor	RCA Socket
D	Stirrer	6-pin Connector
Ε	USB interface	USB Standard B
F	Power input connector (24VDC)	DC Power Jack Connector
G	External PC keyboard	6-pin Mini DIN (Standard PS2)
Η	Printer	DB-25 Socket
Ι	Connector for autosampler interface	5-pin Connector
J	RS232 interface (Balance Interface)	DB-9 Socket
K	Power switch	

CHAPTER 3. USER INTERFACE

3.1. START UP	3-3
3.2. DESCRIPTION	3-4
3.2.1. KEYPAD	3-4
3.2.1.1. FUNCTION KEYS	3-4
3.2.1.2. OPTION KEYS	3-4
3.2.1.3. ARROW KEYS	3-5
3.2.1.4. NUMERIC KEYS	3-5
3.2.1.5. ENTER KEY	3-5
3.2.2. DISPLAY	
3.2.3. THE MAIN SCREEN	3-6
3.3. MENU NAVIGATION	3-7
3.3.1. SELECTING AN OPTION	3-7
3.3.2. SELECTING A MENU ITEM	3-7
	3-7
	3-8

USER INTERFACE

3.1. START UP

Once the instrument is assembled and installed, follow the steps below to start the titrator:

- Connect the titrator to a power outlet with the supplied power adapter.
- Turn on the titrator from the power switch located on the back of the instrument.
- Wait until the titrator finishes the initialization process.
- Press enter when prompted or wait a few seconds for titrator to start.

HI 932 Titrator v1.00	
	ANNA struments
Analog Board 1 Analog Board 2 Pump 1 Pump 2	Potentiometric Potentiometric Burette/Peristaltic Not Detected
Press DEVICE to en Press HELP to view Press ENTER to con	

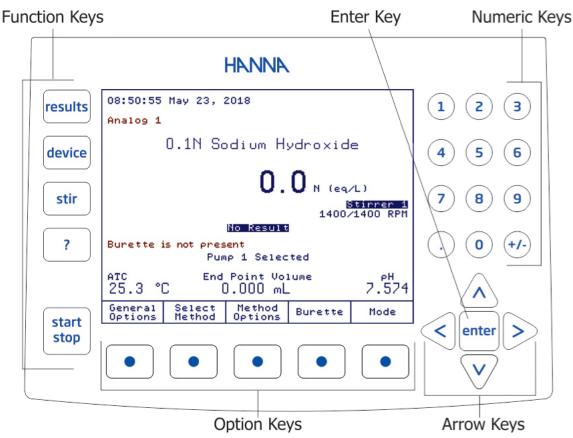
Note: All the performed initialization processes must be successfully completed. If one of the initialization processes fails, restart the titrator. If the problem persists contact your nearest Hanna Service Center.

3.2. DESCRIPTION

This chapter describes the basic principles of navigating through the user interface, selecting fields and entering values from the keypad.

3.2.1. **KEYPAD**

The titrator's keypad is grouped into five categories, as follows:



3.2.1.1. FUNCTION KEYS

If one of these keys is pressed, the associated function is immediately performed. Some of the keys are active only in specific screens:

Starts or Stops a titration

stir Turns the selected stirrer On and Off

device Access the Autosampler

results Access the Data Parameters Menu (reports, GLP, meter information, report setup)



3.2.1.2. OPTION KEYS

These keys are assigned to the virtual keys on the display. Their functions are listed in the boxes above the buttons and vary depending on the displayed screen.

An underlined virtual key can also be activated by pressing enter.

start stop

3.2.1.3. ARROW KEYS

These keys have the following functions:

- Move the on-screen cursor.
- Increase and decrease the stirrer speed and other settings.
- In the alphanumeric screen, to select a character.
- Navigate through menu options.

3.2.1.4. NUMERIC KEYS

(+/-)

 (\cdot)

Keys o to 9

Used for numeric entries.

- Toggles between positive and negative values.
-) Decimal point.

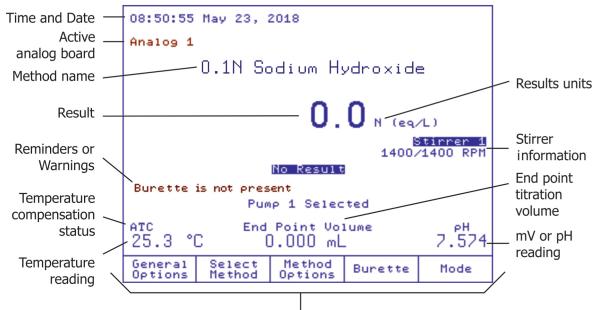
3.2.1.5. ENTER KEY

This key has the following functions:

- Accept alphanumeric data entry.
- Executes the default (underlined) virtual option key.

3.2.2. DISPLAY

The titrator has a large color graphical display. The main screen is shown below with short explanations of the screen segments.



Virtual option keys

The user interface contains several screens. For each titrator function, one or more screens are used.

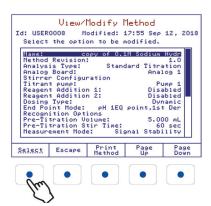
3.2.3. THE MAIN SCREEN

After start up and initialization, the first screen displayed is the main screen. Main screen fields:

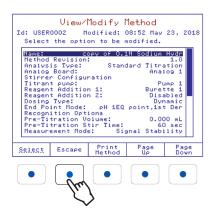
Method name:	Displays the name of the selected method.
Time and date:	Displays the current date and time.
Temperature reading:	Displays the measured temperature.
ATC:	Automatic temperature compensation
Manual:	Manual temperature compensation
Manual :	Temperature probe is not connected, manual temperature compensation
Stirrer information:	The selected stirrer and actual/set stirrer speed is displayed in RPM. When stirrer is off,
	the stirrer information is not displayed.
End point volume:	Displays the volume delivered to reach the titration end point. When no titration has been performed, the displayed volume is "0.000 mL".
Result:	Displays the titration result or the direct reading measurement.
mV or pH reading:	Displays the current readings. The reading will be in mV or pH.
mV:	Indicates actual potential reading.
rel mV:	Indicates relative potential reading.
pH:	Indicates actual pH value.
Titration status:	Displays the status of the selected titration.
	No Results is displayed when a titration has not been performed.
Reminders:	Indicates when a task needs to be performed and displays error
Pump 1 selected:	Displays the active pump.
Analog 1:	When two analog boards are present, the active one is shown.

3.3. MENU NAVIGATION





the ar Select	the high row keys the empt	ethod Na lighted 1 then pres	etter by i s "Enter" or a space	e.
Press	■ A B C P C A B C P C A B C P C A B C P C A B C P C A B C	save the DEFGH QESTGH	I U W I 4 30 1 U I 1 0 2 4 30 1 0 1 0 1 0 1 0 10 1 0 1 0 10 10 10 10 10 10 10 10 10 10 10	ane.
Accept	Escape	Delete Letter	Cursor Left	Cursor Right
- Erw				•



3.3.1. SELECTING AN OPTION

To select an option, simply press the option key below the virtual key. For example, to access the **Method Options** screen press the option key below it.

3.3.2. SELECTING A MENU ITEM

To select an item from the menu screen, use the arrow keys \triangle and \bigtriangledown to move the cursor.

When the menu is larger than the display, a scroll bar is active on the right side.

The $\begin{bmatrix} Page \\ Up \end{bmatrix}$ and $\begin{bmatrix} Page \\ Down \end{bmatrix}$ keys can be used to scroll through the pages.

To activate the selected menu item, press enter or Select .

3.3.3. ENTERING TEXT

To enter text in an alphanumeric input box, first erase the previous text by using To enter a letter, highlight it using the arrow keys then press enter. Use the same procedure to enter the whole name.

For editing, use the $\begin{bmatrix} Cursor\\ Left \end{bmatrix}$ and $\begin{bmatrix} Cursor\\ Right \end{bmatrix}$ keys.

When editing is complete, press Accept .

The method name will be updated and displayed in the name field of the **View/Modify Method** screen.

When all the desired parameters have been set, press Escape

	Sa	ving Met	hod	
Select	a menu o	ption.		
<u>Save M</u> Exit W	ethod ithout Sa	ving Meth	od	
"Escap	e" - exit	without	saving me	thod.
Select	Escape			
Im				
- Y 1				

3.3.4. SAVING MODIFICATIONS

The Saving Method screen allows the user to save the modifications. To exit from Saving Method screen without saving, press Escape or highlight the Exit Without Saving Method option and then press Select. To save the modifications highlight the Save Method option and then press Select.

Note: To access the contextual help menu, press ? at any time. Help is related to the displayed screen. Press escape or r to return to the previous screen.

CHAPTER 4. GENERAL OPTIONS

4.1. SAVE FILES TO USB STORAGE DEVICE	
4.2. RESTORE FILES FROM USB STORAGE DEVICE	4-4
4.3. ADMINISTRATION	
4.4. TEMPERATURE	4-6
4.4.1. TEMPERATURE SOURCE	
4.4.2. MANUAL TEMPERATURE SETTING	4-7
4.4.3. TEMPERATURE UNITS	
4.5. DATE AND TIME SETTING	4-8
4.6. DISPLAY SETTINGS	
4.7. BEEPER	4-9
4.8. STIRRER	
4.9. LANGUAGE	4-10
4.10. TOTAL VOLUME ALERT	
4.11. TITRANT AGE REMINDER	4-11
4.12. USB LINK WITH PC	4-11
4.13. SETUP BALANCE INTERFACE	4-12
4.14. PRINTER MODE	4-13
4.15. RESET TO DEFAULT SETTINGS	4-13
4.16. OPTIMIZE MEMORY SPACE	
4.17. UPDATE SOFTWARE	4-14

GENERAL OPTIONS

The **General Options** screen gives access to options that are not directly related to the titration process or pH/mV/ISE measurement. To access this screen, press General Options from the main screen.

	General Op	tions	
Select	the option to be	modified.	
Save to			
Admini: Temper: Date a	nd Time Setting	Disabled °C, ATC	
Beeper Stirre Langua	^: 9€:	Off Enabled English	
Titran Titran	t 1 Volume Alert: t 1 Age Reminder: t 2 Volume Alert: t 2 Age Reminder:	Off O days Off O days	
Select	Escape		

4.1. SAVE FILES TO USB STORAGE DEVICE

This option allows the user to save files from the titrator to a USB storage device.

On the titrator, the available file types are:

Standard Method Files User Method Files Report Files -HIXXXXYY.MTD (e.g.: HI0001EN.MTD, HI1004EN.MTD) -USERXXXX.MTD (e.g.: USER0001.MTD) -Ti_XXXXX.RPT, mV_XXXXX.RPT, pH_XXXXX.RPT, ISEXXXXX.RPT, mVrXXXXX.RPT

(e.g.: Ti_00001.RPT, mV_00001.RPT, pH_00001.RPT, ISE00001.RPT, mVr00001.RPT) Use the << and >> keys to select the file type. The number of files and the file name on the titrator will be displayed.

List of Files on Titrator Use <-/-> arrow keys to select file type				
13 sta	ndard meth	nod files		
H10004				
HI0002				
HI0010				
HI0200	EN.MTD			
HI1004				
HI1005				
HI1008				
HI1009EN.MTD				
HI1011				
HI1012EN.MTD HI1014EN.MTD				
Escape	Сору	Сору	Delete	Delete

The option keys allow the following operations:

Delete File	Delet
Delete All	Delet
Copy File	Соріє
Copy All	Соріє
Escape	Retur

Deletes the highlighted file.

Deletes all currently displayed files.

Copies the highlighted file from titrator to a USB storage device.

Copies all currently displayed files from titrator to a USB storage device.

Returns to the **General Options** screen.

The status of the transfer ("Successful"/"Unsuccessful") and the file name of the currently processed file are displayed during copying or deleting.

Note: The saved files will be stored on the USB key in the *HI932* folder, as follows:

- Methods: USB Drive:\HI932\Methods*.mtd
- Reports: USB Drive:\H1932\Reports*.rpt

4.2. RESTORE FILES FROM USB STORAGE DEVICE

This screen allows the user to transfer files from the USB storage device to the titrator.

Standard Method Files	-HIXXXXYY.MTD (e.g.: HI0001EN.MTD, HI1004EN.MTD)
User Method Files	-USERXXXX.MTD (e.g.: USER0001.MTD)
Report Files	-Ti_XXXXX.RPT, mV_XXXXX.RPT, pH_XXXXX.RPT, ISEXXXXX.RPT, mVrXXXXX.RPT (e.g.: Ti_00001.
	RPT, mV_00001.RPT, pH_00001.RPT, ISE00001.RPT, mVr00001.RPT)

Use the \lessdot and \triangleright keys to select the file type.

The number of files and the file name will be displayed.

List of Files on USB Use <-/-> arrow keys to select file type				
21 rep	ort files			
21 report files				
Escape	Copy file	Сору А11	Delete File	Delete All

GENERAL OPTIONS

The option keys allow the following operations:

Deletes the highlighted file from the USB storage device.

Deletes all currently displayed files from the USB storage device.

Copy File Copies the highlighted file from the USB storage device to the titrator.

Copy Copies all currently displayed files from the USB storage device to the titrator.

Escape Returns to the General Options screen.

Note: In order to restore files from a USB key, please ensure that the methods and/or reports you wish to transfer to the titrator are in the correct folder:

- Methods: USB Drive:\HI932\Methods*.mtd

- Reports: USB Drive:\HI932\Reports*.rpt

4.3. ADMINISTRATION

A 4-digit numeric PIN can be set to prevent unauthorized changes from being made.

When the user enters administration and a pin has not been set, the user will be prompted to enter a new PIN.

Titrator Administration					
Enter a	Administrator PIN has not been set. Enter a 4-digit PIN to enable Administrator function.				
	Enter	PIN:			
	Confirm	PIN: -			
Your PIN must be 4-digits long.					
Next.	Escape	Delete Digit			

Once a PIN has been set, the titrator can be locked. When a titrator is locked, the users cannot modify methods or delete reports. Basic functions are still available (review reports, save to USB, etc.).

]	litrator	Admini	stratior	ı
Titrato	r is UNLO	CKED.		
	Lock Titr Enter			
Accept	Escape	Delete Digit		

To return to administrator mode, the titrator can be unlocked by entering the PIN.

I	itrator	Admini	stratio	n
Titrato	∽ is LOCKE	D.		
Unlock <u>Titrator</u>	Escape			Recovery PIN

If the PIN is lost or forgotten, press recovery pin and contact technical support to supply the required information.

	Rei	covery H	PIN	
Vendor. When re	overy PIN questing ng inform	PIN please	contact yo e provide	our
	tor Seria 0078	1 Number:	1234562	28
	Recovery	PIN:		
Accept	Escape	Delete Digit		

4.4. TEMPERATURE

The **Temperature Menu** allows access to all of the settings related to temperature.

Temperature Menu							
Select	temperatur	e option	to be	modified.			
Manual	ature Sourc Temperatur ature Units	e Settin	9				
Select	Escape						

GENERAL OPTIONS

4.4.1. TEMPERATURE SOURCE

Option: Automatic Temperature or Manual Temperature

Select the temperature source used for temperature compensation.

When Automatic Temperature Compensation is selected, "ATC" is displayed on the main screen and the temperature is read by the temperature probe.

When Manual Temperature is selected, "Manual" is displayed on the main screen and a preset temperature value is used for temperature compensation.

Note: The selected temperature source will be indicated in the report files: A for Automatic and M for Manual.

Tembe	erature	Menu	
emperatur	e option	to be mo	dified.
Temperatu	Automat	ic Temper	ature
		Temperati	ine L
F			
	ture Sour Temoeratu	ture Source Temperatur ture Units Manual Manual	Temperatur Automatic Temper ture Units Manual Temperatu

4.4.2. MANUAL TEMPERATURE SETTING

Option: -5.0 to 105.0 °C (23.0 to 221.0 °F, 268.2 to 378.2 K)

If the temperature probe is not connected, the user can manually set the temperature used by the titrator for compensation.

	Manua	l Temper	rature			
when t	he temper	ature prol	ture to be de is bein ture probe	9		
		25.0	u∎ °C			
The temperature range is from -5.0 to 105.0°C.						
Accept	Escape	Delete Digit				

4.4.3. TEMPERATURE UNITS

Option: °C, °F or K

The temperature ranges are as displayed in the **Temperature Units** screen.

	Temperature Menu
Select	temperature option to be modified.
Manual	ture Source Temperature Setting ture Units
	Celsius -5.0 to 105.0 *C Fahrenheit 23.0 to 221.0 *F Kelvin 268.2 to 378.2 K
Select	Escape

4.5. DATE AND TIME SETTING

This screen allows the user to set the date and time.

Use the \bigtriangleup and \bigtriangledown keys or the numeric keys to modify the date and time.

Press <u>Next</u> to move the cursor to the next field.

Press AM / PM or 24-hour to change the time format.

	Date an	id Time	Setting	I
Enter	the date.			
	5 month	23 day	2018 year	
Enter	the time.			
	9 hour	11 minute	36 second	
Press	Next to m	ove to the	e next ent	try.
Accept	Escape	Delete Digit	Next	AM∠PM

4.6. **DISPLAY SETTINGS**

This screen allows the user to customize the display settings. Option Keys:

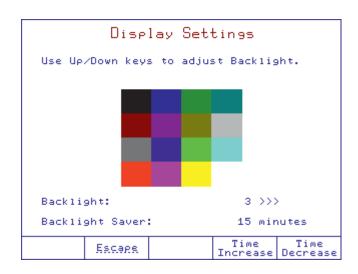


Increases the backlight saver time interval

me rease Decreases the backlight saver time interval

The backlight intensity can be adjusted using \triangle and \bigvee keys.

There are 8 levels of backlight intensity, ranging from 0 to 7.



A color palette is displayed in the center of the screen allowing an easy selection of the appropriate backlight intensity.

The backlight saver option protects the display during standby periods when no keys have been pressed for a set amount of time. If the display backlight is off, any keystroke will activate the backlight without performing any action.

The range for the backlight saver timer is 1 to 60 minutes. To disable the backlight saver, increase the time to the maximum allowed. The "Off" indication will appear.

4.7. BEEPER

Option: On or Off

If enabled (on) an audible alert will sound after a titration is completed, when an invalid key is pressed or when a critical error occurs during titration.

General Options	
Select the option to be modified.	
Save to USB Restore from USB Administration: Disabled Temperature: °C, ATC Date and Time Setting Display Settings	
Beeper: Off Stirrer: E Language: E Titrant 1 Volume Alert: On Titrant 1 Age Reminder: Off Titrant 2 Volume Alert: Off Titrant 2 Age Reminder: Odays	
<u>Select</u> Escape	

4.8. STIRRER

Option: Enabled or Disabled

Select the option to be modified. Save to USB Restore from USB Administration: Disabled Temperature: °C, ATC Date and Time Setting Display Settings Beeper: Off Stinner: Enabled Language: Titrant 1 Volume Alert: Titrant 1 Age Reminder: Titrant 2 Volume Alert: Titrant 2 Age Reminder: 0 days		Gene	eral Opt	tions		
Restore from USB Administration: Disabled Temperature: °C, ATC Date and Time Setting Display Settings Beeper: Off Stirren: Enabled Language: Titrant 1 Volume Alert: Disabled Titrant 1 Age Reminder: Enabled Titrant 2 Volume Alert:	Select	the opti	on to be	modified		
Titrant 1 Volume Alert: Titrant 1 Age Reminder: Titrant 2 Volume Alert:	Restore Adminis Tempera Date an Display Beeper:	from USE tration: ture: d Time Se Settings	≥tting		°C, ATC Off	
	Titrant Titrant Titrant	1 Volume 1 Age Re 2 Volume	Alert:		oled	

The stirrer can be disabled in individual titration method, if necessary.

4.9. LANGUAGE

Option: English, Português, or Español

General Options					
Output the sector is he had didied					
Select the option to be modified.					
	٦				
Save to USB					
Restore from USB					
Administration: Disabled					
Temperature: °C, ATC					
Date and Time Setting					
Display Settings					
Beeper: Off					
Stirrer: Enabled					
Language: English					
Titrant 1 Volume Alert:					
TTO and I Hge Keminder.					
Titrant 2 Age Reminder: Espanol					
Select Escape					
<u>Select</u> Escape					

4.10. TOTAL VOLUME ALERT

Option: Off, 0 to 10000 mL

This screen allows a programmable reminder to appear when the titrant reservoir is below 100 mL. The titrant volume will decrease as the titrant is used.

After the new titrant volume has been entered in the **Total Volume Alert** screen, a warning message appears on the main screen reminding the user re-standardize the titrant.

Т	itrant	1 Volu	me Aler	t	
the titr reservoi	ration/re ir. The (t of titra eagent sys MLs will o is deplet	stem from Jecrease	its	
		1000	u n mL		
A reminder will appear when less than 100 mLs of titrant volume is left.					
Accept	Escape	Delete		Off	

4.11. TITRANT AGE REMINDER

Option: Off, 0 to 31 days

A programmable reminder will appear when it is time to verify the titrant concentration or to change the titrant.

	Titrant	1 Age	Reminder	
last Ti	tr. Vol. (updating	to pass si or the las reminder a	st Start
		3	0 days	
The ran	ge is from	n 0 to 31	days.	
Start	Escape	Delete Digit		Off

4.12. USB LINK WITH PC

In order to use this feature, the USB cable needs to be connected from the titrator to the PC. Make sure that H1900 PC application is running on the PC.

USB Link with PC							
Inactive							
Speed 19200							
Escape							

"Active/Inactive": shows the status of the USB link with the PC.

"Active" means that the titrator is using the USB communication with the PC and not with another device.

"Ready" shows that the titrator is able to communicate with the PC.

During transfer of any information between the PC and the titrator, "Transmit" and information about the percentage of current file already transferred are displayed.

4.13. SETUP BALANCE INTERFACE

This screen allows the users to connect an analytical balance for automatic acquisition of sample mass prior to titration or standardization.

S	etup Ba	lance I	nterfac	e
Select t	he baland	to be a	activated.	
× Lab ba	alance			
Disable <u>Balance</u>	Escape	New Balance	Edit	

The balance is connected to the titrator via RS 232 interface.

- Press New Balance to add a new balance to the list.
- Press Enable Balance to enable the selected balance.
- Press Disable Balance to disable the selected balance (automatic weight acquisition will be not available).
- Press Edit to customize the name and serial communication parameters used by the selected balance.

	Balance	: Conf	iguration	
Select	the optio	n to be	modified.	
Balance Baud Ra Data Bi Parity: Stop Bi Request	ate: it:			9600 9600 8 bits Parity 1 bit 8
Select	Escape		Test Balance	

Be sure that the settings on the titrator *Balance Configuration* menu match the settings for your particular balance (baud rate, data bits, parity, stop bits number, request command syntax). It may be necessary to change settings on your balance. Users should consult their balance instruction manual.

Before leaving this screen, be sure the connection with the balance is working properly by pressing the Balance key.

4.14. PRINTER MODE

Option: Ansi, Ascii, or Text

	General Op	tions	
Select	the option to be	modified.	
	nd Time Setting / Settings	°C, ATC	
Stirrer Languag Titran	∿: 9e: t 1 Volume Alert:	Enabled English 1000.0 mL	
Titran Titran	t 1 Age Reminder: t 2 Volume Alert: t 2 Age Reminder: nk with PC	Ansi Ascii Text	
Setup B Printer	Balance Interface • Mode:	Ansi	
Select	Escape		

- Ansi mode: Use this mode when your printer is set as Ansi. In this case all the accented characters/symbols available in titrator will be printed on your printer.
- Ascii mode: Use this mode when your printer is set as Ascii. In this case only some of the accented characters/symbols available in titrator will be printed on your printer.
- Text mode: Use this mode when you don't need to print the accented characters.

4.15. RESET TO DEFAULT SETTINGS

Note: This will also delete all the user - created methods and restore all manufacturer settings such as titrator configuration, standard method parameters, etc.

	Confirm	nation o	f Reset	
		u want to ufacturer		
		e the cal: s, balance		
Reset	Escape			

4.16. OPTIMIZE MEMORY SPACE

Optimize Memory Space This option is used in order to clean up the memory space. Please ensure the power is not disconnected during this operation.					
the memory space. Please ensure the power is not		Optimiz	e Memor	у Ѕрасе	2
Please ensure the power is not				der to ci	lean up
					n.
	Accept	Escape			

4.17. UPDATE SOFTWARE

This screen allows the user to update the titrator software from a USB storage device containing a software setup kit.

	Update Software					
Curren	Current version: HI932 v1.00					
New ve	rsion:	H	HI932 v1.01			
Are you sure you want to update the current software with the new version?						
Accept	Escape	Refresh				

To update the software:

- Copy the "Setup932" folder to a USB storage device.
- Insert the USB storage device into the titrator.
- Go to General Options, then Update Software. The titrator will display the current and new software versions.
- Press Accept . When prompted, remove the USB storage and restart the titrator.

CHAPTER 5. TITRATION METHODS

5.1. SELECTING METHODS	5-3
5.2. STANDARD METHODS	5-3
5.2.1. UPGRADING STANDARD METHODS	
5.2.2. DELETING STANDARD METHODS	5-4
5.2.3. RESTORE THE STANDARD METHODS TO THE MANUFACTURER SETTINGS	
5.3. USER METHODS	5-5
5.3.1. CREATING USER METHODS	
5.3.2. DELETING USER METHODS	5-5
5.4. VIEW / MODIFY METHOD	5-6
5.5. METHOD OPTIONS	5-6
5.5.1. NAME	5-6
5.5.2. METHOD REVISION	5-7
5.5.3. ANALYSIS TYPE	
5.5.3.1. STANDARD TITRATION	5-7
5.5.3.2. BACK TITRATION	5-7
5.5.3.3. DIRECT READING	5-8
5.5.4. ANALOG BOARD	
5.5.5. STIRRER CONFIGURATION	5-9
5.5.5.1. STIRRER	5-9
5.5.5.2. STIRRER SPEED	5-9
5.5.6. PUMP CONFIGURATION	
5.5.7. REAGENT ADDITION	5-10
5.5.7.1. ADDITION VOLUME (BURETTE)	5-11
5.5.7.2. DISPENSING TIME (PERISTALTIC PUMP)	5-11
5.5.7.3. STIRRING TIME	5-12
5.5.7.4. WAIT TIME	5-12
5.5.8. MEASUREMENT PARAMETER (DIRECT READING ONLY)	5-13
5.5.9. DOSING TYPE	5-13
5.5.9.1. LINEAR DOSING	5-13
5.5.9.2. DYNAMIC DOSING	5-14
5.5.10. END POINT MODE	5-15
5.5.10.1. FIXED END POINT (pH OR mV)	5-15
5.5.10.2. EQUIVALENCE END POINT (pH OR mV)	5-16
5.5.11. RECOGNITION OPTIONS (EQUIVALENCE END POINT ONLY)	5-19
5.5.11.1. THRESHOLD	5-19
5.5.11.2. RANGE	5-20
5.5.11.3. FILTERED DERIVATIVES	5-21
5.5.12 .PRE-TITRATION VOLUME	5-22
5.5.13 .PRE-TITRATION STIR TIME	

TITRATION METHODS

5.5.14 .MEASUREMENT MODE	5-23
5.5.14.1 SIGNAL STABILITY	5-23
5.5.14.2 TIMED INCREMENT	
5.5.15 .ELECTRODE TYPE	5-25
5.5.16 .BLANK OPTION	
5.5.17. CALCULATIONS	5-26
5.5.17.1. STANDARD TITRATION	5-26
5.5.17.1.1 EDIT VARIABLE VALUES	5-26
5.5.17.1.2. NO FORMULA (mL ONLY)	
5.5.17.1.3. NO FORMULA (L ONLY)	5-26
5.5.17.1.4. SAMPLE CALCULATION BY WEIGHT	5-27
5.5.17.1.5. SAMPLE CALCULATION BY VOLUME	5-28
5.5.17.1.6. STANDARDIZE TITRANT BY WEIGHT	
5.5.17.1.7. STANDARDIZE TITRANT BY VOLUME	5-30
5.5.17.1.8. GENERIC FORMULA	5-31
5.5.17.2. BACK TITRATIONS	5-33
5.5.17.2.1 SAMPLE CALCULATION BY WEIGHT	
5.5.17.2.2 SAMPLE CALCULATION BY VOLUME	5-35
5.5.17.2.3 GENERIC FORMULA	5-37
5.5.18. DILUTION OPTION	5-37
5.5.19. TITRANT NAME	5-37
5.5.20. TITRANT CONCENTRATION	5-38
5.5.21. ANALYTE SIZE	5-38
5.5.22. ANALYTE ENTRY	5-38
5.5.22.1. FIXED WEIGHT OR VOLUME	5-39
5.5.22.2. MANUAL WEIGHT OR VOLUME	
5.5.22.3. SAME AS PREVIOUS (LINKED METHODS ONLY)	
5.5.23. TITRANT 1 ENTRY (BACK TITRATION ONLY)	
5.5.23.1. CALCULATED BY FORMULA	5-39
5.5.23.2. FIXED BY USER	5-39
5.5.24. MAXIMUM TITRANT VOLUME	5-40
5.5.25. POTENTIAL RANGE	5-40
5.5.26. VOLUME/FLOW RATE	
5.5.27. SIGNAL AVERAGING	5-41
5.5.28. SIGNIFICANT FIGURES	5-42
5.5.29. LINKED METHOD	5-42
5.5.30. START LINKED METHOD (LINKED METHOD ONLY)	5-43
5.6. PRINTING	5-43

All of the parameters required to complete an analysis are grouped into a method.

The titrator is supplied with a pack of standard methods, these methods have been developed by Hanna Instruments and can be used to create user methods.

Standard and user methods can be upgraded, saved or deleted by connecting the titrator to a PC using the H1900 PC application or a USB storage device.

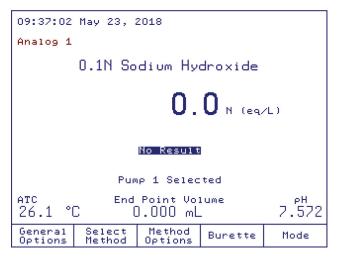
5.1. SELECTING METHODS

To select a method, press Select Method from the main screen. A list of available methods will be displayed.

	F	ìnal	ysis Met	chods	
Select	the	meth	od to be a	activated.	
H100	01EN	0.1N	Sodium Hy	/droxide	
HIOO	02EN	0.1N	Hydroch1d	pric Acid	
HIOO	03EN		Sodium Th		
HTOO	10EN	0.1M	EAS		
			M Silver N	litrate	
			linity of		
			ity of Wat		
			ride in Wa		
			ralizatior		
			ralizatior ralizatior		
			bleshootir		
			bleshootir		
			entration		
USER	0001	COPY	of Acidit	y of wate	er
-					
Select		≅W.	Reset to	Page	Page
A MARKA	Met	hod	Default	Uρ	Down

In the Analysis Methods screen, you can view the list of all available methods (standard and user methods).

To select a method, highlight the method then press Select, the name of the selected method will be displayed on the main screen.



5.2. STANDARD METHODS

The standard methods are developed for the most common types of analysis. Only specific method parameters can be modified by the user (see **Method Options** section). Also, standard methods can be used as a template to create new user methods.

5.2.1. UPGRADING STANDARD METHODS

To upgrade the titrator with new standard methods, follow the steps below:

From USB Storage Device:

- Insert the USB storage device into the USB port, located on the right side of the titrator.
- Press General from the main screen.
- Using 🛆 and 👽 keys, highlight the *Restore from USB* Storage Device option and choose select
- Using <a> and >> keys, navigate through file types to find "standard method files". The list with available standard methods will be displayed.
- Press the Copy File or All key to upgrade the titrator with the standard methods.
- Press Escape to return to General Options screen.

From PC:

You can upgrade the titrator with standard methods from a PC using the H1900 PC application (see General Options section).

5.2.2. DELETING STANDARD METHODS

Unnecessary standard methods can be removed from the titrator by following the procedure below:

From General Options Screen:

- Using the 🛆 and 👽 keys, highlight the *Save to USB* Storage Device option and press select.
- Using the <a> and >> keys, navigate through the file types menu to find "standard method files". The available standard methods will be displayed.
- Press the Delete or All keys to remove unnecessary standard methods.
- Press Escape to return to the General Options screen.

From PC:

Unnecessary standard methods can be removed from the titrator using the H1900 PC application (see General Options section).

5.2.3. RESTORE THE STANDARD METHODS TO THE MANUFACTURER SETTINGS

You can restore the standard methods to the default setting by highlighting a standard method and pressing Reset to Default

Analysis Methods	Confirmation of Reset Methods
Select the method to be activated.	
HI0001EN 0.1N Sodium Hydroxide HI0002EN 0.1N Hydrochloric Acid HI0003EN 0.1M Sodium Thiosulfate HI0010EN 0.1M FAS HI0200EN 0.02M Silver Nitrate HI1004EN Alkalinity of Water HI1005EN Acidity of Water HI1007EN Chloride in Water HI1007EN Neutralization w/ H2S04 HI1009EN Neutralization w/ NaOH HI1011EN Troubleshooting 1 HI1012EN Troubleshooting 2 HI1014EN Concentration of H3P04 USER0001 copy of Acidity of Water	Are you sure you want to reset all Standard Methods to default?
Select New Reset to Page Page Method Default Up Down	Reset Escape

TITRATION METHODS

5.3. USER METHODS

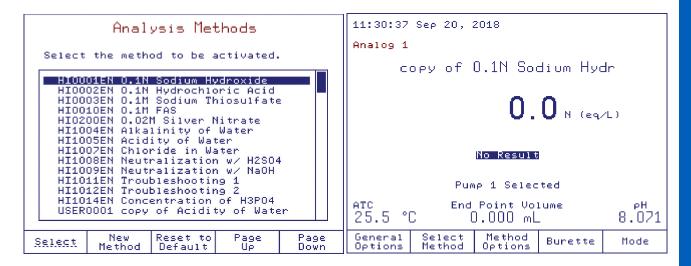
These methods are defined by the user (usually by modifying a standard method).

The user methods can be developed in accordance with the requirements of the user. All method parameters can be modified by the user.

5.3.1. CREATING USER METHODS

To create a new user method, start from a standard or user method and follow these steps:

- Press Method from the main screen.
- Using the \bigwedge and \bigvee keys, highlight an existing method from the method list.
- Press <u>Method</u>. A new user method will be generated.
- Press to activate the new user method.



Note: Only a limited number of methods can be installed on the titrator. The titrator can hold 100 methods (standard and user). When it is reached, a warning message will be displayed.

5.3.2. DELETING USER METHODS

To remove a user method, press Select Method from the main screen. Highlight the user method that you want to delete and press Delete A screen will appear in order to confirm the deletion. Press Delete again to confirm, or press Escape to cancel the operation.

Confi	rmation	of Met	hod Del	etion
A				
	u sure you ed method'		delete tr	ne
сору о	f 0.1N Soc	dium Hydr		
Delete	Escape			

5.4. VIEW / MODIFY METHOD

To modify the method parameters, press Method displayed. Using the A and keys, highlight the option that you want to modify and choose selected. To exit the **View / Modify Method** screen, press Escape.

	· · · · · · · · · · · · · · · · · · ·	*		
	View/	Modify N	1ethod	
Id: USER0	0008 M	odified: :	17:55 Sep	12, 2018
Select	the optio	on to be r	nodified.	
		opy of 0.1	LN Sodium	Hydn
Analysi Analog Stirrer Titran Reagen Reagen Dosing End Poi Recogni Pre-Tit Pre-Tit	Board: Configur t Configur t Addition t Addition Type: Int Mode: Ition Opt: tration S	Stand ration n 1: n 2: PH 1EQ	Anal Pu Disa Disa Dyr point,1st 5.00 60	.og 1 abled abled amic ; Der)0 mL) sec
Select	Escape	Print Method	Page Up	Page Down

You can choose to save the modifications or to discard them.

Saving Method					
Select a menu option.					
<mark>Save Me</mark> Exit Wi		ving Metho	od		
"Escape	e" = exit	without s	saving met	hod.	
Select	Escape				

5.5. METHOD OPTIONS

Note: Only certain method options can be changed for standard methods.

5.5.1. NAME

Option: Up to 24 characters

	Method Name			
Select the highlighted letter by using the arrow keys then press "Enter". Select the empty field for a space. Press Accept to save the entire name.				
	Z a b c m n o p z A A A b o o é é ù ú ü ¿ 0 1 2 3 ? ! ()	D E F G H Q R S T U d e f 9 h q A C E E び ひ ひ じ B さ i 1 元 \$ 6 i 2 6 7 = [] く 7 = U.1N Soci	U W X 1 1 j k 1 1 v X 1 2 a a a a 6 a a a a 1 v X 1 2 a a a 6 a a a 1 v X 1 2 a a 6 a a 7 x - 7 x - 7 x -	
Accept	Escape	Delete Letter	Cursor Left	Cursor Right

5.5.2. METHOD REVISION

Option: Up to 3 characters

	Metł	nod Revi	sion	
the ar Select	row keys the empt	lighted le then press y field fo ring forma	s "Enter". or a space	2.
	Zabc mnop zàáâ bdóm	D E F G H Q R S T U d e f 9 h q r s t U 浴 ひ í n ð i * 1 ~ 7 8 [] く > =	VWXY ijk1 vwxy ÉIIÑ àáàā	
Accept	Escape	Delete Letter	Cursor Left	Cursor Right

5.5.3. ANALYSIS TYPE

Option: Standard Titration, Back Titration or Direct Reading

	Âna	lysis T	уре	
Select	the analy	vsis type.		
Back T	nd Titrati itration Reading	ion		
Select	Escape			

5.5.3.1. STANDARD TITRATION

- A titration with a pH or mV equivalence point detection (single or multiple equivalence points).
- A titration with fixed pH or mV end point.
- A titrant standardization.

5.5.3.2. BACK TITRATION

A titration with a pH or mV equivalence point detection consisting of two titration phases:

- Phase 1 the sample is consumed by a known volume and concentration of titrant 1. A sufficient amount of titrant 1 is dispensed to surpass the equivalence point in order to react quickly with the sample.
- Phase 2 the excess of titrant 1 is titrated with the titrant 2 to the equivalence point. The concentration of the sample is determined by the amount of titrant used in phase 2.

BREAK AT TITRANT CHANGING

Option: Yes or No

Select "Yes" to stop the titration temporarily between the titration phases, this allows users to perform a task related to the analysis (e. g.: boiling the sample to remove carbon dioxide, pH adjustment, etc.)

Br	eak at	Titrant	Changi	ng
Select	the opti	on.		
NO Yes				
		break at t eak at tit		
Select	Escape			

5.5.3.3. DIRECT READING

A direct pH, mV or ISE reading with an optional reagent addition. The titrator will take the measurement automatically once a stable reading has been obtained.

5.5.4. ANALOG BOARD

Option: Analog 1 or Analog 2 (if installed)

	0003 M	Modify odified: on to be (09:40 May	23, 2018
Method Analys Stirre Titran Titran Reagen Reagen Dosing End Po Recogn Pre-Ti	Revision is Type: Board: r Configu t 1 pump: t 2 pump: t Addition t Addition Type:	Stand ration n 1: n 2: PH 1EQ ions olume:	dard Titra Anal Analo Analo Disa Dyr point,1st	1.0 ation og 1 g 2 abled aamic ; Der
Select	Escape	Print Method	Page Up	Page Down

5.5.5. STIRRER CONFIGURATION

Use the arrow keys to select the menu option.

	Stirrer Conf	iguration
Select	a menu option.	
<mark>Stirrer</mark> Stirrin	9 Speed:	Stirrer 1 1400 RPM
Select	Escape	
XXXXXXX		

5.5.5.1. STIRRER

Option: Stirrer 1, Stirrer 2 (if available), or Disabled

	Stirrer	Config	ura	tion		
Select	a menu op	tion.				_
<mark>Stirrer</mark> Stirrin	g Speed:			Disa		
				Stir	rer 1 rer 2	
Select	Escape					

5.5.5.2. STIRRER SPEED

Option: 200 to 2500 RPM

	Stirring Speed				
Enter the below rai		of the st	tirrer wit	thin:	
		1400	RPM		
The range	e is fro	om 200 to	2500 RPM.		
Accept B	Escape	Delete Digit			

5.5.6. PUMP CONFIGURATION

Option: Pump 1 or Pump 2 (if installed)

Note: For back titrations, the pump for titrant 1 and titrant 2 need to be selected.

Name:	03 M he opti- cvision Type: oard:	on to be r opy of 0.: Stand	D9:40 May modified. 1N Sodium dard Titra	Hydr	018
Titrant Titrant Reagent Dosing T End Poin Recognit Pre-Titr	1 pump: 2 pump: Additio Additio ype: t Mode: ion Opt ation V	n 1: n 2: PH 1EQ	Point,1s1	p 2 ; Der	
Select	Escape	Print Method	Page Up	Pag Dow	

5.5.7. REAGENT ADDITION

Option: Burette, Peristaltic Pump or Disabled

	Reagent Addition 1				
Select	Select the option to be modified.				
Reagent	Reagent Pump: Disabled				
			Uisabled Burette 1 Peristalt Burette 2 Peristalt	ic 1	
Select	Escape				

Use the arrow keys to select the menu option.

Reagent Addition 1	Reagent Addition 1			
Select the option to be modified.	Select the option to be modified.			
Reagent Pump:Burette 1Addition Volume:0.000 mLStirring Time:2 secWait Time:1 sec	Reagent Pump:Peristaltic 1Uispensing Time:10 secStirring Time:15 secWait Time:30 sec			
Select Escape	<u>Select</u> Escape			

5.5.7.1. ADDITION VOLUME (BURETTE)

Use the numeric keypad to enter the volume to be dispensed.

	Add i	tion Vo	lume	
	e additio the samp		volume to) be
5.000 mL				
Press Help to view the valid ranges for the addition volume.				
Accept	Escape	Delete Digit		

The volume dispensed must be between the limits shown below:

0.001 to 4.750 mL for a 5 mL Burette

0.001 to 9.500 mL for a 10 mL Burette

0.005 to 23.750 mL for a 25 mL Burette

0.005 to 47.500 mL for a 50 mL Burette

5.5.7.2. DISPENSING TIME (PERISTALTIC PUMP)

Option: 1 to 300 seconds

Enter the dispensing time required to add the desired amount of reagent.

Note: The user should determine this value experimentally. The approximate dispensing rate is 200 mL/min.

Dispensing Time					
	ne period ry pump.	of time (for runnir	19	
5 sec					
Low limit: 1 second High limit: 300 seconds					
Accept	Escape	Delete Digit			

5.5.7.3. STIRRING TIME

Option: 1 to 1800 seconds

The timer will start after the reagent has been added.



5.5.7.4. WAIT TIME

Option: 1 to 1800 seconds

The timer will start after the stirring timer.

Wait Time					
Please	enter the	wait time	in seconds.		
			sec 🗧		
Low limit: 1 second					
High limit: 1800 seconds					
Accept	Escape	Delete Digit			

5.5.8. MEASUREMENT PARAMETER (DIRECT READING ONLY)

Option: pH, ISE or mV $% \left({{\left| {{{\bf{N}}} \right|}} \right)$

Select the measurement parameter for the direct reading. Setup screen for the selected parameter is visible in the method options.

Id: USER Select			09:40 May	23, 2018
Analys Analog Stirre Reagen Reagen	Revision is Type: Board: r Configu t Additio t Additio gment Par	ration n 1: n 2:		PH 1.0 MV ISE
	ode Type: To:	ameter.	No	рН рН Link
			_	
Select	Escape	Print Method	Page Up	Page Down

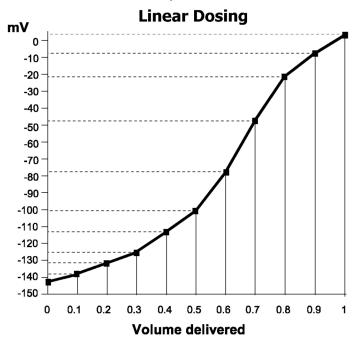
5.5.9. DOSING TYPE

Option: Linear or Dynamic

Dosing Type					
Select	the dosi	ng type.			
Linear Dosing Dynamic Dosing					
Select	Escape				

5.5.9.1. LINEAR DOSING

Linear dosing dispenses a pre-defined volume of titrant with every addition.



The *Linear Dosing* option is recommended for titrations with a slower reaction rate, difficult nonaqueous titrations, and specific applications.

Note: For steep and normal titration curves, smaller volume increments are recommended, to obtain many points around the equivalence point.

For flat titration curves, larger volume increments are recommended for equivalence point detection.

To set the dosing volume, select *Linear Dosing* and enter the optimum dose.

Dosing volume ranges are:

	0 001		
5 mL burette	0.001	to	4.750 mL
10 mL burette	0.001	to	9.500 mL
25 mL burette	0.005	to	23.750 mL
50 mL burette	0.005	to	47.500 mL

5.5.9.2. DYNAMIC DOSING

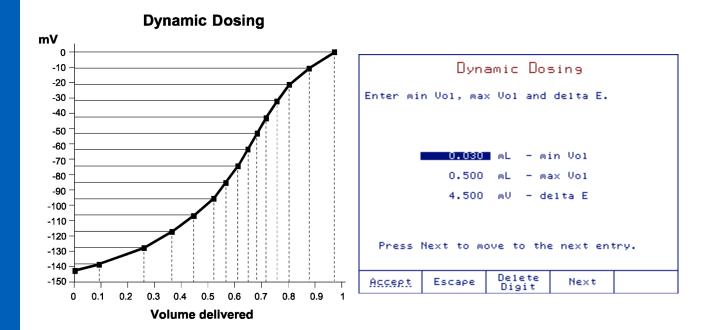
The titrator determines the titrant dose by trying to maintain a certain potential change (*delta E*) with each addition.

After a titrant dose, if the potential change is lower than the set *delta E*, the next dose will be progressively increased until *max Vol* is attained. If the potential change is still lower than the set value, the titration will continue with *max Vol* doses.

After a titrant dose, if the potential change is higher than the set *delta E*, the next dose will be progressively decreased until *min Vol* is attained. If the potential change is still higher than the set value, the titration will continue with *min Vol* doses.

The titrant is added in volumes that depend on the proximity of the end point as shown in the graph below.

Dynamic dosing allows for larger doses far from the end point, reducing the total titration time. Closer to the end point, smaller doses are made, providing more data and improved accuracy.



The following parameters must be set:

min Vol: The smallest dose to be dispensed during a titration.

The *min Vol* must be greater than or equal to: 0.001 mL for a 5 mL burette 0.001 mL for a 10 mL burette 0.005 mL for a 25 mL burette 0.005 mL for a 50 mL burette The max Vol must be less than or equal to 4.000 mL.

delta E: Sets the fixed potential jump that has to be achieved after each titrant dose. The allowed range is between 0.1 and 99.999 mV.

Recommendations for dosing parameters:

For steep and normal titration curves the recommended settings are:

delta E	3.5	to	9 mV
min Vol	0.010	to	0.025 mL (for a 25 mL burette)
max Vol	0.075	to	0.250 mL (for a 25 mL burette)
For flat titration curves the recomm	nended settings a	Ire:	
delta E	10	to	15 mV
min Vol	0.050	to	0.150 mL (for a 25 mL burette)
max Vol	0.400	to	0.600 mL (for a 25 mL burette)

To achieve the highest levels of accuracy and reproducibility, it is recommended that 20-80% of the nominal burette volume used for each titration is consumed. If lower volumes of titrant are required, a smaller burette can be used.

5.5.10. END POINT MODE

Option: Equivalence Point (pH or mV) or Fixed End Point (pH or mV)

Т	itratio	n End	Point	t Mod	de	
Select	the end ρ	point d	detecti	on.		
Equiva Fixed	lence End lence End nd Point	Point (pH)				
Fixed	End Point	(MV)				
Select	Escape					

5.5.10.1. FIXED END POINT (pH OR mV)

Fixed End Point (pH):

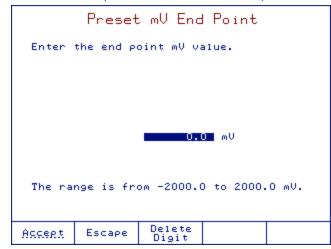
Option: -2.000 to 20.000 pH

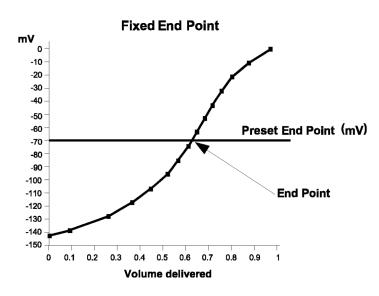
The titration is terminated when the preset pH value has been exceeded. The end point volume is a calculated value based on the dispensed volume when pH is under the preset value and the dispensed volume when pH exceeded the preset value.

	Preset	: eH End	l Point	
Enter	the end po	oint pH v∶	alue.	
		8.600	PH D	
The ra	nge is fro	om -2.000	to 20.000) рН.
Accept	Escape	Delete Digit		

Fixed End Point (mV): Option: -2000.0 to 2000.0 mV

The end point detection algorithm is the same as for pH, but the threshold value is expressed in mV.



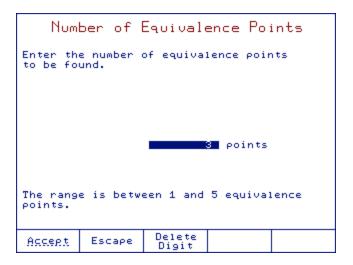


5.5.10.2. EQUIVALENCE END POINT (pH OR mV)

The titration is terminated when the equivalence point is detected (the point where the added quantity of titrant equals the quantity of analyte present in the sample).

Number of Equivalence Points:

Option: 1 to 5



End Point Determination
Select the end point determination.
<mark>Ast derivative</mark> 2nd derivative
Select Escape

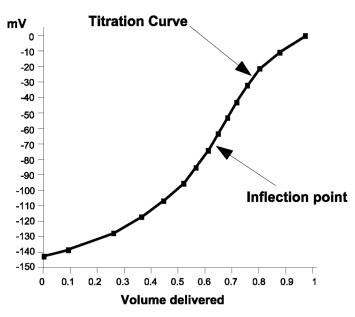
The equivalence point detection algorithm requires three additional titrant doses to be dispensed after the equivalence point is reached.

The reported end point volume is a calculated value based on a number of points around the equivalence point.

The potentiometric titration curve is the response in mV potential or pH between the indication of the electrode versus the volume of titrant added.

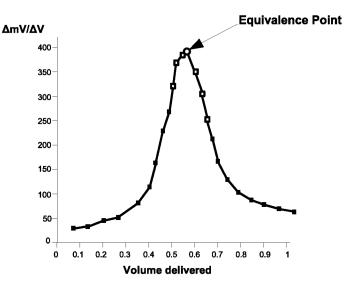
The inflection point of the titration curve is assumed to be the equivalence point of the chemical reaction.

For non-symmetric titration curves, the theoretical error can be reduced by using the dynamic dosing.



1st Derivative:

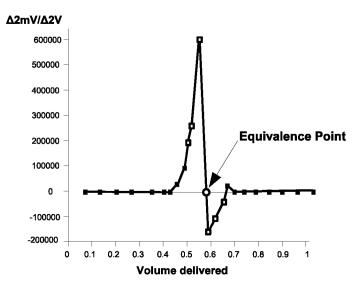
When first derivative is used to recognize the equivalence point, the titration curve inflection point (EQP) is the point where the first derivative reaches its maximum value.



The detection algorithm looks for the maximum value of the first derivative. The first derivative must be greater than the threshold value at the maximum point (see **Recognition Options** section).

2nd Derivative:

When second derivative is used to recognize the equivalence point, the titration curve inflection point (EQP) is the point where the second derivative crosses zero.



The detection algorithm looks for the point where the second derivative changes sign.

The checked point, or first derivative, must be greater than the threshold value (see Recognition Options section).

TITRATION METHODS

5.5.11. RECOGNITION OPTIONS (EQUIVALENCE END POINT ONLY)

The **Recognition Options** screen is a set of parameters used to avoid false detection of the equivalence point due to the chemical system (titrant/sample species and concentrations) and/or electrode response.

	Recogn	ition	Option	5	
Select recogn	the optic ition.	ons for	equivale	nce point	
had been a second se					
Thresh	010		500) mV/mL	
Range				NO	
Filter	ed Derivat	tives		NO	
Select	Escape				

5.5.11.1. THRESHOLD

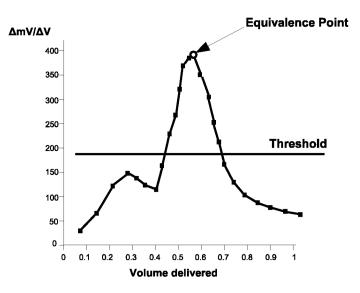
Option: 1 to 9999 mV/mL

This parameter must be set by the user according to the analysis.

The threshold represents the absolute value of the first derivative, expressed in mV/mL, below which the detection algorithm does not search for the equivalence point.

]	lhresho l	Ь	
Enter th detectio	e thresho n.	ld for equ	Jivalence	point
EQ 1	Threshold	: 50	u∎ mV∕mL	
1 and 4 450 and	ded value 50 mV/mL f 1800 mV/n d 9999 mV/	or FLAT C AL for NOR	Curve, MAL Curve	
Accept	Escape	Delete Digit		Next Threshold

The recommended value is 40% of the absolute value of the first derivative.



Depending on the titration curve profile, the following guide can be used:

TITRATION CURVE PROFILE	THRESHOLD (mV/mL)
Flat	1 to 450
Normal	50 to 1800
Steep	1800 to 9999

5.5.11.2. RANGE

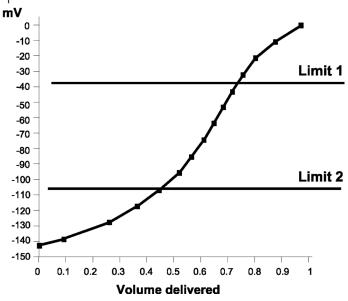
Option: -2.000 to 20.000 pH or -2000.0 to 2000.0 mV

Range is an optional feature for equivalence point recognition. The titrator will only look for an equivalence point between the set values.

The Range option can be enabled by selecting "Yes" in the Range Options screen.

Range Options	Range Limits
Select option for equivalence point range.	Enter Limit 1 and Limit 2 for range.
NO YES	
	-2.0 mV - EQ 1 Limit1
	20 mV - EQ 1 Limit2
"NO" - without equivalence point range. "YES" - with equivalence point range.	Press <next eq="" range=""> for the next range.</next>
<u>Select</u> Escape	Accept Escape Delete Next Next Digit Limit EQ Range

The Limit 2 value must not be equal to the Limit 1 value.



5.5.11.3. FILTERED DERIVATIVES

Option: Yes or No

This option adds a filtering procedure in the 1st and 2nd derivative computation algorithm that reduces the influence of pH or mV noise.

The *Filtered Derivative*s option can be enabled by selecting "*Yes*" in the **Filtered Derivatives Option** screen.

Fil	tered [)erivati	ves Opt	ion
Select o	ption for	filtered	derivativ	ves.
NO YES				
		filtered tered der	derivativ ivatives.	es.
Select	Escape			

Noise can be due to:

- Chemical system properties (sample, titrant, solvent), such as slow chemical reactions or unbuffered samples such as wastewater, tap water, wine
- Electrode response
- Incorrect method parameters settings such as Signal Stability, Stirring Speed, etc.
- Insufficient titrant additions

Note: A shift in the end point volume by 1 or 2 doses may be seen due to filtering.

5.5.12. PRE-TITRATION VOLUME

During a titration, the equivalence point is reached after many titrant doses. These doses take up extra time while having no relevance for equivalence point detection.

Pre-titration volume adds a large initial dose to jump directly to the proximity of the equivalence point.

This first dose occurs after the pre-titration stir time is completed.

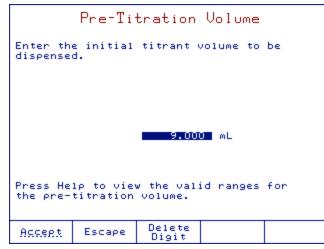
The ranges for pre-titration volumes are shown below:

0.001 to 4.750 mL for a 5 mL burette

0.001 to 9.500 mL for a 10 mL burette

0.005 to 23.750 mL for a 25 mL burette

0.005 to 47.500 mL for a 50 mL burette



To disable a pre-titration volume, enter 0.000 mL.

Note: A pre-titration volume is highly recommended whenever possible. Fewer doses will considerably shorten the overall titration duration.

5.5.13. PRE-TITRATION STIR TIME

Option: 0 to 180 seconds

When enabled, the sample is mixed for a set period of time before any titrant is added. This allows the sample to become homogeneous.

P	re-Titr	ation S	tir Tim	e
		mixing ti titration.		to
		1	J second:	5
The range	: is from	0 to 180	seconds.	
Accept	Escape	Delete Digit		

The Pre-Titration Stir Time option is disabled if 0 seconds is entered.

5.5.14. MEASUREMENT MODE

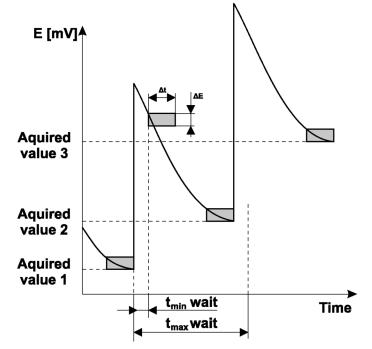
Option: Signal Stability or Timed Increment

During titration, the acquisition of the potential (mV) value of the solution can be done in two ways: by using either *Signal Stability* or *Timed Increment* option.

	Meas	urement	Mode	
Select	the meas	urement mo	ode.	
	Stabilit Increment	ý		
Select	Escape			

5.5.14.1. SIGNAL STABILITY

When *signal stability* is selected, the titrator acquires the potential (mV) only when stable conditions are reached. The principles of signal stability are plotted below:



The signal stability window (condition) represents the time interval (Δt) during which the potential measured in solution (mV) is confined inside the potential interval (ΔE).

The new signal value is acquired if the stability condition is reached after the minimum (t min) wait time.

If the stability condition is not reached and the maximum (t max) wait time has elapsed, the potential is acquired.

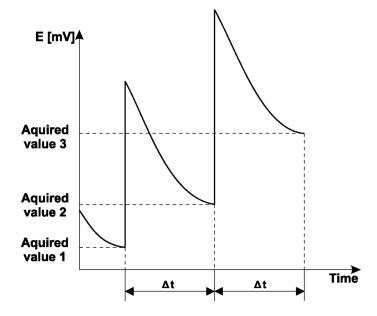
	Signa	al Stab:	ility	
interval	variation (delta t)) the next	min and	max wait	time
	0.3	μŲ	- delta	E
	2	seconds	- delta	t
	3	seconds	- t min	wait
	30	seconds	- t max	wait
Accept	Escape	Delete Digit	Next	

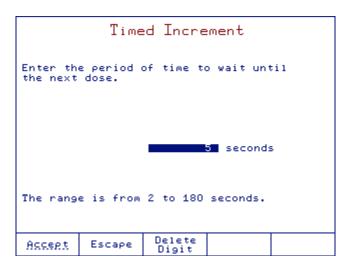
delta E:	maximum change in potential during <i>delta t</i> The range is from 0.1 to 99.9 mV.
delta t:	the time interval during which the potential is measured. The range is from 1 to 10 seconds.
t min wait:	the minimum elapsed time before a stability check. This is also the minimum elapsed time between two doses. The range is from 2 seconds to <i>t max wait</i> time.
t max wait:	the maximum elapsed time between two successive doses. If the <i>t max wait</i> has elapsed, a new dose is added even if the signal stability condition is not reached. The range is from t min wait time to 180 seconds.

5.5.14.2. TIMED INCREMENT

Option: 2 to 180 seconds

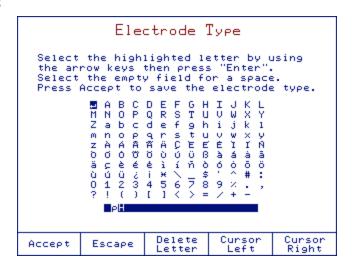
When *timed increment* is selected, the titrator acquires the potential (mV) at a fixed time interval (no signal stability check). The time period between two acquisitions must be set according to the reaction and the response time of the electrode.





5.5.15. ELECTRODE TYPE

Option: Up to 20 characters



5.5.16. BLANK OPTION

Option: Disabled, V-Blank or Blank-V

This feature allows the user to select the procedure for the blank calculations (where V is the volume of titrant dispensed during the titration and *blank* is the volume of titrant consumed by the blank sample).

Id: USER(Select	0003 M	Modify odified: (on to be (09:40 May	23, 2018
Analog Stirrer Titran Reagen Dosing End Poi Recogni Pre-Ti Measure	Board: Configuu t pump: t Addition Type: int Mode: int Mode: tration U tration So ment Mode ode Type:	ration n 1: n 2: mV 3EQ ; ions olume: tir Time:	Pu Disa Dyr Points,1st V - B1 Blank No B1a	ump 1 abled abled namic t Der ank - V
Select	Escape	Print Method	Page Up	Page Down

If one of the options (*V-Blank* or *Blank-V*) is selected in the **View / Modify Method** screen, the *blank value* will be active on the **View/Modify Method** screen and the value of the blank can be set (in liters).

				•		'	
			Bla	ank Va	alu	e	
E	nter	the bl	ank (volume	in	liters.	
				0.00	0125		
As	cept	Esca	Pe	Delete Digit			Exponent

5.5.17. CALCULATIONS

The final result is calculated using the end point volume (titrant volume at the equivalence point or at the fixed end point), and a formula selected by the user.

Calculations
Select either the calculation to be performed or modify the variables.
Edit Variable Values No Formula (mL only) No Formula (L only) Sample Calc. by Weight Sample Calc. by Volume Stdz. Titrant by Weight Stdz. Titrant by Volume Generic Formula
Select Escape

5.5.17.1. STANDARD TITRATION

5.5.17.1.1. EDIT VARIABLE VALUES

Edit the variables in a previously selected calculation.

For each formula, selected variables can be changed.

5.5.17.1.2. NO FORMULA (mL ONLY)

Only the volume of titrant (mL) required to reach the end point will be displayed.

5.5.17.1.3. NO FORMULA (L ONLY)

Only the volume of titrant (L) required to reach the end point is displayed.

TITRATION METHODS

5.5.17.1.4. SAMPLE CALCULATIONS BY WEIGHT

This calculation is used when the concentration of an analyte is determined by the weight of the sample. The results are based on the initial sample weight (in grams).

The titrator will calculate the results based on the selected units.

Titrant Units	Final Result Units
Select the titrant unit.	Select the unit for your results.
M (mol/L) N (eq/L) g/L mg/L	ppt (g/kg) ppm (mg/kg) ppb (ug/kg) X = (g/100g) mg/g mg/kg mol/kg mmol/g eq/kg
<u>Select</u> Escape	Select Escape

The titrator will provide the results based on the titrant and sample units selected. Titrant Units:

 $M \pmod{L}$ moles/liter equivalents/liter N (eq/L)g/L grams/liter mg/L milligrams/liter Final Result Units: parts per thousand (grams/kilogram) ppt (g/kg) ppm (mg/kg) parts per million (milligrams/kilogram) ppb (µg/kg) parts per billion (micrograms/kilogram) % (g/100 g) percentage in weight (grams/100 grams) milligrams/gram mg/g milligrams/kilogram mg/kg mol/kg moles/kilogram mmol/g millimoles/gram eq/kg equivalents/kilogram milliequivalents/kilogram meq/kg

A formula example is shown below using M (mol/L) as the titrant unit and ppt (g/kg) as the final result unit:

Calculating Sample Concentration
M (mol/L)> ppt (g/kg)
The calculation is:
$\frac{\frac{V \times m01}{L} \times m01}{9 \times \frac{k9}{10009}}$
Select the variables to change value. V = volume dispensed in liters.
1.000 mol/L -> titrant conc. 1.000 mol/mol -> (sample/titrant) 1.000 g/mol -> mw of sample 1.000 g -> sample weight
Select Escape Save / Exit

Variables can be set according to the amount of sample and titrant used.

5.5.17.1.5. SAMPLE CALCULATIONS BY VOLUME

This calculation is used when the concentration of an analyte is determined in terms of the volume of sample. The results are based on the initial sample volume (in milliliters).

The titrator will calculate the results based on the selected units.

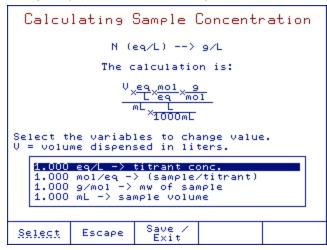
Titrant Units	Final Result Units
Select the titrant unit.	Select the unit for your results.
M (mol/L) N (eq/L) g/L mg/L	<pre>% = (g/100g) mg/g mg/kg mo1/kg mmo1/g eq/kg meq/kg ppt (g/L) ppm (mg/L) ppb (µg/L) M (mo1/L) N (eq/L) g/L</pre>
Select Escape	Select Escape Page Page Down

Titrant Units:

M (mol/L)	moles/liter
N (eq/L)	equivalents/liter
g/L	grams/liter
mg/L	milligrams/liter

Final Result Units:	
ppt (g/L)	parts per thousand (grams/liter)
ppm (mg/L)	parts per million (milligrams/liter)
ppb (µg/L)	parts per billion (micrograms/liter)
M (mol/L)	Molarity (moles/liter)
N (eq/L)	Normality (equivalents/liter)
mg/L	milligrams/liter
μg/L	micrograms/liter
mmol/L	millimoles/liter
mg/mL	milligrams/milliliter
mg/100 mL	milligrams/100 milliliters
g/100 mL	grams/100 milliliters
eq/L	equivalents/liter
meq/L	milliequivalents/liter

A formula example is shown below using N (eq/L) as the titrant units and g/L as the final result units:



Variables can be set according to the amount of sample and titrant used.

5.5.17.1.6. STANDARDIZE TITRANT BY WEIGHT

This calculation is used when the concentration of the titrant is determined using a solid standard. Determination of the titrant concentration is based on the primary standard weight (in grams).

The calculation is based on the selected titrant unit. If the titrant unit is M (mol/L), the formula used to calculate the result is displayed below:

Titrant 1 Units	Calculating Titrant Concentration
Select the titrant 1 unit.	The titrant concentration unit is M (mol/L).
M (mo1/L)	The calculation is:
N (eq/L) g/L mg/L	$\frac{9 \times \frac{mo1}{9} \times \frac{mo1}{mo1}}{V}$
	Select the variables to change value. V = volume dispensed in liters.
	D.200 g → standard weight 204.23 g/mol → mw of standard 1.000 mol/mol → (titrant/standard)
Select Escape	Select Escape Save / Exit

5.5.17.1.7. STANDARDIZE TITRANT BY VOLUME

This calculation is used when the concentration of the titrant is determined using a primary standard solution. Determination of the titrant concentration is based on the primary standard volume (in milliliters).

The calculation is based on the selected titrant unit. If the titrant unit is N (eq/L), the formula used to calculate the result is displayed below:

Titrant Units	Calculating Titrant Concentration
Select the titrant unit.	The titrant concentration unit is N (eq/L).
M (mo1/L)	The calculation is:
N (eq/L) g/L mg/L	^{ML} × <u>L</u> ×≊۹ 1000mL× ^E L V
	Select the variables to change value. V = volume dispensed in liters.
	1.684 mL -> standard volume 2.375 eq/L -> standard conc.
Select Escape	Select Escape Save / Exit

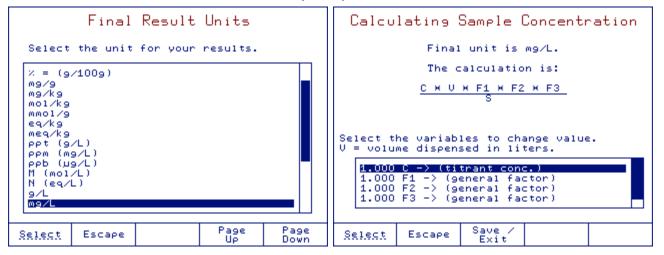
5.5.17.1.8. GENERIC FORMULA

The user can define their own calculation formula based on the final result units in a solid or liquid sample. Final Result Units:

ppt (g/kg)	parts per thousand (grams/kilogram)
ppt (g/L)	parts per thousand (grams/liter)
ppm (mg/kg)	parts per million (milligrams/kilogram)
ppm (mg/L)	parts per million (milligrams/liter)
ppb (µg/kg)	parts per billion (micrograms/kilogram)
ppb (µg/L)	parts per billion (micrograms/liter)
% (g/100 g)	percentage in weight (grams/100 grams)
M (mol/L)	Molarity (moles/liter)
mg/g	milligrams/gram
N (eq/L)	Normality (equivalents/liter)
g/L	gram/liter
mg/kg	milligrams/kilogram
mg/L	milligrams/liter
mol/kg	moles/kilogram
μ g/L	micrograms/liter
mol/L	moles/liter
mmol/g	millimoles/gram
eq/kg	equivalents/kilogram
mmol/L	millimoles/liter
meq/kg	milliequivalents/kilogram
mg/mL	milligrams/milliliter
mg/100 mL	milligrams/100 milliliters
g/100 mL	grams/100 milliliters
eq/L	equivalents/liter
meq/L	milliequivalents/liter
No Unit	No result unit

The titrator will calculate the results based on the selected unit.

The formula can be either for titrant standardization or sample analysis.



TITRATION METHODS

Where:

- C = the concentration of the titrant
- F1 = general factor
- F2 = general factor
- F3= general factor
- S = sample size, in grams or milliliters

V = the volume delivered, in liters, to reach the preset or equivalence end point (determined by the titrator) General factors:

Weight Conversion:

One of the general factors should be a weight conversion factor.

Examples of concentration units:

mol/L	moles/Liter
eq/L	equivalents/Liter
g/L	grams/Liter
mg/L	milligram/Liter

Reaction Ratio:

The reaction ratio is the ratio between the analyte and titrant or standard and titrant.

Examples of ratios:

mol/mol	moles of sample/moles of titrant
mol/eq	moles of sample/equivalents of titrant
eq/mol	equivalents of sample/moles of titrant
mol/mol	moles of titrant/moles of standard
eq/mol	equivalents of titrant/moles of standard
male 2 males of NaOH	react with 1 male of H SO

Example: 2 moles of NaOH react with 1 mole of $\mathrm{H_2SO_4}$

Unit Conversion factor:

Used to convert between various measurement units. Examples: L/1000 - mL

g/1000 —> mg

Weight Conversion factor:

Used to convert between weight measurement bases (kg, g, mg, μ g, mole or mmole). Example: g —> mol

5.5.17.2. BACK TITRATIONS

Calculations				
Select either the calculation to be performed or modify the variables.				
Sample Calc. by Weight Sample Calc. by Volume Generic Formula				
		•		
Select	Escape			

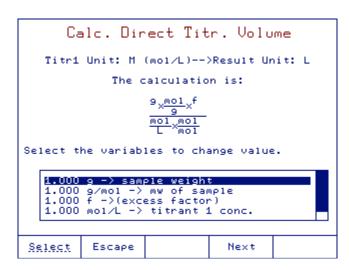
5.5.17.2.1. SAMPLE CALCULATIONS BY WEIGHT

Select the titrant 1 unit, titrant 2 unit, and final result unit.

Titrant 1 Units	Titrant 2 Units				
Select the titrant 1 unit.	Select the titrant 2 unit.				
H (mol/L) N (eq/L) g/L mg/L	M (mol/L) N (eq/L) g/L mg/L				
<u>Select</u> Escape	Select Escape				

Final Result Units	
Select the unit for your results.	
<pre>ppt (g/kg) ppm (mg/kg) ppb (µg/kg) % = (g/100g) mg/kg mo1/kg mmo1/g eq/kg meq/kg</pre>	
Select Escape	

A formula example is shown below using M (mol/L) as the titrant 1 units, M (mol/L) as the titrant 2 units, mg/g and the final result units. This formula is used to calculate the amount of titrant 1 to dispense:



The formula is based on the assumption that the sample concentration is 100% w/w.

The titrator will calculate the volume of titrant 1 needed to consume the sample and multiply it with the excess factor in order to raise or lower the amount of titrant 1 dispensed.

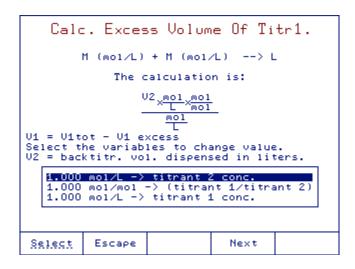
Variables can be set according to the amount of sample and titrant used.

Press Next to proceed to the next formula.

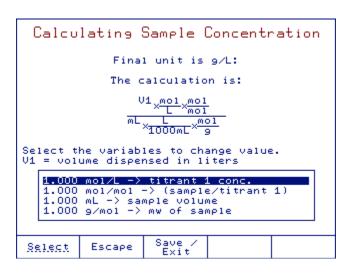
If you do not want the titrator to calculate the volume of titrant 1 to add, see Titrant 1 Entry section.

The remaining volume of titrant 1 needs to be calculated.

The following formula is used to calculate the remaining volume of titrant 1 after the reaction with the sample:



When all of the variables are set, press Next to proceed with the "Calculating Sample Concentration" formula:

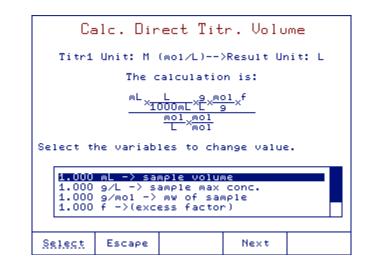


5.5.17.2.2. SAMPLE CALCULATIONS BY VOLUME

Select the titrant 1 unit, titrant 2 unit, and the final result unit.

	Titr	rant 1 l	Jnits	Б				[itr	ant 2 l	Jnits	
Select <mark>M (mol</mark> N (eq/ 9/L mg/L	the titr /L)	ant 1 uni		>		Selec N (eq 9/L m9/L	t the 1/L)		nt 2 uni		
Select	Escape		elect pt (9 pm (m pb ()) (mol (eq) 9/L 9/L 9/L 9/L 9/L 9/L 100	the un /L) 9/L) 9/L) /L) L)		Select Sult Un: Your rest	its	аре			
			q∕L lect	Escap	e	P	age Jp	Pag			

After you have selected the titrant 1, titrant 2, and the final result units, the titrator will display a screen with a formula used to calculate the amount of titrant 1 (used in the first stage of back titration) to be dispensed.

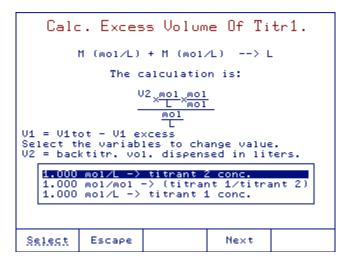


The formula is based on the assumption that the sample concentration is 100% v/v. The titrator will calculate the volume of titrant 1 needed to consume the sample and multiply it with the excess factor in order to raise or lower the amount of titrant 1 dispensed. Variables can be set according to the amount of sample and titrant used.

Press Next to proceed to the next formula.

If you do not want the titrator to calculate the volume of titrant 1 to add, see Titrant 1 Entry section.

The following formula is used to calculate the remaining volume of titrant 1 after the reaction with the sample.



When all the variables are set, press vext to proceed with the "Calculating Sample Concentration" formula:

Calculating Sample Concentration					
Final unit is g/L:					
The calculation is:					
V1 × mo1 mL × mo1 mL × mo1 1000mL × 9					
Select the variables to change value. V1 = volume dispensed in liters					
1.000 mol/L -> titrant 1 conc. 1.000 mol/mol -> (sample/titrant 1) 1.000 mL -> sample volume 1.000 g/mol -> mw of sample					
Select Escape Save / Exit					

TITRATION METHODS

5.5.17.2.3. GENERIC FORMULA

The user can define their calculation formula for the "Direct Titration Volume", "Calculating Excess Volume of Titrant 1" and "Final Sample Concentration" in a solid or liquid sample.

5.5.18. DILUTION OPTION

Option: Enabled or Disabled

When the initial sample is diluted, a titration is made with an aliquot of the diluted sample, dilution calculations can be used. The calculations are based on the original sample weight (volume) in order to express the results for the initial sample.

Dilution Parameters					
Select	the option				
	Dilution Vo	lume:	100.000 mL	1	
	t Volume: e size to b	e diluted	10.000 mL 1.0000 mL		
Select	Escape				

Final Dilution Volume: Aliquot Volume: Analyte size to be diluted: The volume of the sample after dilution Volume of sample taken from the dilution for titration The initial sample weight (volume)

5.5.19. TITRANT NAME

Option: Up to 15 characters

Titrant1 Name using the highlighted letter Ъy arrow keys then press y field for "Enter". a space. ect the Accep to save entered text. C P AN a n A O BODOAO DQd E R e FSfSCO GT 9t EÜ HULUEBÒ\$ IJ KXk×1aō# MZ mzbäù0? J⊌jwi Ļ Y è i 1 9 # 0 ĕ Ň P A U n Ä Ù á è ü 2 é ė ì ñ ò ö 9 0 1 5 ŝ 1 Delete Letter Cursor Left Cursor Right Accept Escape

Note: For back titrations the name of titrant 1 and titrant 2 can be entered.

TITRATION METHODS

5.5.20. TITRANT CONCENTRATION

Enter the concentration of the titrant to be used. When determining the titrant concentration, only the concentration unit is displayed.

Titrant1 Conc.				
Enter the titrant 1 concentration.				
		0.106	7 <mark>6 </mark> M (mo	(17L)
Accept	Escape	Delete Digit		Exponent

Note: For back titration the concentration of titrant 1 and titrant 2 can be entered.

5.5.21. ANALYTE SIZE

Option: 0.001 to 250.0

Enter the size of the sample (for sample concentration determinations) or standard (for titrant concentration determination).

	Sam	nele Vol	ume	
Enter millil	the initi: iters.	al sample	volume ir	1
		1.000	U n L	
	olume wil: size is s		when fixe	≥d
Accept	Escape	Delete Digit		Exponent

5.5.22. ANALYTE ENTRY

Option: Fixed, Manual or Same as Previous (linked methods only)

Analyte Entry	Analyte Entry
Select the entry mode of analyte.	Select the entry mode of analyte.
Fixed Weight or Volume Manual Weight or Volume Verify the correct formula is being used, i.e. weight or volume analyte type.	Fixed Weight or Volume Manual Weight or Volume Same as Previous Verify the correct formula is being used, i.e. weight or volume analyte type.
Select Escape	Select Escape

5.5.22.1. FIXED WEIGHT OR VOLUME

Each titration will use a set weight or volume in the calculations.

5.5.22.2. MANUAL WEIGHT OR VOLUME

Each titration, the exact weight or volume can be entered. The titrator will prompt for the analyte weight or volume at the beginning of each titration.

5.5.22.3. SAME AS PREVIOUS (LINKED METHOD ONLY)

The same weight or volume is used for both methods.

5.5.23. TITRANT 1. ENTRY (BACK TITRATION ONLY)

Select the mode for evaluating the necessary quantity of titrant 1 used in the back titration process (phase 1).

	Titr	ant 1 E	intry	
Select	the entry	mode of	titrant 1.	
<mark>Calcul</mark> a Fixed B	i <mark>ted By Fo</mark> r Wy User	rmula		
Select	Escape			

5.5.23.1. CALCULATED BY FORMULA

The volume of titrant 1 to be dispensed in the phase 1 of back titration will be calculated by the titrator (see **Back Titrations** section).

5.5.23.2. FIXED BY USER

A fixed volume of titrant 1 will be used during the first phase of back titration process (direct titration).

Direct Titration Volume				
	the volume pensed dur			
		10.00	J mL	
This volume will be dispensed when Fixed By User option is selected.				
Accept	Escape	Delete Digit		

5.5.24. MAXIMUM TITRANT VOLUME

Option: 0.100 to 100.000 mL

The maximum titrant volume used in the titration must be set according to the analysis.

If the titration end point (fixed or equivalence End Point) is not reached, the titration will be terminated after the maximum titrant volume has been dispensed. The error message ("Limits Exceeded") will appear on the display.

Maximum Titrant Volume					
Enter dispen:		um titran [.]	t volume to	be	
		15.00	u mL		
Recommend the total volume of the burette.					
Accept	Escape	Delete Digit			

5.5.25. POTENTIAL RANGE

Option: -2000.0 to 2000.0 mV

The input potential range can be set by the user. The titration will be terminated and an error message will appear if the potential is outside these limits.

These limits provide protection against a titration that does not generate an end point due to potential over-range.

Potential Range				
Enter	the upper	and lower	- potentia	al.
	2000.0	mV - Upp	er Limit	
-2000.0 mV - Lower Limit				
Press Next to move to the next entry.				
Accept	Escape	Delete Digit	Next	

5.5.26. VOLUME/FLOW RATE

The flow rate for the dosing system can be set by the user in an interval of 0.3 to two times the burette volume:

- 0.3 to 10 mL/min for a 5 mL burette
- 0.3 to 20 mL/min for a 10 mL burette
- 0.3 to 50 mL/min for a 25 mL burette
- 0.3 to 100 mL/min for a 50 mL burette

Flow Rate					
Enter	the titra	nt/reagen:	t flow ra	te.	
		50.0	💵 mL/min		
The range is from 0.1 to twice the total volume of the burette.					
Accept	Escape	Delete Digit			

Note: The titrator will automatically detect the burette size and display the correct high limit volume. The flow rate is set for all burette operations.

5.5.27. SIGNAL AVERAGING

Option: 1,2,3 or 4 readings

This option enables filtering on the mV/pH reading.

If 1 Reading is selected, the filtering is disabled. The titrator will take the last reading and place it into a "moving window" along with the last 2, 3 or 4 readings (depending on the selected option). The average of those readings is displayed and used for calculations. Averaging more readings is helpful when a noisy signal is received from the electrode.

	View/ 0001 M the optic		15:58 Jun	28, 2018
Blank Calcul Diluti Titran Analyt Analyt Maximu Potent Volume	ode Type: Option: ations: on Option t Name: t Conc.: e Size: e Entry: m Titrant ial Range ∕Flow Rat	: 0. Volume: : -2000.(e: 25 mL	alc. by Vo Disa 0.1N 1 Readi 2 Readi 3 Readi 4 Readi	abled NaOH ngs ngs ngs
Signal Averaging: 1 Reading Significant Figures: XXXXX Linked To: No Link				XXXXX
Select	Escape	Print Method	Page Up	Page Down

5.5.28. SIGNIFICANT FIGURES

Option: Two (XX), Three (XXX), Four (XXXX) or Five (XXXXX)

This option allows you to set the format for displaying the final titration result.

	0005 M	Modify odified: :	10:18 May	23, 2018
Pre-Ti Measurd Electr Blank (Calcul Diluti Titran Analyt Analyt Maximu Potent Volume.	tration S ement Mod ode Type: Option: ations: S ton Option t Name: e Size: e Entry: m Titrant ial Range	Volume: : -2000.(e: 25 mL	anal Stabi No E Disa 0.1N 0.1N 0.1X 15 XX 0 to 20 XX	1ity PH Blank Sight bled NaOH X XX
Signif	icant Fig	ures:		
Select	Escape	Print Method	Page Up	Page Down

5.5.29. LINKED METHOD

This option allows the user to link two titration methods. If No Link is selected, only the current method will run. If a method is selected, it will run after the current method.

Link Titration Method				
Select the method to be linked.				
No Link USER0001 Acidity of Water				
USER0009 Free acidity USER0010 Wine acidity				
<u>Select</u> Escape				

TITRATION METHODS

5.5.30. START LINKED METHOD (LINKED METHOD ONLY)

Option: Manually or Automatically

Selecting Manually will stop the titration temporarily between the methods. This break allows you to perform a task related to the analysis (e.g.: boiling the sample to remove carbon dioxide).

Start Linked Method				
Select the start linked method mode.				
Automatically Manually				
Select Escape				

5.6. PRINTING

To print method parameters, press Method Options from the main screen.

Press Print and wait a few seconds until the printer completes the job.

If no printer is connected to the dedicated socket, or if the printer is offline, an error message will appear on the display (see **Connecting a Printer** section, for information about connecting a printer to the titrator).

CHAPTER 6. TITRATION/DIRECT READING MODE

6.1. RUNNING A TITRATION/DIRECT READING	
6.1.1. STARTING A TITRATION/DIRECT READING	
6.1.2. SUSPENDING A TITRATION/DIRECT READING	
6.1.3. VIEWING THE TITRATION CURVE	
6.2. STOPPING A TITRATION/DIRECT READING	

TITRATION MODE

6.1. RUNNING A TITRATION/DIRECT READING

Before beginning a titration, make sure that the following conditions are met:

- At least one pump is properly installed.
- A burette is inserted in the pump and filled with titrant.
- The aspiration tube is inserted in the titrant bottle and primed. The dispensing tube is over the titration beaker.
- The tubes are installed on the peristaltic pump and filled with reagent.
- The standard or sample has been carefully weighed/measured into the beaker.
- The electrode(s) and the temperature probe are submersed in the beaker.
- The desired method is selected and the parameters are set to the optimal values.

6.1.1. STARTING A TITRATION/DIRECT READING

To start a new analysis, press stop from the main screen. When an analysis begins:

- The stirrer will turn on (if enabled).
- The pre-titration stir time starts, if enabled (see Titration Methods section).
- After the pre-titration stir time is complete the pre-titration volume will be displayed, if enabled. (see **Titration Methods** section).
- If *Reagent Addition* is enabled the quantity of reagent will be entered according to the parameters set in the method.
- The titrator will start the analysis and continue to deliver titrant until the end point is detected or the titration is terminated.

6.1.2. SUSPENDING A TITRATION/DIRECT READING

While a titration/analysis is in progress, you can temporarily stop it by pressing Suspend. This will stop the dosing pump if it is running.

To continue the titration/analysis press Resume

6.1.3. VIEWING THE TITRATION CURVE

During a titration, the potentiometric curve and the derivative curve (equivalence point only) can be displayed on the **Titration Graph** screen by pressing View Curve.

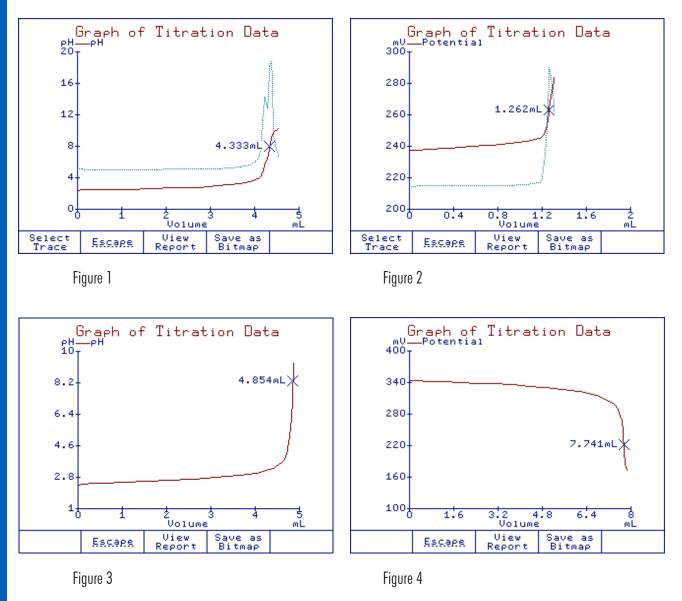
The potentiometric curve and the derivative curve are scaled to fit simultaneously inside the display.

When a titration end point is successfully detected, the volume is displayed on the graph and marked with an "x".

TITRATION MODE

The contents of the graph as related to an end point type are as follows:

Equivalence End Point (pH): The pH readings and the selected derivative vs. volume of titrant are displayed (see Figure 1).
Equivalence End Point (mV): The mV readings and the selected derivative vs. volume of titrant are displayed (see Figure 2).
Fixed End Point (pH): The pH readings vs. volume of titrant are displayed (see Figure 3).
Fixed End Point (mV): The mV readings vs. volume of titrant are displayed (see Figure 4).



Select Trace - Changes the y-axis scale to either the mV (or pH) readings or the selected derivative values (of mV or pH). Available only for titrations with equivalence end points.

Save as Bitmap - Allows you to save the graph as a bitmap file. Available only when the titration is finished.

TITRATION MODE

6.2. STOPPING A TITRATION/DIRECT READING

The titration/analysis is terminated when one of the following conditions is met:

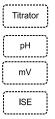
- Analysis Completed: This is the only mode with valid final result values. The end point/stable reading was successfully detected, the final results will be displayed.
- Manually Terminated: The current titration/analysis terminated by the user before the end point was detected.
- Limits Exceeded: The maximum titrant volume was delivered without reaching the end point. An error message is displayed on the screen.
- Critical Error: A critical error occurred and the titration was stopped. These errors are typically related to the dosing system. An error message is displayed on the screen.
- **Potential Out of Range:** The measured values from the electrode are outside the potential range. An error message is displayed on the screen.

CHAPTER 7. pH MODE

7.1. DISPLAY
7.2. pH SETUP
7.2.1. BUFFER ENTRY TYPE
7.2.2. FIRST CALIBRATION POINT
7.2.3. EDIT CUSTOM BUFFERS
7.2.4. EDIT BUFFER GROUP
7.2.5. CALIBRATION REMINDER
7.2.6. SET REMINDER PERIOD
7.2.7. CLEAR CALIBRATION
7.2.8. pH GLP DATA
7.2.9. LOGGING INTERVAL
7.2.10. STABILITY CRITERIA
7.2.11. pH RESOLUTION
7.2.12. STIRRER CONFIGURATION
7.2.13. STIRRING SPEED
7.3. pH CALIBRATION
7.4. LOGGING
7.4.1. INTERVAL LOGGING
7.4.2. MANUAL LOGGING

By pressing Mode from the main screen, the titrator can be switched to **Titrator**, **pH**, **mV** or **ISE** modes. **One Analog Board**

Wa	orking M	ode	
Select the worl	king mode.		
<u>Titrator</u>	ρH	mV	ISE



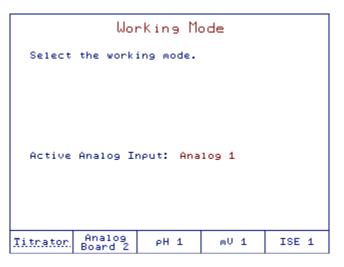
Switches to Titrator mode.

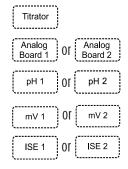
Switches to **pH** mode.

Switches to **mV** mode.

Switches to **ISE** mode.

Two Analog Boards





Switches to **Titrator** mode.

Switches the Analog Input for **pH**, **mV** and **ISE** mode.

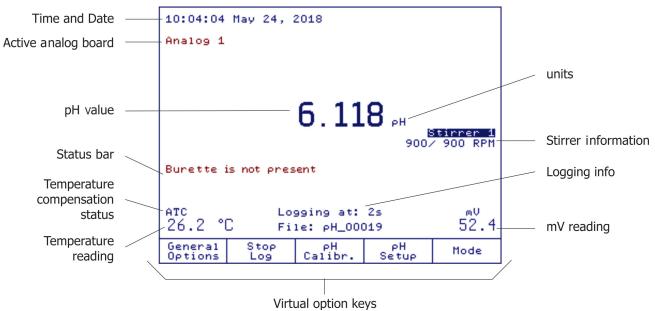
Switches to $\mathbf{p}\mathbf{H}$ mode.

Switches to \boldsymbol{mV} mode.

Switches to $\ensuremath{\mathsf{ISE}}$ mode.

7.1. DISPLAY

pH MODE



pH Mode Option keys:

General Options screen gives access to options that are not directly related to the measurement process (see General Options section).



Stores the current pH reading (see Manual Logging section).

Starts the interval log (see Automatic Logging section).

Enter the pH calibration screen (see pH Calibration section).

Enter the pH setup screen, parameters are associated with pH measurements and calibration (see **pH Setup** section). Allow the user to switch between the available measurement modes: **Titrator**, **pH**, **mV** or **ISE** mode.

7.2. pH SETUP

To access pH Setup, press pH Setup option key while in pH mode.

Select a menu option.	
Buffer Entry Type:	Manual
First Cal Point: Edit Custom Buffers Edit Buffer Group	Point
Calibration Reminder:	Periodio
Set Reminder Period: Clear Calibration pH GLP Data	10d:02h:30≬
Logging Interval:	0h:00m:02s
Stability Criteria: pH Resolution:	Medium X.XXX
Stirrer Configuration: Stirring Speed:	Stirrer 1 1200 RPN

Use \bigwedge and \bigvee keys to highlight the desired option.

Press select or enter to access the selected option.

7.2.1. BUFFER ENTRY TYPE

Option: Automatic, Semiautomatic or Manual

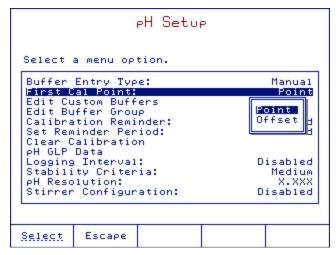
PH S	ietup
Select a menu option.	
Buffer Entry Type:	Manual
First Cal Point: Edit Custom Buffers Edit Buffer Group Calibration Reminder Set Reminder Period: Clear Calibration pH GLP Data Logging Interval: Stability Criteria: pH Resolution: Stirrer Configuration	Disabled Medium X.XXX
<u>Select</u> Escape	

Automatic: The instrument automatically selects the pH calibration point as the closest buffer from the predefined buffer group (see **Edit Buffer Group** section).

Semiautomatic: The instrument automatically selects the closest buffer from the available buffers (standard and custom buffers). Manual: The calibration buffer must be manually selected by the user during calibration from the available buffer list (standard and custom buffers).

7.2.2. FIRST CALIBRATION POINT

Option: Point or Offset



If *Point* is selected, the slope values adjacent to the calibration points will be reevaluated (normal calibration). If *Offset* is selected the existing slope values will not be changed.

7.2.3. EDIT CUSTOM BUFFERS

If you wish to use buffers other than the standard ones, use the *Edit Custom Buffers* option to set the desired pH value, up to five pH custom buffers can be set.

Note: Custom buffers are not temperature compensated, enter the value of the buffer at the calibration temperature.

	Edit C	Custom B	luffers	
Press	<edit> to</edit>	edit sele	ected buff	er.
	(Remove B) stom buff)	uffer> to er.	delete	
Cust 6.870 Use ar	9.230	to select	Cust	Cust
Remove Buffer	Escape	Edit	\triangleleft	\triangleright

- Use \lt and \triangleright keys to select the desired buffer.
- Press $\left[\begin{array}{c} {}^{\text{Remove}} \\ {}^{\text{Buffer}} \end{array}
 ight]$ to delete the selected buffer.
- Press Edit to edit the selected buffer.

	Edit C	Custom B	offers	
Enter	the custo	m buffer (value.	
		9.23	О П РН	
Cust 6.870		Cust 12.750	Cust	Cust
Low lin High 1:	nit: -2 imit: 20			
Accept	Escape	Delete Digit		

- Use the numeric keypad to enter the pH buffer value.
- Press Accept to save the value.
- Press Escape to return to pH Setup menu.

7.2.4. EDIT BUFFER GROUP

Option: Up to five

Select up to five buffers from the available buffers (Hanna or Custom) to be used for automatic buffer recognition.

Within the buffer group, pH values must be at least 1.5 pH far apart.

If the buffer group already contains five pH buffers, at least one pH buffer has to be removed in order to add another buffer.

	Edit	Buffer	Group	
Availa	ble Buffe	rs		
Hanna 1.679 Hanna 9.177	Hanna 3.000 Hanna 10.010	Hanna 4.010 Hanna 12.450		Hanna 7.010
Buff	er Group			
Hanna 1.679	Hanna 4.010	Hanna 6.862		Hanna 2.450
Remove	Escape	\triangleright	Δ	∇

- Use the arrow keys to select the pH buffer to be included/removed in/from the buffer group.
- Press Add or Remove to add/remove the selected pH buffer to/from buffer group.
- Press Escape to return to pH Setup menu.

7.2.5. CALIBRATION REMINDER

Option: Daily, Periodic or Disabled

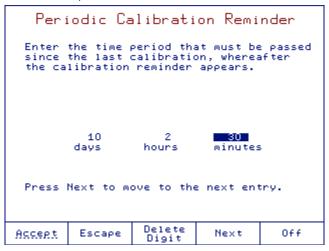
	Calibration	Reminder	
Select	a menu option.		
Daily Period Disabl			
_	_		
Select	Escape		

- Daily: The calibration reminder will appear daily at a specified time.
- Periodic: The calibration reminder will appear after the set time has elapsed since the last calibration.
- Disabled: The calibration reminder will not appear.

7.2.6. SET REMINDER PERIOD

If *Daily* or *Periodic* option was selected for the calibration reminder, the reminder period must also be set. For a daily reminder period, the time of day can be set.

For a periodic reminder period, the number of days, hours and minutes can be set.



- Press Next to move the cursor to the next field.
- Press Accept to save the changes or Escape to return to the previous screen.
- Press Or to disable the calibration reminder and return to pH setup.

7.2.7. CLEAR CALIBRATION

This option clears the existing pH calibration for the selected channel. If the calibration is cleared, the factory calibration will be used.

• Press Clear to clear the previous calibration or Escape to return to the previous screen without clearing the calibration.

	Clear	Calibr	ration	
Press points	(Clear) to •	o clear a	ll calibr	ration
	(Escape) libration		without	clearing
<u>Clear</u>	Escape			

7.2.8. pH GLP DATA

Displays the pH calibration data.

PH GLP Data
Analog 1 Last Calibration: 10:13 May 24, 2018 Offset: -0.1 mV Average Slope: 100.7%
1.679рН (Hanna) 316.2mV 26.3°С А 10:10:30 Мау 24, 2018
4.010pH (Hanna) 177.5mV 26.3°C A 10:09:11 May 24, 2018
7.010pH (Hanna) -0.6mV 26.3°C A 10:08:40 May 24, 2018
10.010pH (Hanna) -179.1mV 26.3°С А 10:09:43 Мау 24, 2018
12.450pH (Hanna) -325.6mV 26.3°C A 10:13:15 May 24, 2018
Escars

7.2.9. LOGGING INTERVAL

Option: 2 seconds to 8h 59 min 59 sec

Set the logging interval to be used for automatic logging.

Logging Interval				
Enter	the data (logging ir	nterval.	
	0 hours	0 minutes	2 seconds	
Press Next to move to the next entry.				
Accept	Escape	Delete Digit	Next	Off

7.2.10. STABILITY CRITERIA

Option: Fast, Medium, Accurate

PH Setup	
Select a menu option.	
Buffer Entry Type: First Cal Point: Edit Custom Buffers Edit Buffer Group Calibration Reminder: Set Reminder Period: Clear Calibration pH GLP Data Logging Interval:	Manual Point Fast Miedium Accurate
Stability Criteria: pH Resolution: Stirrer Configuration:	Medium X.XXX Disabled
Select Escape	

Select the signal stability criteria:

Fast: Quicker results with less accuracy

Medium: Medium speed results with medium accuracy

Accurate: Slower results with high accuracy

pH MODE

7.2.11. pH RESOLUTION

Option: One (X.X), Two (X.XX) or Three (X.XXX) decimal places

et	Setup
Select a menu opti	n.
Buffer Entry Type: First Cal Point: Edit Custom Buffer Edit Buffer Group	Manual Point
Calibration Remind Set Reminder Perio Clear Calibration pH GLP Data Logging Interval:	
Stability Criteria PH Resolution: Stirrer Configurat	×.×××

7.2.12. STIRRER CONFIGURATION

Option: Stirrer 1, Stirrer 2 (if available) or Disabled

PH Setur	
Select a menu option.	
Buffer Entry Type:	Manual
First Cal Point: Edit Custom Buffers Edit Buffer Group	Point
Calibration Reminder:	Periodic
Set Reminder Period:	10d:02h:30m
Clear Calibration PH GLP Data	Disabled
Logging Interval:	Stirrer 1
Stability Criteria:	Stirrer 2 m
pH Resolution: Stirrer Configuration:	Disabled
Select Escape	

7.2.13. STIRRING SPEED

Option: 200 to 2500 RPM

	Stirring Speed				
Enter below		of the st	tirrer wit	hin:	
		110	U RPM		
The range is from 200 to 2500 RPM.					
Accept	Escape	Delete Digit			

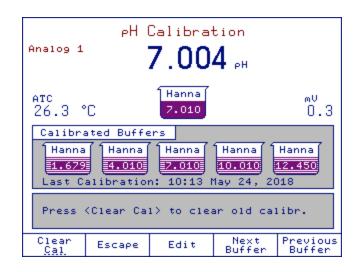
pH MODE

7.3. pH CALIBRATION

Calibrate the instrument often, especially if high accuracy is required.

The instrument should be recalibrated:

- Whenever the pH electrode is replaced.
- At least once a week.
- After testing aggressive chemicals.
- When "No pH Calibration" or "pH Calibration Expired" message appears on the display.



PREPARATION

Pour small quantities of the buffer solutions into clean beakers. If possible, use plastic beakers to minimize any EMC interferences. For accurate calibration and to minimize cross-contamination, use two beakers for each buffer solution: one for rinsing the electrode and one for calibration. If you are measuring in the acidic range, use pH 7.01 or 6.86 as the first buffer and pH 4.01/3.00 or 1.68 as the second buffer. If you are measuring in the alkaline range, use pH 7.01 or 6.86 as the first buffer and pH 10.01/9.18 or 12.45 as the second buffer.

For extended range measurements (acidic and alkaline), perform a five-point calibration by selecting five buffers across the entire pH range.

CALIBRATION PROCEDURE

During calibration, the user has a choice of 8 standard buffers: (pH 1.68, 3.00, 4.01, 6.86, 7.01, 9.18, 10.01, 12.45) and up to 5 custom buffers.

For accurate measurements it is recommended to perform a five-point calibration. However, at least a two-point calibration is suggested. For pH titrations, the selected buffers should bracket your end point (e.g.: if your end point value is at 8.5, use 7.01 or 6.86 and 9.18 or 10.01 for calibration).

To begin calibration:

• Press Press I the instrument was calibrated before the old calibration can be cleared by pressing Clear Cal

Note: It is very important to clear calibration history when a new electrode is used.

- Immerse the pH electrode and the temperature probe approximately 4 cm (1.5") into a buffer solution and stir gently.
- If necessary, select the pH calibration buffer value with Next Buffer or Previous Buffer.
- Once the reading has stabilized press Accept to update the calibration. The calibration buffer will be added to the Calibrated Buffers section.
- Rinse the pH electrode and the temperature probe, then immerse them into the next buffer solution and follow the above procedure or press Escape to exit the calibration.

Notes:

- The new calibration points will replace old ones if the difference between them is ± 0.2 pH.
- Buffers used in older calibrations will not have a solid background.
- If calibrating with a standard buffer in MTC mode, the pH value and temperature can be modified by pressing [Edit]. The values can be adjusted using the numeric keys. Press Accept to save the new values.

Manual Edit					
Edit p	H buffer -	and manual	temperat	ture.	
Buffer: 7.010 pH					
Temperature: 25.0 °C					
Low limit: 6.990 pH High limit: 7.030 pH					
Press	Next to m	ove to the	e next ent	try.	
Accept	Escape	Delete Digit	Next		
. ,			,		

- In ATC mode, the pH value for custom buffers can be modified by pressing Edit.
- If the Automatic Buffer entry type was selected for the calibration procedure, the titrator will automatically select the closest buffer to the measured pH value from the buffer group.
- If the Semiautomatic Buffer entry type was selected use the Previous or Next Buffer or Buffer. Only buffers in the buffer group will be displayed.

CALIBRATION MESSAGES:

- Wrong Buffer. Please check the buffer: This message appears when the difference between the pH reading and the value of the selected calibration buffer is significant. If the message is displayed, check if you have selected the appropriate calibration buffer.
- Wrong buffer temperature: This message appears if the buffer temperature is out of the defined temperature range.
- Clean the electrode or check the buffer. Press Accept to update calibration: This message alerts the user that some dirt or deposits could be on the electrode, or the buffer is contaminated.
- Slope too low. Please check the buffer: This message appears if the current slope is under 80% of the default slope. Recalibrate the instrument using fresh buffers.
- Slope too high. Press to clear the old calibration: This message appears as a result of an erroneous slope condition.

pH MODE

7.4. LOGGING

Data logging is available in pH mode. It can be logging on demand (Manual Logging) or automatically (Interval Logging) at predefined time intervals.

To customize the logging report:

- Press results to display the Data Parameters screen.
- Highlight the Setup pH/mV/ISE Report option and press select to display the Setup pH/mV/ISE Report screen.

Setup pH/mV/ISE Report
Select fields to be saved in the report.
<pre>* Result and Units * Potential * Temperature and Units * Date and Time Calibration Data Sample Name Company Name Operator Name Electrode Name Field 1 Field 2 Field 3 Software Versions Serial Numbers</pre>
Escape Save Report

- Use the A and V keys to highlight the data field that you want to show/hide in the pH/mV/ISE report and then press
- Each field marked by "*" is an active field selected for the report.
- Press Save Report to save the customized report.

7.4.1. INTERVAL LOGGING

The logging interval is set in the pH setup screen.

 $\Press \left[\begin{array}{c} Start\\ Log \end{array} \right] to start the log.$

The logging interval and name of logging file will be displayed on the **pH measurement** screen.

To stop the automatic logging, press stop.

7.4.2. MANUAL LOGGING

To manually log pH readings, press Save Reading from the **pH measurement** screen. A new record will be added to the report every time Reading is pressed.

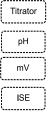
CHAPTER 8. mV MODE

8.1. DISPLAY	8-4
8.2. mV SETUP	
8.2.1. CLEAR RELATIVE mV OFFSET	
8.2.2. LOGGING INTERVAL	
8.2.3. STABILITY CRITERIA	
8.2.4. STIRRER CONFIGURATION	
8.2.5. STIRRING SPEED	
8.3. RELATIVE mV CALIBRATION	
8.4. LOGGING	
8.4.1. INTERVAL LOGGING	
8.4.2. MANUAL LOGGING	

mv mode

By pressing Mode from the main screen, the Titrator can be switched to **Titrator**, **pH**, **mV** or **ISE** modes. **One Analog Board**

Working Mode				
Select the work	ing mode.			
<u>Titrator</u>	ρH	mŲ	ISE	



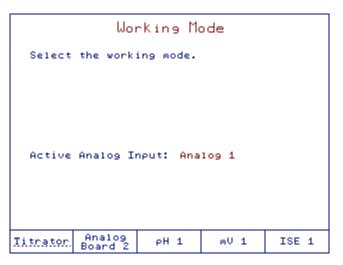
Switches to Titrator mode.

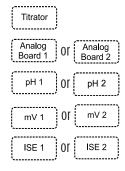
Switches to **pH** mode.

Switches to **mV** mode.

Switches to **ISE** mode.

Two Analog Boards





Switches to **Titrator** mode.

Switches the Analog Input for pH, mV and ISE mode.

Switches to **pH** mode.

Switches to \mathbf{mV} mode.

Switches to $\ensuremath{\mathsf{ISE}}$ mode.

8.1. DISPLAY

The **mV** screen is shown below.

10:21:21 Analog 1	May	24,	2018			
2.8 MU 200/ 200 RPM						
атс 26.3 °(_			
General Options		art 09		Rel ibr.	mŲ Setup	Mode

mV Mode Option Keys:



The General Options screen gives access to options that are not directly related to the measurement process (see **General Options** section).

Save Reading

٥r

	Start Log mV Rel Calibr.)))
i Ci C	mV Setup Mode	

Starts the mV interval log (see Automatic Logging section).

Stores the current mV reading (see Manual Logging section).

Enter the mV calibration screen (see **pH Calibration** section).

Enter the mV setup screen, parameters are associated with pH measurements and calibration (see pH Setup section).

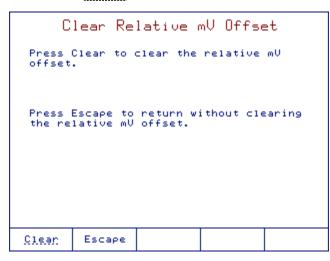
Allow the user to switch between the available measurement modes: Titrator, pH, mV or ISE mode.

8.2. mV SETUP

mV Setup				
Select	a menu optior			
Clear Relative mV Offset Logging Interval: 0h:00m:02s Stability Criteria: Fast Stirrer Configuration: Stirrer 1 Stirring Speed: 1200 RPM				
Select	Escape			

8.2.1. CLEAR RELATIVE mV OFFSET

Press Clear to clear the relative mV offset or Escape to return to the previous screen.



8.2.2. LOGGING INTERVAL

Option: 2 seconds to 8h 59min 59sec

Logging Interval				
Enter	the data	logging ir	nterval.	
	0	0	2	
	hours	minutes	seconds	
Press Next to move to the next entry.				
Accept	Escape	Delete Digit	Next	Off

8.2.3. STABILITY CRITERIA

Option: Fast, Medium or Accurate:

	I	mV Setu	P	
Select	a menu op	tion.		
Logging Stabili Stirrer	elative m Interval ty Criter Configur g Speed:	: ia:	l eas Med	00m:02s Fast 1 t ium urate
Select	Escape			

Fast: Quicker results with less accuracy

Medium: Medium speed results with medium accuracy

Accurate: Slower results with high accuracy

8.2.4. STIRRER CONFIGURATION

Option: Stirrer 1, Stirrer 2 (if available) or Disabled

	(mV Setu	P	
Select	a menu op	tion.		
Logging Stabili	elative m Interval ty Criter Configur	: ia:		:00m:02s Fast tirrer 1
	g Speed:	ation.	Dis	abled rrer 1 rrer 2
Select	Escape			

8.2.5. STIRRING SPEED

Option: 200 to 2500 RPM

	Stirring Speed				
Enter below (of the st	tirrer wit	thin:	
1100 RPM					
The range is from 200 to 2500 RPM.					
Accept	Escape	Delete Digit			

8.3. RELATIVE mV CALIBRATION

Relative mV				
Analog 1				
Set the value for the relative mV offset.				
Absolute mV: 2.7 mV				
<mark>Stirrer 1</mark> 1100/1100 RPM				
Relative mV: 2.7 mV				
Low limit: -1997.3 mV				
High limit: 2002.7 mV				
Accept Escape Delete Digit				

- Press Accept to accept the value.
- Press Escape to cancel this operation and return to the previous screen.
- Press Delete Digit to delete the last digit.

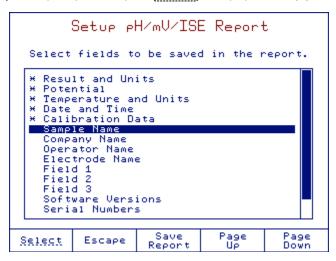
mV MODE

8.4. LOGGING

Data logging is available in mV mode. It can be logging on demand (Manual Logging) or automatically (Interval Logging) at predefined time intervals.

To customize the logging report:

- Press (results) to display the Data Parameters screen.
- Highlight the Setup pH/mV/ISE Report option and press select to display the Setup pH/mV/ISE Report screen.



- Use the A and keys to highlight the data field that you want to show/hide in the pH/mV/ISE report and then press
 Select to activate/deactivate it.
- Each field marked by "*" is an active field selected for the report.
- Press save Report to save the customized report.

8.4.1. INTERVAL LOGGING

The logging interval is set in the mV Setup screen.

 $\Press \xrightarrow[Log]{Start} to start the log.$

The logging interval and name of logging file will be displayed on the measure screen. To stop the automatic logging, press stop again.

8.4.2. MANUAL LOGGING

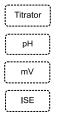
To manually log mV readings, press $\begin{bmatrix} Save \\ Reading \end{bmatrix}$ from the **mV** measurement screen. A new record will be added to the report every time $\begin{bmatrix} Save \\ Reading \end{bmatrix}$ is pressed.

CHAPTER 9. ISE MODE

9.1. DISPLAY	4
9.2. ISE SETUP	4
9.2.1. CALIBRATION GROUP	5
9.2.2. TEMPERATURE COMPENSATION	5
9.2.3. ISOPOTENTIAL POINT	6
9.2.4. EDIT CUSTOM STANDARDS	6
9.2.5. EDIT STANDARD GROUP	7
9.2.6. CALIBRATION REMINDER	7
9.2.7. SET REMINDER PERIOD	7
9.2.8. CLEAR CALIBRATION	8
9.2.9. ISE GLP DATA	.9
9.2.10. ELECTRODE TYPE	9
9.2.11. CONCENTRATION UNIT	0
9.2.12. LOGGING INTERVAL	10
9.2.13. STABILITY CRITERIA	0
9.2.14. ISE SIGNIFICANT DIGITS	11
9.2.15. STIRRER CONFIGURATION	1
9.2.16. STIRRING SPEED	11
9.3. ISE CALIBRATION	12
9.4. LOGGING	13
9.4.1. INTERVAL LOGGING	4
9.4.2. MANUAL LOGGING	4

By pressing Mode from the main screen, the Titrator can be switched to **Titrator**, **pH**, **mV** or **ISE** modes. **One Analog Board**

Working Mode					
Select the working mode.					
Titrator.		ρH	mŲ	ISE	



Switches to **Titrator** mode.

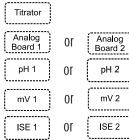
Switches to $\mathbf{p}\mathbf{H}$ mode.

Switches to ${\bf mV}$ mode.

Switches to $\ensuremath{\mathsf{ISE}}$ mode.

Two Analog Boards

Working Mode					
Select	the work	ing mode.			
Active Analog Input: Analog 1					
<u>Titrator</u>	Analog Board 2	ρH 1	mV 1	ISE 1	



Switches to Titrator mode.

Switches the Analog Input for pH, mV and ISE mode.

Switches to **pH** mode.

Switches to **mV** mode.

² Switches to **ISE** mode.

9.1. DISPLAY

The **ISE** screen is shown below.

10:56:51	May 24,	2018		
Analog 1				
			_	
		64.6	D PPM	
				tirrer 1 1100 RPM
		ISE: Silve		1100 1.11
ATC				mŲ
26.1 °C 197.7				
General Options	Start Log	ISE Calibr.	ISE Setup	Mode

ISE Mode option keys:



The General Options screen gives access to options that are not directly related to the measurement process (see **General Options** section).

Save Reading

0ľ

Start Log St Calibr. Er ISE Setup Er

Mode

Starts the ISE interval log (see Interval Logging section).

Enter the ISE calibration screen (see ISE Calibration section).

Stores the current concentration reading (see Manual Logging section).

Enter the ISE setup screen. Parameters are associated with ISE measurements and calibration.

Allows the user to switch between the available measurement modes: **Titrator**, **pH**, **mV** and **ISE** mode.

9.2. ISE SETUP

To access the ISE Setup, press setup option key in ISE mode.

	ISE Setu	Р	
Select	a menu option.		_
Calibr	ation Group:	All Standards	
	ature Compensation:	Disabled	
	ential Point:	None	
Edit C	ustom Standards:		
Edit S	tandards Group:		
Calibr	ation Reminder:	Disabled	
Set Re	minder Period:	Disabled	
	Calibration		
ISE GL			
	ode Type:	Silver	
	tration Unit:	PPM P	
	a Interval:	0h:00m:02s	
Stabil	ity Criteria:	Medium	
L			
Select	Escape		
Serect	COCOPE		

ISE MODE

9.2.1. CALIBRATION GROUP

Option: All Standards or Standards Group

ISE S	etup
Select a menu option.	
Calibration Group:	All Standards
Temperature Compensat Isopotential Point: Edit Custom Standards Edit Standards Group:	All Standards Standards Group
Calibration Reminder: Set Reminder Period: Clear Calibration ISE GLP Data	Disabled Disabled
Electrode Type: Concentration Unit: Logging Interval: Stability Criteria:	Silver PPM Oh:OOm:O2s Medium
Select Escape	

All Standards:The set of available standards includes the Standard solutions and Custom solutions.Standards Group:The set of available standards includes only the standards selected by the user.

9.2.2. TEMPERATURE COMPENSATION

Option: Enabled or Disabled

ISE Setup				
Select a menu option.				
Calibration Group: Temperature Compensation:	All Standards			
Isopotential Point: Edit Custom Standards: Edit Standards Group: Calibration Reminder:				
Set Reminder Period: Clear Calibration ISE GLP Data	Disabled			
Electrode Type: Concentration Unit: Logging Interval: Stability Criteria:	Silver PPM Oh:OOm:O2s Medium			
<u>Select</u> Escape				

Note: If you enabled Temperature Compensation, then the isopotential point must be set.

9.2.3. ISOPOTENTIAL POINT (TEMPERATURE COMPENSATION)

Option: 1.00 E⁻² to 1.00 E⁺⁵ ppm

This option allows the user to set an isopotential point for the selected electrode when temperature compensation is enabled. The isopotential point is edited in ppm units only. The isopotential point will vary for different electrodes, if measurements are going to be made at several temperatures, the value should be entered if it is known.

Isopotential Point				
Enter	the value	for isop	otential	point.
		20.0	0 PPM	
Low li	Low limit: 1.00E-2 ppm			
High limit: 1.00E+5 ppm				
Accept	Escape	Delete Digit		Exponent

9.2.4. EDIT CUSTOM STANDARDS

Option: Up to five

	Edit Cu	stom St	andards	
Press	<edit> to</edit>	edit sele	ected star	ndard.
	(Remove S [.] stom stand	tandard> ∶ dard.	to delete	
4.00	40.0	400		
PPM	PPM	PPM		
Use arrows keys to select the standard.				
Remove Standard	Escare	Edit	\triangleleft	\triangleright

- Use the \lt and \triangleright keys to select the standard.
- Press Remove to delete the standard.
- Press Edit to edit the selected custom standard; use the numeric keys to edit the standard.

9.2.5. EDIT STANDARD GROUP

Option: Up to 5 standards

- Use the arrow keys to select the standard to be included/removed in/from the standard group.
- Press Add or Remove to add/remove the selected standard to/from standard group.
- Press Escape to return to ISE Setup menu.

Edit Standards Group				
Available Standards ppm				
E-1 1.00 1.00 2.00 10.0	100			
Standards Group ppm				
E-1 1.00 2.00 100 1000 10000				
Remove Escape D				

9.2.6. CALIBRATION REMINDER

Option: Daily, Periodic or Disabled

	Calibration Reminder	
Select	a menu option.	
Daily Period Disabl		
0		
Select	Escape	

Daily: The calibration reminder will appear daily at specified time.

Periodic: The calibration reminder will appear after the set time has elapsed since the last calibration.

Disable: The calibration reminder will not appear.

9.2.7. SET REMINDER PERIOD

If Daily or Periodic option was selected for the calibration reminder, the reminder period must also be set. For a daily reminder period the time of day can be set.

For a periodic reminder period the number of days, hours and minutes can be set.

Peri	odic Ca	alibrati	on Remi	nder
Enter the time period that must be passed since the last calibration, whereafter the calibration reminder appears.				
	10 days	2 hours	30 minutes	ŝ
Press	Next to m	ove to the	2 next ent	ny.
Accept	Escape	Delete Digit	Next	Off

- Press Next to move the cursor to the next field.
- Press Accept to save the changes or Escape to return to the previous screen.
- Press of to disable the calibration reminder and return to ISE setup menu.

9.2.8. CLEAR CALIBRATION

This option clears the existing ISE calibration. If the calibration is cleared, a new calibration must be done in order to take measurements.

• Press clear to clear the previous calibration or Escape to return to the previous screen.

```
Clear Calibration

Press (Clear) to clear all calibration

points.

Press (Escape) to return without clearing

the calibration points.

<u>Clear</u> Escape
```

9.2.9. ISE GLP DATA

Displays the ISE calibration data

```
ISE GLP Data
Analog 1
  Last Calibration: 13:42 |
Slope: 100.8%
Isopotential Point: 20.0 ppm
                                    13:42 May 24, 2018
ISE: Silver
                      0.1mU 28.1°C A
13:39:43 May 24, 2018
   1.00E-1 ppm,
   1.00 ppm,
                   59.5mV
                      ).5mV 28.1°C A
13:40:39 May 24, 2018
                   77.6mU 28.1°C A
13:41:25 May 24, 2018
   2.00 ppm,
                   120.0mV
                      0.0mV 28.1°C A
13:41:45 May 24, 2018
   10.0 ppm,
                      .0mV 28.2°C A
13:42:17 May 24, 2018
   100 ppm,
                  181.0mV
              Escape
```

9.2.10. ELECTRODE TYPE

Option: Ammonia, Bromide, Cadmium, Calcium, Carbon Dioxide, Chloride, Cupric, Cyanide, Fluoride, Iodide, Lead, Nitrate, Potassium, Silver, Sodium, Sulfate, Sulfide or five custom electrodes

	Elec	trode (Гуре	
Select	a menu og	stion.		
Ammoni Bromid Cadmiu Carbon Chlori Cupric Cyanid Fluori Iodide Lead Nitrat Potass Silver	e M Dioxide de e de			
Select	Escape	View	Page Up	Page Down

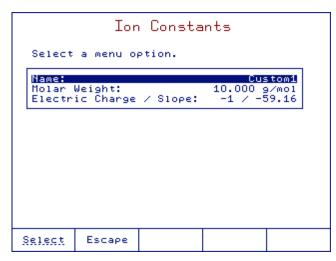
For Standard ISE:

Press Select to see the ion constants (name, molar weight, electric charge/slope), press Escape to return to the setup screen.

Ion Constants
View Ion constants.
Name: Silver Molar Weight: 107.868 g/mol Electric Charge / Slope: 1 / 59.16
Escape

For Custom ISE:

• Press view to edit the Ion constants for the selected custom ISE.



- Use the \triangle and \bigtriangledown keys to highlight the desired ion constant and press select to edit the value.
- Set the Ion Name (up to 10 characters can be entered).
- Set the appropriate Molecular Weight (in g/mol) using the numeric keys. Press Accept to save the value or press Escape to return to the previous screen.
- Select the appropriate Electric Charge/Slope. Use the A and keys to select the value and then press select. If the electric charge is none, manually set the slope by pressing select. Press Accept to save the value or press screen.

9.2.11. CONCENTRATION UNIT

Options: ppt (g/L), ppm (mg/L), ppb (µg/L), mg/mL, M (mol/L), mmol/L, %w/v or user defined

ISE Setur				
Select a menu option.				
Calibration Group: Temperature Compensation: Isopotential Point: Edit Custom Standards:	All Standards Disabled None			
Edit Standards Group: Calibration Reminder: Set Reminder Period: Clear Calibration ISE GLP Data Electrode Type:	PPt g∕L ppm Mg/L			
Concentration Unit: Logging Interval:	ррм Oh:00m:02s			
Stability Criteria:	Medium			
<u>Select</u> Escape				

9.2.12. LOGGING INTERVAL

Option: 2 seconds to 8h 59 min 59 sec

Logging Interval					
Enter	the data :	logging in	nterval.		
	0	o	2		
	hours	minutes	seconds		
Press	Next to m	ove to the	e next ent	my.	
Accept	Escape	Delete Digit	Next	Off	

9.2.13. STABILITY CRITERIA

Option: Fast, Medium, Accurate

ISE Setu	Ρ
Select a menu option.	
Calibration Group: Temperature Compensation: Isopotential Point: Edit Custom Standards:	All Standards Enabled 20.0 ppm
Edit Standards Group: Calibration Reminder: Set Reminder Period: Clear Calibration	Disabled Disabled
ISE GLP Data Electrode Type: Concentration Unit: Logging Interval:	Fast Medium Accurate
Stability Criteria:	Medium
<u>Select</u> Escape	

Fast: Quicker results with less accuracy

Medium: Medium speed results with medium accuracy

Accurate: Slower results with high accuracy

9.2.14. ISE SIGNIFICANT DIGITS

Option: One (X), Two (XX) or Three (XXX).

	ISE	E Setur		
Select	a menu optic	on.		
Edit Cu	ature Compen Antial Point Astom Standa	: rds:	Enabled 20.0 ppm	
Calibra Set Ren Clear (ation Remind Minder Perio Calibration	len:	Disabled Disabled	
Concent Logging	' Data de Type: ration Unit Interval: ty Criteria	-	0h:0	
ISE Significant Digits: XXX				
Select	Escape			

9.2.15. STIRRER CONFIGURATION

Option: Stirrer 1, Stirrer 2 (if available) or Disabled

Select a menu option.	
Isopotential Point: Edit Custom Standards: Edit Standards Group:	20.0 ppm
Calibration Reminder: Set Reminder Period: Clear Calibration ISE GLP Data	Disabled Disabled
Electrode Type: Concentration Unit: Logging Interval: Stability Criteria:	Disabled Stirrer 1 Stirrer 2
ISE Significant Digits: Stirrer Configuration:	Stirrer 1

9.2.16. STIRRING SPEED

Option: 200 to 2500 RPM

	Stirring Speed					
Enter below (of the s	tirrer wit	hin:		
		250	D RPM			
The ra	nge is fro	om 200 to	2500 RPM.			
Accept	Escape	Delete Digit				

9.3. ISE CALIBRATION

It is recommended to calibrate the instruments frequently if high accuracy is required. The instrument should also be recalibrated whenever the "Calibrate Electrode" message appears on the LCD.

Due to electrode conditioning time, the electrode must be immersed for several seconds to stabilize. The user will be guided step by step during calibration with easy-to-follow messages on the display. This will make the calibration a simple and error-free procedure. **PREPARATION**:

Pour small quantities of the standard solution into clean beakers. If possible, use plastic beakers to minimize any EMC interferences. For accurate calibration and to minimize cross-contamination, use two beakers for each standard solution: one for rinsing the electrode and one for calibration.

Note: For accurate measurements, add the appropriate ISA (Ionic Strength Adjustment) to the calibration standards. CALIBRATION PROCEDURE:

Before calibrating, make sure that the electrode type and concentration unit has been selected in ISE Setup.

Up to a five points calibration is possible using any combination of five standard solutions and five custom solutions.

The ISE calibration and measurement can be performed with or without temperature compensation. If the temperature compensation option is enabled, the isopotential point of the electrode must be set in ISE Setup.

- Press IsE Calibr. from the main screen. If the instrument was calibrated before and the calibration was not cleared, the old calibration can be cleared by pressing Clear Cal.
- Immerse the ISE and the temperature probe approximately 2 cm into the standard with the lowest concentration.

	ISE	Calibra	ntion	
Analog 1		9.99	9 ^{E-2}	
	I	SE: Silve	in in	
атс 24.8 °	°C	E-1 1.00		∾V 0.0
Calibrat	ted Standa	ands		
E-1 1.00 Last C		10.0 n: 10:40	100 Ju1 03, 20	1000
Press	(Accept)	to update	calibrati	ion.
Accept	Escape	Edit		Previous Standard

- Select the standard concentration with <u>Standard</u> or <u>Previous</u> standard.
- When the reading has stabilized, press Accept to update the calibration. The calibration point value will be added to the Calibrated Standard list.
- Select Next Standard and repeat the procedure with all of the available standards.
- Press Escape to exit the calibration.

9.4. LOGGING

Data logging is available in ISE mode. It can be logging on demand (Manual Logging) or automatically (Interval Logging) at predefined time intervals.

To customize the logging report:

- Press results to display the Data Parameters screen.
- Highlight the *Setup pH/mV/ISE Report* option and press [Select] to display the **Setup pH/mV/ISE Report** screen.

Setup pH/mV/ISE Report	
Select fields to be saved in the report.	
<pre>* Result and Units * Potential * Temperature and Units * Date and Time * Calibration Data Sample Name Company Name Operator Name Electrode Name Field 1 Field 2 Field 3 Software Versions Serial Numbers</pre>	
Select Escape Save Page Page Report Up Down	

- Use the 🛆 and 🤍 keys to highlight the data field that you want to show/hide in the pH/mV/ISE report and then press
- Each field marked by "*" is an active field selected for the report.
- Press Save Report to save the customized report.

9.4.1. INTERVAL LOGGING

The logging interval is set in the ISE Setup screen.

 $\operatorname{Press} \underbrace{[]_{\text{Log}}^{\text{Start}}}_{\text{Log}} \text{ to start the log.}$

The logging interval and name of logging file will be displayed on the measure screen.

To stop the automatic logging, press stop again.

9.4.2. MANUAL LOGGING

To manually log ISE readings, press $\begin{bmatrix} Save \\ Reading \end{bmatrix}$ from the **ISE** screen. A new record will be added to the report every time $\begin{bmatrix} Save \\ Reading \end{bmatrix}$ is pressed.

CHAPTER 10. AUXILIARY FUNCTIONS

10.1. BURETTE	. 10-3
10.1.1. PRIME BURETTE	. 10-3
10.1.3. MANUAL DISPENSE	10-4
10.1.4. PURGE BURETTE	. 10-5
10.1.5. PERISTALTIC PUMP	. 10-6
10.2. STIRRER	. 10-6
10.3. RESULTS	. 10-6
10.3.1. REVIEW LAST ANALYSIS REPORT	. 10-7
10.3.2. REVIEW AVAILABLE REPORTS	
10.3.3. GLP DATA	. 10-8
10.3.4. METER INFORMATION	. 10-9
10.3.5. SETUP pH/mV/ISE REPORT	.10-10
10.3.6. SETUP TITRATION REPORT	10-10

AUXILIARY FUNCTIONS

10.1. BURETTE

To access the **Burette** screen, press Burette from the main titration screen. Highlight the desired option and then press Select.

		Burette	:	
Select	a menu op	tion.		
Prime B Rinse T Manual Purge B	ip Dispense			
	rrent pum; ≥ is not ;	⊃ is: Pump ⊃resent.	> 1	
Select	Escape	Choose Pump	Perist.1 On	Perist.2 On

^{Choose} Pump allows you to select the desired pump for burette operations (it is only active if two pumps are connected).

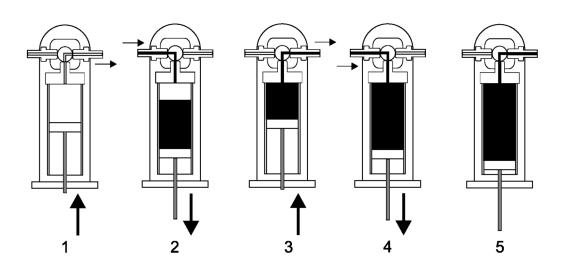
		Pur	mp Sett	ing	
	Select	the curre	ent pump.		
	Pump 1 Pump 2				
9	elect	Escape			

10.1.1. PRIME BURETTE

Option: Up to 5

The *Prime Burette* option is used to fill the burette with titrant or reagent before starting a titration. The priming process consists of several cycles of filling and emptying the burette with titrant.

Two rinse cycles of burette are shown in the figure below. The dispensing tube is connected on the right side and the aspiration tube on the left side.



Note: Before starting this operation, the aspiration tube must be inserted in the titrant bottle. A waste container should be placed under the dispensing tip to collect the waste solution.

To prime the burette, select *Prime Burette*, enter the number of rinses and press Accept. We recommend at least three rinses to assure that the air bubbles are completely removed.

Total Burette Rinses						
Enter	the total	number of	f burette	rinses.		
			3			
A mini	A minimum of three rinses is recommended.					
Accept	Escape	Delete Digit				

10.1.2. **RINSE TIP**

A 2 mL dose of titrant will be dispensed from the burette when this operation is selected, this will eliminate any air in the dispensing tip.

10.1.3. MANUAL DISPENSE

Manual Dispense option allows a defined titrant volume to be dosed. Select the Manual Dispense option and press

	Manual (Volume [Dispense	2		
Enter dispen	the amoun sed.	t of volu	ne to be			
		1.00	u mL			
Curren	Current burette volume is 25 mL.					
Accent	Escape	Delete Digit				

Use the numeric keypad to enter the volume to be dispensed.

The manual dispense volume must be between the limits shown below:

 0.001 to 4.750 mL
 for a
 5 mL
 burette

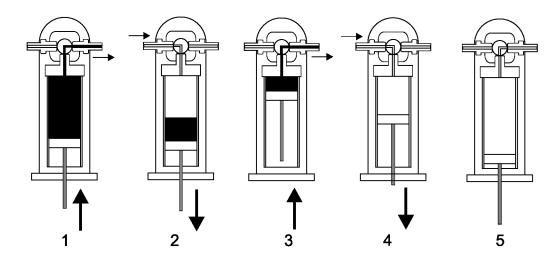
 0.001 to 9.500 mL
 for a
 10 mL
 burette

 0.005 to 23.750 mL
 for a
 25 mL
 burette

 0.005 to 47.500 mL
 for a
 50 mL
 burette

10.1.4. PURGE BURETTE

This option allows the burette to be emptied before cleaning and/or storing the burette. The burette is flushed twice. *Note: Before starting this operation, remove the aspiration tube from the titrant bottle.* The figures below show the steps in a purge burette operation.



10.1.5. PERISTALTIC PUMP

To manually control the peristaltic pump, press Burette from the main titration screen.

, ,		·				
Press Perist.1 or Perist.2 on the selected peristaltic pump.						
Press Off Of Off To tu	Press Perist 1 Off Off Off Off To turn off the selected peristaltic pump.					
			Burette	2		
	Select	a menu op	tion.			
	<mark>Prime Burette</mark> Rinse Tip Manual Dispense Purge Burette					
		rrent pum; t burette				
	Select	Escape	Choose Pump	Perist.1 Off	Perist.2 On	

Note: If the peristaltic pump is not turned off, it will turn off automatically after 10 minutes.

10.2. STIRRER

The stirrer can be turned on and off by pressing stir.

During the titration process, the stirring speed can be manually adjusted using the \triangle and $\overline{\bigtriangledown}$ keys.

10.3. RESULTS

From the Data Parameters screen, you can access the following options:

	Data	a Parame	ters:	
Select	a menu op	tion.		
Review GLP Dat Meter I Setup p	Last Anal Available a nformatio H/mV/ISE itration	Reports n Report	rt	
Select	Escape			

AUXILIARY FUNCTIONS

10.3.1. REVIEW LAST ANALYSIS REPORT

Review Result					
ISEOC	0020.RPT =				
	HI93:	2 - ISE Re	eport		
Method Name: pH/mV/ISE logging Time & Date: 14:11 May 24, 2018 Logging ID ISE00020					
	Cal	ibration [Data		
Standard Potential Efficiency Temp. Time and Date 1.00E-1ppm 0.1mV 99.4% 28.1°C A 13:39 May 24, 2018 1.00ppm 59.5mV 100.5% 28.1°C A 13:40 May 24, 2018					
View Graph	Escars	Print Report	Page Up	Page Down	

The information seen in the report is based on the selections made in the **Setup Titration Report** and **Setup ISE/pH/mV Report** screen.

The following option keys are available:

Review the graph.

View Graph

Print Report Print the titration report.

Escape Return to the previous screen.

Page Down Keys can be used to scroll through the pages.

10.3.2. REVIEW AVAILABLE REPORTS

Up to 100 reports can be saved on the titrator. To view one of the saved reports, highlight a report and then press view Report.

to see	ght a rep the deta	iled data.	ress View	
PH/mV/ ISE Re	ISE loggin cont		ID:ISE(1 May 24,	
PH/mV/ PH Rep 1.0N N Titrat PH/mV/ MV Rep PH/mV/ mV Rep	ISE loggi ort aOH Titr. ion Repor ISE loggi ort ISE loggi ISE loggi ISE loggi	ng 10:00 Conc. t 09:00 ng 09:02 ng 09:02	ID:pH_0 3 May 24,	00019 2018 00018 2018 00017 2018 00016 2018 00016 2018
	ISE loggi	ng	ID:рН_(L May 23,	00014
View Graph	Escape	View Report	Print Report	Delete Report

The report contains only the information selected in the **Setup Titration Report** and **Setup pH/mV/ISE Report** screens during report configuration.

The following option keys are available:

View Graph	Review the selected graph.
View Report	Review the selected report.
Print Report	Print the selected report.
Delete Report	Delete the selected report.
Escape	Return to the previous screen.

10.3.3. GLP DATA

Option: Up to 20 characters

	y Name:			
Operator Name: Electrode Name: Field 1:				
Field 2: Field 3:				

Sample Name: Allows the sample name to be recorded in each report. The sample name will increase by one, with each new titration or logging report, if the last character is a number.

Company Name: Allows the company name to be recorded in each report.

Operator Name: Allows the operator name to be recorded in each report.

Electrode Name: Allows the electrode name to be recorded in each report.

Fields 1, 2, 3: Allows any additional information to be recorded in each report.

The fields must be selected from **Setup Titration Report** screen (see **Setup Titration Report** section) in order to be displayed in the titration report.

10.3.4. METER INFORMATION

Displays titrator configuration data.

Titrato Analog Analog Pump 1	. NUMBER r Serial Board1 Se Board2 Se Serial No	Inform 932 Titra Number: erial Numb erial Numb umber: 1 Number:	tor 121 per: 301 per: 300 700	33404404 34202202 0000000 94513513 91703703	
SOFTWARE VERSION Titrator Software Version: v1.00 Base Board Software Version: v1.00 Pump 1 Software Version: v1.00 Stirrer 1 Software Version: v1.00					
Analog 1 Calibration Date: May 22, 2018 Analog 2 Calibration Date: May 10, 2018					
	Escare	Print			

Titrator Serial Number: The serial number of the titrator base board.

Analog Board 1 (and/or 2) Serial Number: The serial number of the analog board.

Pump 1 (and/or 2) Serial Number: The serial number of the connected pump.

Titrator Software Version: The current software version installed on the titrator.

Base Board Software Version: The current software version present on the base board of the titrator.

Pump 1 (and/or 2) Software Version: The current software version for the pump.

Analog 1 (and/or 2) Calibration Date: Manufacturer calibration date of the analog board.

Note: If more than 1 year elapsed from the calibration date of the analog board 1 and/or 2, the message *Analog 1 Calibration Due* and/or *Analog 2 Calibration Due* will appear on the main screen. The analog board(s) need to be recalibrated.

10.3.5. SETUP pH/mV/ISE REPORT

Customize a unique report to record the pH, mV, and ISE measurements. An asterisk means that it will be included in the report.

	Setup pl	HZmUZTSF	- Report	F.
Select fields to be saved in the report.				
* Pote * Temp * Date * Cali Samp Oper Elec Fiel Fiel Fiel Soft	erature and Time bration D: le Name any Name ator Name trode Name d 1	nd Units ata 2		
Select	Escape	Save Report	Page Up	Page Down

C	Select)	
ĺ	Unselec)	
C	Escape)	
C	Save Report)	
ĺ	Page Up)(Page Down

Adds the highlighted information to the report.

Removes the highlighted information from the report.

Returns to the Data Parameter Screen. Report is not updated.

Update the report with the select items. Report previously saved will not be updated.

Scroll through the options.

10.3.6. SETUP TITRATION REPORT

Customize a unique report to record the titration results. An asterisk means that it will be included in the titration report.

Setup Titration Report Select fields to be saved in the report.					
Result and Units Titration Method Initial and Final Readings Analyte Size End Point Volume Titration Duration Date and Time Titration Ended By All Data Points Method Parameters Calibration Data Sample Name Operator Name					
Unselect	Escape	Save Report	Page Up	Page Down	

Select)
Unselect]
Escape)
Save Report]
Page Up	Page Dowi

Adds the highlighted information to the report.

Scroll through the options.

Removes the highlighted information from the report.

Returns to the Data Parameter Screen. Report is not updated.

Update the report with the select items. Report previously saved will not be updated.

e n

CHAPTER 11. MAINTENANCE, PERIPHERALS

11.1. BURETTE MAINTENANCE	11-3
11.1.1. BURETTE ASSEMBLY	11-3
11.1.2. CHANGING THE BURETTE	11-3
11.1.3. DISASSEMBLING THE BURETTE	11-3
11.1.4. ASSEMBLING THE BURETTE	11-4
11.1.5. CLEANING THE BURETTE	11-4
11.1.6. BURETTE PREPARATION (FILLING WITH TITRANT)	11-5
11.2. PERIPHERALS	11-6
11.2.1. CONNECTING TO A PRINTER	11-6
11.2.2. CONNECTING AN EXTERNAL PC KEYBOARD	11-6
11.2.3. CONNECTING TO A COMPUTER	

MAINTENANCE PERIPHERALS

The 25-mL burette included with the titrator exceeds the ISO 8655 standard for accurate delivery of liquids by a motor-driven piston burette.

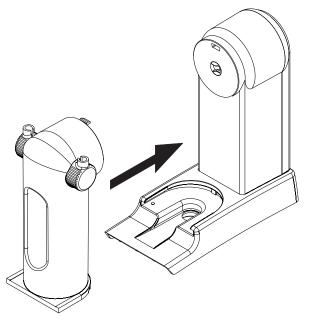
11.1. BURETTE MAINTENANCE

11.1.1. BURETTE ASSEMBLY

The burette is delivered with a 25-mL syringe inside and with all of the accessories mounted (see **Setup** chapter). The burette assembly consists of a rigid housing which holds the glass syringe, a 3-way valve and titrant tubing.

11.1.2. CHANGING THE BURETTE

Remove the burette from the pump assembly by sliding it forward and then slide the new burette into place.

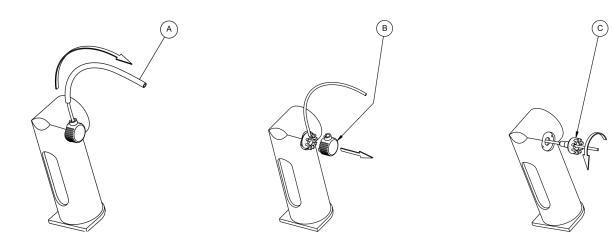


11.1.3. DISASSEMBLING THE BURETTE

The aspiration and the dispensing tubes have fittings and tube protectors. The aspiration tube is mounted in the left side and the dispensing tube is mounted in the right side of the burette.

To remove the dispensing tube and the aspiration tube follow these steps:

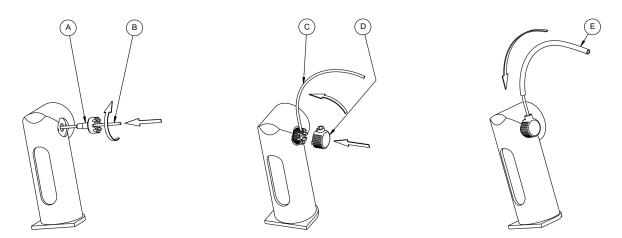
- Remove the blue tube protector (A) by sliding it off the clear titrant tubing.
- Remove the tube lock (B) from the burette holder.
- Turn the fitting (C) counter-clock wise to remove it from the burette holder.
- Slide the clear titrant tubing through the fitting.



11.1.4. ASSEMBLING THE BURETTE

To attach the dispensing tube and the aspiration tube, follow these steps:

- Insert the flat-shaped end of the titrant tubing into the valve outlet (A) and screw the fitting clock-wise to tighten. The highest of the 9 cuts should be vertical in the final position.
- Bend the tube up into the vertical position to enter the highest cut of the fitting (C).
- Replace the tube lock fitting (D).
- Replace the blue tube protector (E) by sliding it over the clear titrant tubing, the protector will sit in the tube lock fitting.



11.1.5. CLEANING THE BURETTE

To clean the burette, follow these steps:

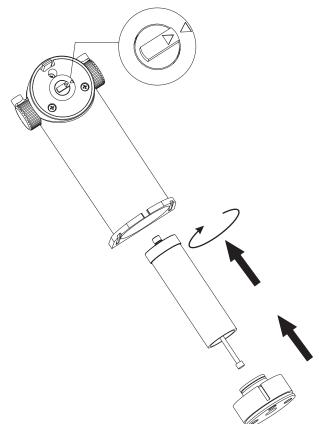
- If the burette is filled with titrant, remove the aspiration tube from the titrant bottle and purge burette (see Auxiliary Functions chapter).
- Insert the aspiration tube into cleaning solution, deionized water or titrant solvent.
- Prime burette to fill the burette (use 2 rinses) (see Auxiliary Functions chapter).
- During second cycle remove the aspiration tube from the cleaning solution, deionized water or solvent and allow the air to replace the liquid in the burette. This will clean the aspiration tube.

If this simple cleaning procedure is not adequate, continue with these steps:

- Remove the burette assembly from the pump.
- Remove the dispensing and aspiration tubes. Clean them separately or insert new ones.
- Remove the protective cap from the bottom of the burette assembly by using the burette removal tool.
- Remove the syringe from the burette assembly by unscrewing it with your fingers.
- Extract the piston from the syringe.
- Clean both the piston and the syringe with appropriate cleaning solution. Rinse with deionized water.
- Remove the excess liquid.

Warning: Avoid contacting the titrant with bare hands.
 Avoid spilling titrant.
 Clean the external side of the syringe and piston to remove aggressive chemicals.
 Do not touch the white PTFE part of the piston or internal walls of the burette with bare hands or greasy materials.

- Reinsert the piston into the syringe.
- Reinsert the syringe by screwing it in the valve with your fingers.
- Reinsert the protective cap to the bottom of the burette assembly. Carefully position the cap into the burette.
- Slide the burette into the burette stand. Notice the position of the piston shaft to the pump couple.
- Priming the burette three times with new titrant is recommended.



11.1.6. BURETTE PREPARATION (TITRANT FILLING)

Before starting a titration, the burette must be properly filled with titrant in order to obtain an accurate and repeatable result. To fill the burette, follow the next steps and recommendations:

- If necessary, clean the burette and make sure it is empty.
- From the main screen press
- Highlight *Prime Burette* option and press select
- Enter the number of times the burette needs to be rinsed (minimum three rinses are recommend allowing air bubbles to be evacuated).
- Press Accept .

To avoid the presence of the air bubbles inside the burette, make sure to have a continuous liquid flow inside the burette. A little air just above the liquid level at the first filling is normal. The next filling will evacuate all of the air; no air will be left in the valve. Sometimes during this process, slight finger tapping on the tubes is helpful to remove any residual air bubbles from the tubes. If air bubbles are still present:

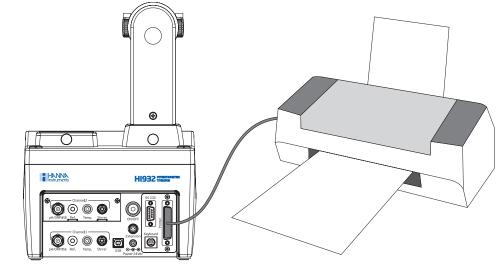
- Remove the aspiration tube from the titrant bottle.
- Repeat burette preparation procedure.
- If this is not successful, clean the burette again.

11.2. PERIPHERALS

Warning! Connection/disconnection of POWER, PUMP ASSEMBLY, PRINTER, RS232 INTERFACE or AUTOSAMPLER must only be done when Titrator and external devices are turned off.

11.2.1. CONNECTING TO A PRINTER

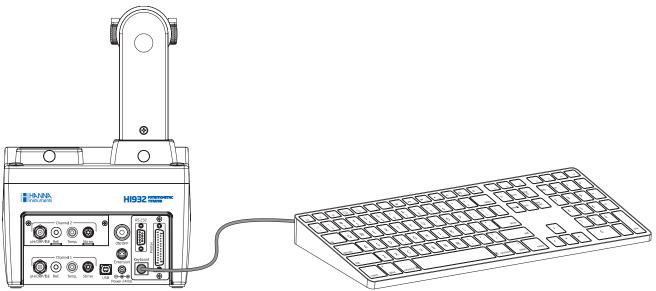
A variety of parallel printers can be connected to the parallel port of the titrator using a DB25 cable.



Warning: The titrator and the external printer must be both turned OFF before they are connected.

11.2.2. CONNECTING AN EXTERNAL PC KEYBOARD

This connection allows you to use an external PS/2 PC Keyboard in addition to the titrator's keypad.

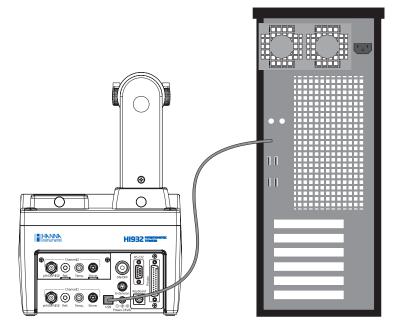


The correspondence between the titrator's keypad and the United States 101-type external keyboard are:

External PC Keyboard (United States 101)	Titrator Keypad
Function Key F-1	?
Function Key F-2	stir
Function Key F-3	results
Function Key F-4	device
Function Key F-5	Option Key 1 (from left to right)
Function Key F-6	Option Key 2 (from left to right)
Function Key F-7	Option Key 3 (from left to right)
Function Key F-8	Option Key 4 (from left to right)
Function Key F-9	Option Key 5 (from left to right)
Function Key F-10	start stop
Arrow Key: Up	
Arrow Key: Down	
Arrow Key: Left	
Arrow Key: Right	
Page Up	Page Up
Page Down	Page Down
Numeric Keys: 0 to 9	(e) ₀₁ (0)
Enter	enter
Alphanumeric Keys	Allow alphanumeric entries.

11.2.3. CONNECTING TO A COMPUTER

The titrator can be connected to a computer using a USB cable. HI900 PC application needs to be installed on the PC.



Connect the cable to the USB port on the rear panel of the titrator.

Connect the cable to the USB port on the PC.

Open the **USB Communication** screen on the titrator (see **General Options** chapter) Launch the **H1900** PC application and then select the appropriate USB Port on the PC.

	USB I	Link wi	th PC	
		Inactive		
Speed 19200				
	Escape			

The HI900 PC application allows the transfer of methods and reports between the titrator and PC.

CHAPTER 12. AUTOSAMPLER

12.1. START UP	
12.2. AUXILIARY COMMANDS	
12.2.1. MANUAL COMMANDS	
12.2.1.1. TRAY	12-5
12.2.1.2. DISPENSER	
12.2.1.3. STIRRER	
12.2.1.4. AUXILIARY PUMPS	
12.2.1.5. BURETTE	
12.2.2. pH CALIBRATION	
12.2.3. RELATIVE mV CALIBRATION	12-7
12.2.4. ISE CALIBRATION	
12.3. AUTOSAMPLER OPTIONS	
12.3.1. SAVE TO USB STORAGE DEVICE	
12.3.2. RESTORE FILES FROM USB STORAGE DEVICE	12-9
12.3.3. ADMINISTRATION	
12.3.4. TOTAL VOLUME ALERT	12-10
12.3.5. TITRANT AGE REMINDER	12-10
12.3.6. TRAY TYPE	12-11
12.3.7. BEAKER DETECTION	12-11
12.3.8. USB LINK WITH PC	12-12
12.3.9. SETUP BALANCE	12-12
12.3.10. RESTORE AUTOSAMPLER SETTINGS	12-12
12.4. AUTOSAMPLER SEQUENCES	12-13
12.4.1. SELECTING A SEQUENCE	12-13
12.4.2. CREATING A SEQUENCE	12-13
12.4.3. DELETING A SEQUENCE	12-14
12.4.4. VIEW/MODIFY A SEQUENCE	12-14
12.4.5. SEQUENCE OPTIONS	12-15
12.4.5.1. SEQUENCE NAME	12-15
12.4.5.2. SEQUENCE REVISION	12-15
12.4.5.3. COMMENTS	
12.4.5.4. STIRRER CONFIGURATION	12-16
12.4.5.5. MISSING BEAKER BEHAVIOR	
12.4.5.6. SAMPLE LEVELING	
12.4.5.6.1 LEVELING PUMP	
12.4.5.6.2 LEVELING TIME	
12.4.5.6.3 DISPENSER HEAD HEIGHT	

AUTOSAMPLER

12.4.5.7. REAGENT ADDITION	<u>12-18</u>
12.4.5.7.1. REAGENT PUMP	
12.4.5.7.2. DISPENSER POSITION	
12.4.5.7.3. DISPENSING TIME	
12.4.5.7.4. STIRRING TIME	
12.4.5.7.5. DISPENSER WAITING POSITION	
12.4.5.7.6. WAIT TIME	
12.4.5.7.7. ADDITION PHASE (LINKED METHODS ONLY)	
12.4.5.8. METHOD	
12.4.5.9. METHOD OPTIONS	
12.4.5.10. DISPENSER POSITION	
12.4.5.11. HEAD UP WAIT TIME	
12.4.5.12. SAMPLE ASPIRATION	
12.4.5.12.1. ASPIRATION PUMP	
12.4.5.12.2. ASPIRATION TIME	
12.4.5.12.3. DISPENSER HEAD HEIGHT	
12.4.5.13. RINSE	
12.4.5.13.1. RINSE BEAKER	
12.4.5.13.2. RINSE TIME	
12.4.5.13.3. DISPENSER POSITION	
12.4.5.13.4. HEAD UP WAIT TIME	
12.4.5.13.5. STIRRER	
12.4.5.14. BEAKER HEIGHT	
12.4.5.15. POSITION WHEN FINISHED	
12.4.5.16. STORAGE BEAKER (POSITION WHEN FINISHED, STORAGE ONLY)	<u>12-27</u>
12.5. SAMPLE TABLE	
12.6. RUNNING THE AUTOSAMPLER	<u>12-29</u>
12.7. REVIEWED RESULTS AND REPORTS	12-29
12.7.1. VIEWING RESULTS FROM THE SAMPLE TABLE	

AUTOSAMPLER

12.1. START UP

Au Se So Tr St

Se Pie

Once the instrument is assembled and installed, follow the steps below to start the titrator and access the autosampler.

- Connect the titrator to a power outlet with the supplied power adapter.
- Connect the autosampler to the titrator using the HI920-933 communication cable.
- Turn on the autosampler then the titrator with the power switches located on the back of each instrument.
- Wait until the titrator finishes the initialization process.
- When prompted, press device to enter the autosampler interface.

The autosampler information screen will be displayed.

If the autosampler has not been detected an X will appear over the autosampler symbol located in the top right corner.

Note: The *HI932* titrator is compatible with the *HI921* and *HI922* autosamplers.

	HI 932 Titrator v1.00	ANNA struments	
	Analog Board 1 Analog Board 2 Pump 1 Pump 2	Potentiometric Potentiometric Burette/Peristaltic Burette/Peristaltic	
	Press DEVICE to en Please press Enten	nter Autosampler menu ^ ■≱	
HI 922 Aut	osampler 🔅	HI 922 (Autosampler 🏾 🌺
<mark>tosampler</mark> rial Number: ftware Version: ay Type: atus:	21153001 v1.0 16 beakers Ready	Autosampler Serial Number: Software Version: Tray Type: Status:	21142001 v1.0 16 beakers Not Connected
quence Name: copy of ase press Enter		Sequence Name: cop Please press Enter	y of Default Sequence
Select Sequence		Select Sequence	

Note: The autosampler can be accessed from the titrator's main screen by pressing device.

The sample table screen will be displayed.

	15:04:28	8 May 24, Defa	<mark>2018</mark> ult Seq	uence	۲
	Last Se	q.: TRAYO	002, 16:1	6 Apr 04,	2018
	# Nai 2 3 5 6 7 8 9 10 11 12 Add Samels	Me 	Size[9] Result	AutoSmp. Setup
<i>i</i> the autosampler's main scr	en press 🔼	utoSmp. Setup			
		0.1N Sodi	ult Seq		۲

To view

15:04:27	May 24, Defa	2018 ult Sequ	vence	
Method: Analog :	0.1N Sodi L	um Hydrox.	ide	
	Pur	ip 1 Selec	ted	
# Nar	ne	Sizelgl	Result	
1				
2				
3				
4				
6				
AutoSmp.	Select	Sequence		Sample
Options	Sequence	Options	Commands	Table

12.2. AUXILIARY COMMANDS

The auxiliary commands menu can be accessed from the main screen by pressing Commands. From this screen you are able to calibrate your electrode and perform manual operations (i.e. running the pumps, moving the tray, etc).

Use the \checkmark and \triangleright to select the analog input to be used for calibration.

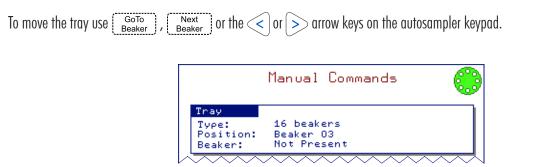
	Auxil	iary Co	mmands	0
Select	the optio	on.		_
	eft> and ∘ t active a		rrows to : ard.	select
Active	Analog I	nput: Ana	alog 1	
Manual Commands	Escape	ρΗ Calibr.	mŲ Calibr.	ISE Calibr.

12.2.1. MANUAL COMMANDS

The manual commands screen is used to manually operate the autosampler dispenser, beaker position, auxiliary pumps, burette and stirrers.

	Manu	al C	omm	ands		
Tray Type: Position: Beaker:	16 b Beak Not		_			
Dispenser Position:	U	₽∎ \$	Sti	rrer		verhead RPM
Auxiliary Pump 1 Present <1>		ent	Pro	imp 3 esent (3)		^P ump 4 Tresent <4>
GoTo Es	ACARS.	Ne> Beak		Buret	te	Magneti Stirre

12.2.1.1. TRAY



Type: Is the tray size currently detected by the autosampler. The tray size can be manually selected, if necessary (see **General Options**). **Position:** Refers to the beaker located under the dispenser.

Beaker: Is the status (present or not present) of the beaker located under the dispenser. Beaker detection can be disabled, if necessary (see **General Options**).

12.2.1.2. DISPENSER

The position (up or down) of the dispensing head will be displayed.

To move the dispenser use the \bigwedge and \bigvee keys on the autosampler or titrator keypad.



12.2.1.3. STIRRER

Use the $\left[\frac{Magnetic}{Stirrer} \right] / \left[\frac{Overhead}{Stirrer} \right]$ to toggle between the stirrer type.

Press stir on the titrator keypad to turn on the stirrer. When active use the << and >> on the titrator keypad to change the stir speed.



12.2.1.4. AUXILIARY PUMPS

To run an active pump press and hold the corresponding number key on the autosampler keypad or titrator keypad. (e.g. press numeric key 1 for auxiliary pump 1, 2 for auxiliary pump 2, etc.).



12.2.1.5. BURETTE

To access the Burette screen, press Burette from the manual commands screen. Highlight the desired option and then press Select See Auxiliary Function, Burette section for additional information.

Burette					
Select	Select a menu option.				
The current pump is: Pump 1 Current burette volume is 25 mL.					
Select	Escape	Choose Pump	Perist.1 On	Perist.2 On	

12.2.2. pH CALIBRATION

From the Auxiliary Commands screen, press <u>
 Calibre</u>
 to view pH calibration screen.
 See pH mode, pH Calibration section for additional information.

Analog 1	PH I	Calibrat 6.90	_	۲
атс 25.0 °	'C	Hanna 7.010		۳۷ 5.76
Hanna 1.679	ated Buffe Hanna 4.010 alibration	Hanna 7.010	Hanna 10.010 1ay 24, 20	Hanna 12.450
Press <clear cal=""> to clear old calibr.</clear>				
Clear <u>Cal</u>	Escape	Edit	Next Buffer	Previous Buffer

12.2.3. RELATIVE mV CALIBRATION

Press $\begin{bmatrix} mV \\ Calibr \end{bmatrix}$ to view mV calibration screen.

See **mV mode**, **Relative mV Calibration** section for additional information.

Relative mV 👸					
Analog 1					
Set th	e value fo	or the rel	lative	mV offse	t.
Abs	olute mV:	3.:	1 mV		
			1	<mark>Uver</mark> 400/1400	
Rel	ative mV:	3.:	i mV		
Low limit: -1996.9 mV					
High limit: 2003.1 mV					
Accept	Escape	Delete Digit			

12.2.4. ISE CALIBRATION

Press ISE to view ISE calibration screen.

See ISE mode, ISE Calibration section for additional information.

Analog 1		Calibra 9.99	9 ^{E-2} PPM	۲
атс 24.8 °	_	SE: Silve E-1 1.00	r	mV 0.0
E-1	ted Standa 1.00 alibratio	10.0	100 Jui 03, 20	1000 018
Press	(Accept)	to update	calibrati	ion.
Accept	Escape	Edit	Next Standard	Previous Standard

12.3. AUTOSAMPLER OPTIONS

The Autosampler Options screen gives access to options that are not directly related to the autosampler sequences. To access this screen, press AutoSmp from the autosampler main screen.

	Autosa	mpler	Opt	io	ns		ę	٢
Select	the menu (option:						
Adminis Titrant Titrant Titrant Titrant	2 from USB stration: 5 1 Volume 5 1 Age Re 5 2 Volume 5 2 Age Re	Alert: minder: Alert:				0	ocked Off days Off days	
USB Lir Setup B	Detection whith PC		ting	18 16 12	bea bea bea to D	ker ker ker	's 's	3
Select	Escape							

12.3.1. SAVE TO USB STORAGE DEVICE

This option allows the user to save files from titrator to a USB storage device.

From the autosampler options, the available file types are:

Standard Method Files	- HIXXXXYY.MTD (e.g.: HI1001EN.MTD, HI1004EN.MTD)
User Method Files	- USERXXXX.MTD (e.g.: USER0001.MTD)
Sequence Files	- SEQXXXX.MTD (e.g.: SEQ0001.MTD)
Autosampler Report Files	- TRAYXXXX.RPT (e.g.: TRAY0001.RPT)

Note: Autosampler Report Files contain the individual titration reports for all samples run on that tray.

Use the \lt and > keys to select the file type. The number of files and the file name on the titrator will be displayed.

Li	st of F	iles on	Titrat	or 👸
	/-> arrow report f:	keys to s iles	select fil	le type
MRAYOU TRAYOU				
		-	-	_
Escape	Copy file	Сору А11	Delete File	Delete All

The option keys allow the following operations:

Delete File Delete All Copy File Copy All Escape

Deletes the highlighted file.

Deletes all currently displayed files.

Copies the highlighted file from the titrator to a USB storage device

Copies all currently displayed files from the titrator to a USB storage device

Returns to the Autosampler Options screen

The status of the transfer ("Successful"/"Unsuccessful") and the file name of the currently processed file are displayed during copying or deleting.

Note: The saved files will be stored on the USB storage device in the H1932 Folder, as follows:

- Methods: USB Drive:\HI932\Methods*.mtd

- Sequences: USB Drive:\HI932\Sequence*.mtd

- Reports: USB Drive:\HI932\ASReport*.rpt

AUTOSAMPLER

12.3.2. RESTORE FILES FROM USB STORAGE DEVICE

This screen allows the user to transfer files from a USB storage device to the titrator. The file types that can be transferred are:

Standard Method Files User Method Files Sequence Files

Autosampler Report Files

- USERXXXX.MTD (e.g.: USER0001.MTD)
 - SEQXXXX.MTD (e.g.: SEQ0001.MTD)

- HIXXXXYY.MTD (e.g.: HI1001EN.MTD, HI1004EN.MTD)

- TRAYXXXX.RPT (e.g.: TRAYOOO1.RPT)

Note: Autosampler Report Files contain the individual titration reports for all samples run on that tray.

Use the < and > keys to select the file type. The number of files and the file name on the titrator will be displayed.

	List o	f Files	on USB	
	/-> arrow y report ·		select fil	le type
WINTY OU TRAYOO TRAYOO TRAYOO TRAYOO TRAYOO TRAYOO TRAYOO TRAYOO TRAYOO TRAYOO TRAYOO TRAYOO TRAYOO TRAYOO	09.RPT 10.RPT 11.RPT 12.RPT 13.RPT 14.RPT 15.RPT 15.RPT 16.RPT 18.RPT 18.RPT 20.RPT			
Escape	Copy file	Сору А11	Delete File	Delete All

The option keys allow the following operations:

Delete File Delete All Copy File Copy All

Deletes the highlighted file from the USB storage device.

Deletes all currently displayed files from the USB storage device.

Copies the highlighted file from the USB storage device to the titrator.

Copies all currently displayed files from the USB storage device to the titrator.

Escape Returns to the Autosampler Options screen

Note: The saved files will be stored on the USB storage device in the HI932 Folder, as follows:

- Methods: USB Drive:\HI932\Methods*.mtd
- Sequences: USB Drive:\HI932\Sequence*.mtd
- Reports: USB Drive:\HI932\ASReport*.rpt

12.3.3. ADMINISTRATION

A 4-digit numeric PIN can be set to prevent unauthorized changes from being made.

See General Options, Administration section for additional information.

	litrator	Admini	stratior	· 🔅
Titrato	r is LOCK	ED.		
	Unlock Ti	trator		
	Enter	PIN: 🍂	**-	
Accept	Escape	Delete Digit		

12.3.4. TOTAL VOLUME ALERT

This screen allows a programmable reminder to appear when the titrant reservoir is below 100 mL. The titrant volume will decrease as the titrant is used.

See General Options, Total Volume Alert section for additional information.

	Titrant	1 Volu	me Alert	- 🛞	
the ti reserv	tration/re oir. The e	eagent sys	ant availa stem from decrease a ted.	its	
		0.0	u mL		
A reminder will appear when less than 100 mLs of titrant volume is left.					
Accept	Escape	Delete Digit		Off	

12.3.5. TITRANT AGE REMINDER

A programmable reminder will appear when it is time to verify the titrant concentration or to change the titrant. See **General Options, Titrant Age Reminder** section for additional information.

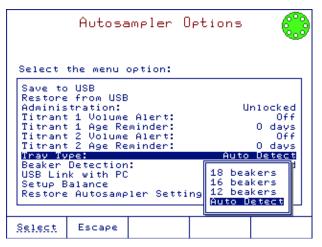
	Titrant	1 Age	Reminder	· 🛞
last Ti	tr. Vol. (updating	to pass si or the las reminder a	st Start
			5 days	
The ran	ge is fro	n O to 31	days.	
Start	Escape	Delete Digit		Off

AUTOSAMPLER

12.3.6. TRAY TYPE

Option: Auto Detect, 18 beakers, 16 beakers or 12 beakers

The autosampler trays have a built in RFID tag that transmits the tray size and serial number directly to the titrator. The tray size can also be selected manually.



12.3.7. BEAKER DETECTION

Option: Enabled or Disabled

The autosampler can detect the presence of a beaker when it is under the dispenser. This prevents titrations from occurring when no beaker is present.

WARNING: If you disable this feature, the autosampler will titrate in any spot on the tray a sample has been entered. Please ensure all beakers are present in the correct positions before starting the sequence. Serious injury could result.

	Autosa	meler	Opt	ions	٩
Select	the menu (option:			
Adminis Titrant Titrant Titrant Tray Ty Beaker USB Lin Setup B	from USB tration: 1 Volume 1 Age Re 2 Volume 2 Age Re 2 Age Re pe: Uetection k with PC	Alert: minder: Alert: minder: :	tings	Auto	locked Off O days Ofs Detect nabled abled
Select	Escape				

12.3.8. USB LINK WITH PC

In order to use this feature, the USB cable needs to be connected from the titrator to the PC. Make sure that the H1900 PC application is running on the PC. See General Options, USB Link with PC section for additional information.

USB Link with PC 🔅						
Inactive						
Speed 19200						
Escape						

12.3.9. SETUP BALANCE

This screen allows the users to connect an analytical balance for automatic acquisition of sample mass prior to titration or standardization.

See General Options, Setup Balance Interface section for additional information.

9	Getup Ba	lance I	nterfac	e 🔅
Select	the balan	ce to be a	activated.	
<mark>× Lab b</mark>	alance			
Disable	Escape	New	Edit	
Balance	Escape	Balance	EUIt	

12.3.10. RESTORE AUTOSAMPLER SETTINGS

This option restores the manufacturer settings for the autosampler interface only! *Note:* This will delete all user created sequences, tray reports, etc.



12.4. AUTOSAMPLER SEQUENCES

All of the parameters required to complete an analysis on the autosampler are grouped into a sequence.

A default sequence is provided; this sequence is used as a starting point for creating user defined sequences. Up to 30 sequences can be created and stored on the titrator.

User defined sequences allow the user to customize reagent additions and rinsing cycles to suit specific applications. New sequences are created in the select sequence screen.

12.4.1. SELECTING A SEQUENCE

To select a sequence, press Sequence from the main screen. A list of available sequences will be displayed.

In the Autosampler Sequence screen, you can view the list of all available sequences.

	Autosampler Sequence	۲
Select	the sequence to be activated:	
SI=000000	1 Default Sequence	
0	New	
Select	Sequence	

To select a sequence, highlight the sequence and press select. The name of the selected sequence will be displayed on the main screen.

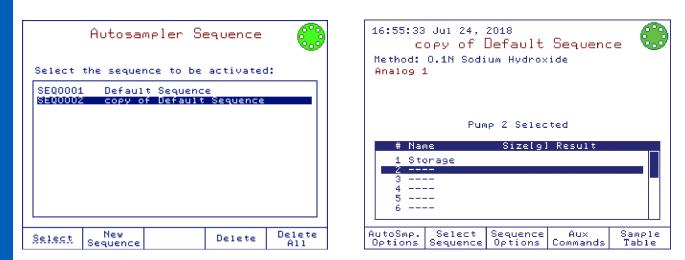
15:10:51 May 24, 2018 Default Sequence Method: 0.1N Sodium Hydroxide Analog 1							
		ip 1 Selec					
# Nar	1e	Sizelgi	Result				
1 Sto	orage						
2							
3							
4							
6							
AutoSmp. Options	Select Sequence	Sequence Options	Aux Commands	Sample Table			

12.4.2. CREATING A SEQUENCE

Sequences are developed by the users in accordance with the analysis requirements. All sequence parameters can be modified by the user.

To create a new sequence, start from the default sequence or a previous created sequence and follow these steps:

- Press Sequence from the main screen.
- Using the \bigwedge and \bigtriangledown keys, highlight an existing sequence from the list.
- Press $\underbrace{\mathbb{N}_{\text{Sequence}}^{\text{New}}}_{\text{Sequence}}$. A new sequence will be generated.
- Press Select to activate the new sequence.



12.4.3. DELETING A SEQUENCE

Unnecessary sequences can be removed from the titrator. To remove a sequence press sequence from the main screen then highlight the sequence you want to delete and press below the sequence will appear in order to confirm the deletion. Press delete again to confirm, or press escape to cancel the operation.

Confi	rmation of Method Deletion	
	u sure you want to delete the ed method?	
COPY O	f 0.1N Sodium Hydr	
Delete	Escape	

12.4.4. VIEW/MODIFY A SEQUENCE

To modify the sequence options, press $\underline{Sequence}_{Options}$ from the main screen. A list of all the parameters for the selected sequence will be displayed. Using the \bigwedge and \bigvee keys, highlight the option you want to modify and press \underline{Select} .

I	Jiew /	Modify 9	Bequence	•			
Id: SEQ0002 Modified: 16:54 Jul 24, 2018 Select the option to be modified.							
Sequence Name: copy of Default Sequence Revision Number:1.0Comments:0verheadStirrer Type:0verheadMissing Beaker Behavior:StopSample Leveling:DisabledReagent Addition 1:DisabledReagent Addition 2:DisabledMethod: HI0001EN 0.1N Sodium HydroxideMethod Options:140 mmHead Up Wait Time:1 sec							
Select	Escape	Print Sequence	Page Up	Page Down			

To exit the View/Modify Sequence screen press [Escape]

You can choose to save the modifications or to discard them.

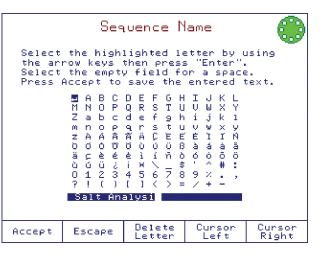
	Savi	ng Seq	vence	۲		
Select	a menu op	tion.				
<mark>Save Se</mark> Exit Wi	quence thout Sav	ing Sequ	ence			
"Escape" - exit without saving sequence.						
Select	Escape					

12.4.5. SEQUENCE OPTIONS

All of the parameters required to complete an analysis are grouped into a sequence. The sequence options screen is arranged as they occur during the sequence.

12.4.5.1. SEQUENCE NAME

Option: Up to 24 characters



12.4.5.2. SEQUENCE REVISION

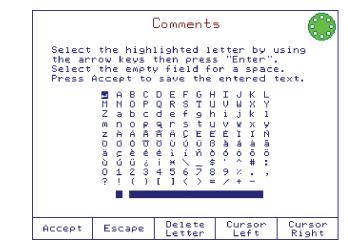
Option: Up to 3 characters

Revision Number 💮						
Select the highlighted letter by using the arrow keys then press "Enter". Select the empty field for a space. Press Accept to save the entered text.						
	M N O P Z a b c m n o p 7 A A A	D E F G H Q R S T U d e f 9 h Q A C E E 0000000000000000000000000000000000	U W X Y i j k 1 V W X Y E i i a a a a a a a a a a a a a a a a a a			
Accept	Escape	Delete Letter	Cursor Left	Cursor Right		

12.4.5.3. COMMENTS

AUTOSAMPLER

Option: Up to 20 characters



12.4.5.4 STIRRER CONFIGURATION

Option: Overhead or Magnetic

View / Modify Sequence 🧃)
Id: SEQ0002 Modified: 16:54 Jul 24, 201 Select the option to be modified.	18
Sequence Name: copy of Default Sequence Revision Number: 1.0 Comments: Sequence Missing Beaker Behavior: Sample Leveling: Uverhead Reagent Addition 1: Magnetic Method: HIO001EN 0.1N Sodium Hydroxide Method Options: 140 mm Head Up Wait Time: 1 sec	
<u>Select</u> Escape	

12.4.5.5. MISSING BEAKER BEHAVIOR

Option: Pause, Skip or Stop

Select the behavior to occur when beaker detection is enabled and no beaker is detected.

	View /	Modify	Sequenc	e 💮
)002 Mc the optior			24, 2018
Revisio Commen Stirren Missin Sample Reagen Reagen Method Dispen:		Sehavior: 1: 2: 1 0.1N So	Ov Si dium Hy B	quence 1.0 erhead Stop d use d top 140 mm 1 sec
Select	Escape			

Pause: The autosampler will pause the sequence at the current beaker and wait for the user before continuing the analysis.Skip: The autosampler will automatically move to the next available sample.

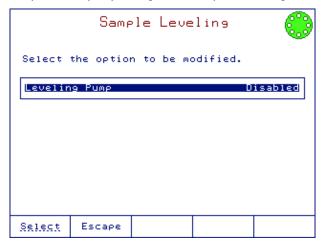
Stop: All operations will stop and the analysis will be stopped.

12.4.5.6. SAMPLE LEVELING

Option: Disabled or Enabled

Volumetric samples that do not require high accuracy can be leveled to the correct volume rather than manually being dispensed with a pipette. Sample leveling involved the user adding excess sample to each beaker, and the autosampler removed the excess sample using an aspiration tube. This allows samples to be quickly poured into each beaker by the user while the autosampler accurately removes excess.

Note: Sample leveling requires one peristaltic pump configured for aspiration using H1920-203 Tubing Set for aspiration.



12.4.5.6.1. LEVELING PUMP

Select the peristaltic pump that is connected to the aspiration tube.

Sample Leveling 🔅	Sample Leveling 🔅
Select the option to be modified.	Select the option to be modified.
Leveling Pump Disabled Aux Pump 1 Aux Pump 2 Aux Pump 3	Leveling Pump Aux Pump 1 Leveling Time 10 sec Head Height 120 mm
Select Escape	Select Escape

12.4.5.6.2. LEVELING TIME

Option: 1 second to 300 seconds

Set the duration that the peristaltic pump will run.

	Leu	eling I	ime	
	he period ry pump.	of time H	for runnir	9
		1	J o sec	
	mit: 1 : imit: 30)			
Accept	Escape	Delete Digit		

12.4.5.6.3. DISPENSER HEAD HEIGHT

Option: 10 to 150 mm

Set the height for the dispenser head. This height should be set to produce the sample volume that is defined within method options. The correct height must be determined experimentally by the user and will depend on the sample size, beaker shape and size, and the aspiration tube position.

The easiest way to determine the volume of a particular height setting is to manually aspirate water from a pre-weighed beaker and weighing the remaining water in the beaker.

	Prese	t Head H	leight	۲	
the hea	d to appr eric keys	opriate po	to positi osition, d lly enter	on	
			U mm		
The range is from 10 to 150 mm. press (Accept) to save the head position.					
Accent	Escape		Δ	∇	

12.4.5.7. REAGENT ADDITION

Option: Disabled or Enabled

Reagents and/or deionized water can be automatically added to each sample using the reagent addition feature.

Note: Reagent addition required a peristaltic pump (for each reagent) configured for dispensing using **H1920-208** Tubing Set for Dispensing. The **H1922** can perform up to two reagent additions.

12.4.5.7.1. REAGENT PUMP

Select the peristaltic pump that is connected to the reagent container.

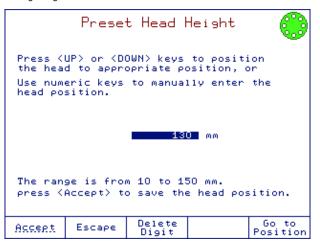
	Rease	nt Addi	tion 1	۲
Select	the option	n to be mo	odified.	
Reagent	: Pump:			Disabled
			Aux F Aux F Aux F	Pump 1 Pump 2 Pump 3
Select	Escape			

	Reage	nt Addi	tion 1	۲
Select	the option	n to be m	odified.	
Dispens	ser Positi sing Time: ng Time: ser Waitin			Pump 1 130 mm 1 sec 0 sec Down 0 sec
Select	Escape			

12.4.5.7.2. DISPENSER POSITION

Option: 10 to 150 mm

Enter the position of the dispenser during reagent addition and stir time.



12.4.5.7.3. DISPENSING TIME

Option: 1 to 300 seconds

Enter the dispensing time required to add the desired amount of reagent.

Note: This time should be determined experimentally. The approximately flow rate it 200 mL/min.



12.4.5.7.4. STIRRING TIME

Option: 0 to 1800 seconds



12.4.5.7.5. DISPENSER WAITING POSITION

Option: Up or Down

Set the position of the dispenser during the wait time. This is useful if it is undesirable for the electrode(s) to be immersed in the solution for extended periods of time.

	Reagent	Addi	tion	1	۲
Select	the option t	o be ma	odifie	d.	
Dispens Stirrir	er Position: ing Time: g Time: er Waiting P	ositio	n:	Aux	Pump 1 130 mm 1 sec 0 sec Down C
Select	Escape				

12.4.5.7.6. WAIT TIME

Option: 0 to 1800 seconds

Set the reaction time. This is the amount of time after the stirring is completed that the autosampler will wait before performing any other actions.

	h	Jait Tim	ne	۲
Please	enter the	wait time	e in seconds.	
			2 sec	
	mit: 0 : imit: 180		5	
Accept	Escape	Delete Digit		

12.4.5.7.7. ADDITION PHASE (LINKED METHODS ONLY)

Option: First Titration or Second Titration

Set the addition phase for the reagent addition. Reagent addition can be done before the first titration or before the second titration.

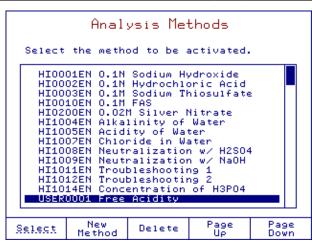
	Reager	nt Addi	tion	1	۲
Select	the option	n to be mo	odifie	d.	
Dispens Stirrin Dispens Wait Ti	er Positi ing Time: g Time: er Waitin	Fir	st Ti ond Ti	trat itra	Pump 1 130 mm c tion c ration
Select	Escape				

AUTOSAMPLER

12.4.5.8. METHOD

The following types of methods can be run on the autosampler:

	Sample Titration (Single End point)
No. Polod	Titrant Standardization
Non-Linked Methods	Back Titration
Memous	Sample Titration (Multi EQ points)
	Direct Reading
	Sample Titration (Single End point) \rightarrow Sample Titration (Single End point)
	Sample Titration (Single End point) \rightarrow Sample Titration (Multi EQ points)
	Sample Titration (Single End point) \rightarrow Direct Reading
	Back Titration → Direct Reading
	Sample Titration (Multi EQ points) \rightarrow Sample Titration (Single End point)
Linked Methods	Sample Titration (Multi EQ points) 🛛 → Sample Titration (Multi EQ points)
	Sample Titration (Multi EQ points) → Direct Reading
	Direct Reading \rightarrow Sample Titration (Single End point)
	Direct Reading -> Back Titration
	Direct Reading 🛛 → Sample Titration (Multi EQ points)
	Direct Reading → Direct Reading



12.4.5.9. METHOD OPTIONS

The analysis method options can be accessed directly from the autosampler interface. Analysis method options can be reviewed and/or modified if necessary.

For more information see Titration Methods, Method Options section.

			15:22 May	24, 2018
Analog Titran Reagen Dosing End Po Recogn Pre-Ti Stirri Measur	int Mode: ition Opt tration V tration S ng Speed:	n 1: n 2: mV 1EQ ions olume: tir Time:	Pu Disa Disa Dyr Point,1st 9.00 10	1.0 Log 1 Jmp 1 abled abled t Der 0 mL 0 RPM
Select	Escape	Print Method	Method 2	Page Down

12.4.5.10. DISPENSER POSITION

Option: 10 to 150 mm

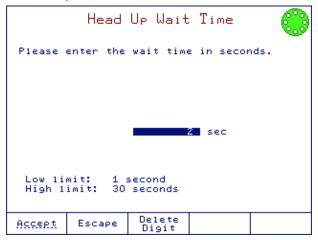
Enter the position of the head during titration (140 mm by default).

	Preset Head Height 🔅					
Press (UP) or (DOWN) keys to position the head to appropriate position, or Use numeric keys to manually enter the						
head po		vo Manda.	riy chiver	one.		
	140 mm					
The range is from 10 to 150 mm. press 〈Accept〉 to save the head position.						
Accept	Escape	Delete Digit		Go to Position		

12.4.5.11. HEAD UP WAIT TIME

Option: 1 to 30 seconds

Set the duration that the autosampler will wait with the dispenser in the up position for any drops of solution to fall off of the electrodes/stirrer before moving to another sample or rinse beaker.



12.4.5.12. SAMPLE ASPIRATION

Option: Disabled, Aspirate Only or Aspirate/Spray Rinse

Reacted samples may be aspirated into a waste container after each titration.

Note: Sample aspiration requires one peristaltic pump configured for aspiration using H1920-203 Tubing Set for Aspiration.

Aspirate Sample 🔅						
Select	the option	n to be m	odified.			
Aspirat	ion Optio	n:		Jisabled		
			ed ate Only ate/Spray	Rinse		
Select	Escape					

Select the aspiration mode:

Aspiration Only: The existing waste from the sample beaker will be removed according to the parameters defined in this menu.

Aspirate/Spray: Reserved for future

12.4.5.12.1. ASPIRATION PUMP

Select the peristaltic pump that is connected to the aspiration tube.

Aspirate S	ample 🔅
Select the option to be a	nodified.
Aspiration Option: Aspiration Pump: Aspiration Time: Head Height:	Aspirate Only Disabled Aux Pump 1 Aux Pump 2 Aux Pump 3
<u>Select</u> Escape	

12.4.5.12.2. ASPIRATION TIME

Option: 1 to 300 seconds

Set the duration that the peristaltic pump will run.

	Aspiration Time 💮					
	he period ry pump.	of time (for runnir	19		
			sec 🖥			
Low limit: 1 second High limit: 300 seconds						
Accept	Escape	Delete Digit				

12.4.5.12.3. DISPENSER HEAD HEIGHT

Option: 10 to 150 mm

Set the height for the dispenser head. The aspiration tube should be set to a height that will reach the bottom of the sample beaker when the dispenser head is positioned at this height.

Preset Head Height 🔅						
Press (UP) or (DOWN) keys to position the head to appropriate position, or Use numeric keys to manually enter the head position.						
	140 mm					
The range is from 10 to 150 mm. press (Accept) to save the head position.						
Accept	Escape	Delete Digit		Go to Position		

12.4.5.13. RINSE

The autosampler can perform a dip rinse function after each analysis. Up to 3 dip rinses can be performed in a dedicated rinse beakers.

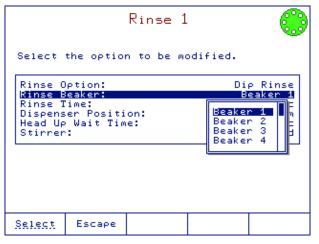
Rinse 1 🔅				
Select	the optio	n to be	modified.	
Rinse (Option:			Disabled
Select	Escape			

Dip rinse: Dip rinse can be used to clean the electrodes and stirrer of contaminants after each analysis using dedicated rinsing beakers.

Spray Rinse: Reserved for future

12.4.5.13.1. RINSE BEAKER

Select the position on the tray for the dedicated rinse beaker.



12.4.5.13.2. RINSE TIME

Option: 1 to 300 seconds

	Rinse Time 💮						
		of time y rinse bea		like			
		1	J sec				
	mit: 1 : imit: 30)						
Accept	Escape	Delete Digit					

12.4.5.13.3. DISPENSER POSITION

Option: 10 to 150 mm

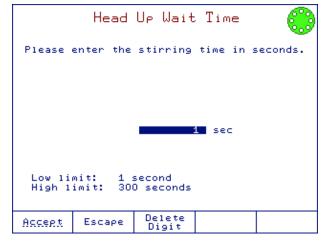
Set the height for the dispenser head during rinsing.

Preset Head Height 🔅					
the hea	Press <up> or <down> keys to position the head to appropriate position, or Use numeric keys to manually enter the</down></up>				
head po		to Marida.	ny enter	one	
140 mm					
The range is from 10 to 150 mm. press (Accept) to save the head position.					
Accept	Escape	Delete Digit		Go to Position	

12.4.5.13.4. HEAD UP WAIT TIME

Option: 1 to 300 seconds

Set the duration that the autosampler will wait with the dispenser in the up position for any drops of solution to fall off of the electrodes/stirrer before moving to another sample or rinse beaker.



12.4.5.13.5. STIRRER

Option: Enabled or Disabled

Select if the stirrer will run during the rinse operation.

	Rinse			۲
Select	the option	n to be m	odified.	
Rinse E Rinse T Dispens Head Up	Rinse Option: Rinse Beaker: Rinse Time: Dispenser Position: Head Up Wait Time: Stirrer:			P Rinse eaker 1 10 sec 140 mm 1 sec inabled abled
Select	Escape			

12.4.5.14. BEAKER HEIGHT

Option: 30 to 120 mm

Set the height of the beaker being used on the autosampler.

Beaker Height 👸				۲
Enter the beaker height in mm.				
		10	U mm	
Low limit: 30 mm High limit: 120 mm				
Accept	Escape	Delete Digit		

AUTOSAMPLER

12.4.5.15. POSITION WHEN FINISHED

Option: Home, Sample or Storage

•				
l	Jiew /	Modify S	Gequence	•
		dified: 1 to be mo	1:48 Jul dified.	24, 2018
Method: Linked Dispens Head Up Aspirat Rinse 1 Rinse 2 Rinse 3 Beaker	To: USER Options: er Positi Wait Tim e Sample: :	USER0003 20004, Dir .on: .e:	, Free Ac ect pH re Asf Home Samp Stor	ading 40 mm 1 sec 4 4 4 1 e
Select	Escape			

Home: The dispenser head will be in the up position over beaker one.

Sample: The dispenser head will remain down in the last sample that was analyzed/titrated.

Storage: The dispenser head will be down in a preset beaker containing storage solution.

12.4.5.16. STORAGE BEAKER (POSITION WHEN FINISHED, STORAGE ONLY)

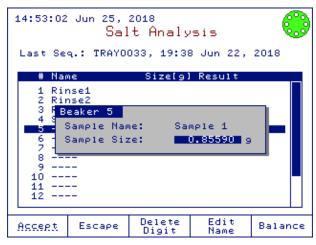
Select the storage beaker. After the sequence has been completed, the autosampler will move to this position automatically and lower the dispenser head.

	View /	Modify 9	Bequence	- (9
Id: SEQ0002 Modified: 11:50 Jul 24, 2018 Select the option to be modified.					
Method Dispen: Head U Aspira Rinse Rinse Beaker Positi	To: USER Options: ser Positi Wait Tin te Sample: 1: 2:	ROOO4, Dir Lon: Me:	1 Beaker Beaker Beaker Beaker	40 mm 1 sec 1 1 2 1	
Select	Escape				

12.5. SAMPLE TABLE

All sample information is entered into the sample table according to the tray position. The sample table screen is the default screen when entering the autosampler interface while the autosampler is idle. The sample table is automatically formatted with the appropriate number of beaker, with rinse/storage beaker positions reserved.

To add a sample to the sample table, highlight an empty beaker position using the \bigwedge and \bigvee keys, then press $\frac{Add}{Sample}$ to open the sample dialog box. The user can then edit the sample name and size.



- Press Accept to enter the current sample name and size into the sample table.
- Press Escape to cancel the sample size entry.
- Press Delete Digit to modify the sample size entry
- Press Edit Name to modify the sample name entry
- Press Balance to access the balance interface for direct entry of sample weight (if available).

Note: Several features have been added to make sample entry faster, depending on your peripheral connections and analysis method. The following are available while an empty table position is highlighted.

- <u>Shortcut to name entry</u>: Typing a sample name using an external keyboard will automatically edit the name if the first character is non-numeric.
- <u>Shortcut to size entry</u>: Typing a sample size using the keypad or external keyboard will automatically edit the sample size. The sample name will be auto-incremented.
- <u>Auto-incrementing name</u>: The default sample name is an auto-increment of the previous sample name.
- Barcode reader: Scanning a barcode with a USB barcode reader automatically enters the barcode into the sample name field.
- <u>Fixed sample size</u>: The sample dialog box is omitted if the sample size entry is set to "Fixed". Typing anything from the keypad or external keyboard will go directly to the Edit Name screen.
- <u>Autofill</u> (Fixed sample size): All empty sample table positions can be automatically filled by pressing [Auto name will be auto-incremented.

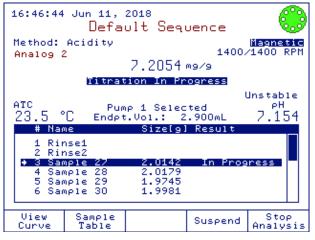
12.6. RUNNING THE AUTOSAMPLER

The autosampler sequence can be started by pressing the $\begin{bmatrix} start \\ stop \end{bmatrix}$ key.

The autosampler will process each sample according to the settings in sequence options.

While the autosampler is running, the top part of the screen shows titration information for the current titration, and the bottom part of the screen shows a portion of the sample table.

The sample in progress is marked with • symbol in the sample table.



If the selected titration method is linked, the linked method will be displayed below the sample when it is in progress.

16:51:56	Jun 11, Defa	2018 ult Sequ	Jence	۲
Method: LinkedTo Analog 2		,].01242;		M <mark>agnetic</mark> ⁄1400 RPM Og)
		ion In Pr		
атс 23.6 °	°C Endet	np 2 Selec .Vol.: O Size[g]		_{рн} 8.476
♦ ¥ Sar 4 Sar		2.0145 2.0145 2.0145 1.9851		iress
View Curve	Sample Table		Suspend	Stop Analysis

- Use the \bigwedge and \bigvee keys to scroll the sample table.
- Press View Curve to view the graph of the current titration.
- Press sample Table to view/modify the sample table entries. Sample can be added to the sample table while the autosampler is running.
- Press Suspend to pause the current titration.
- Press Stop Analysis
 to end the current titration immediately and preceed to the next sample.

12.7. REVIEWED RESULTS AND REPORTS

Results for the current or most recent tray of samples are shown directly in the sample table.

The results of all autosampler titrations can be accessed using the results key.

12.7.1. VIEWING RESULTS FROM THE SAMPLE TABLE

The results of each titration is shown directly in the sample table once the sample has been completed. For more information on these reports press sample reports press highlight the desired sample and press view Results.

APPENDIX 1. TECHNICAL SPECIFICATIONS

A1 HI932 TECHNICAL SPECIFICATIONS	A1-3
A2 HI922 TECHNICAL SPECIFICATIONS	

A1. HI932. TECHNICAL SPECIFICATIONS

	Standard Titrati	on (Standardization, Fixed pH/ mV, Equivalence Point pH/ mV)
Analysis Type	Back Titration	
	Direct Reading	
	Fixed mV	
End Point Mode	Fixed pH	
	mV Equivalence	Point (up to 5 points, 1 st or 2 nd derivate)
	pH Equivalence	Point (up to 5 points, 1 st or 2 nd derivate)
	Size	5 mL/10 mL/25 mL/50 mL
	Resolution	0.001 mL
	Flow Rate	0.3 mL to 2 x Burette volume per minute
Burette		\pm 0.005 mL (5 mL Burette)
	A	\pm 0.010 mL (10 mL Burette)
	Accuracy	\pm 0.025 mL (25 mL Burette)
		\pm 0.050 mL (50 mL Burette)
Cu:	Range	200 to 2500 RPM
Stirrer	Resolution	100 RPM
	Range	-2000.0 to 2000.0 mV
14	Resolution	0.1 mV
mV	Accuracy	\pm 0.1 mV
	Calibration	Single point offset
	Range	-2.000 to 20.000 pH
-11	Resolution	0.1/0.01/0.001 pH
рН	Accuracy	± 0.001 pH
	Calibration	Up to 5 points with standard or custom buffers
	Range	1x10 ⁻⁶ to 9.999x10 ¹⁰
ICE	Resolution	1/0.1/0.01
ISE	Accuracy	± 0.001 pH
	Calibration	Up to 5 points
		-5.0 to 105 °C
	Range	23.0 to 221.0 °F
Temperature	-	268.2 to 378.2 K
	Resolution	0.1 °C/0.1 °F/0.1 K
	Accuracy	\pm 0.1 °C/ \pm 0.2 °F/ \pm 0.1 K
Data Storage	Mathada	up to 100 titration methods (standard and user)
	Methods	up to 30 autosampler sequences
	Decit	up to 100 titration and pH/mV/ISE reports
	Reports	up to 40 autosampler tray reports (e.g. 720 reports for 18 beaker tray)

		1 x BNC Socket (pH, ORP, ISE half-cell and ISE combination electrodes)
	Measurement	1 x 4 mm Banana Socket (reference electrode)
	(per analog board)	1 x RCA Socket (temperature sensor)
	boundy	1 x 6-pin Connector (stirrer)
Connections		1 x 6-pin Mini DIN (external PC keyboard)
		1 x DB-25 Socket (printer)
	Peripheral	1 x USB Standard B (PC connection)
		1 x DB-9 Socket (analytical balance)
		1 x USB Standard A (USB flash drive)
		4 x multi-purpose slots (titrant/reagent tubes)
	Electrode Holder	3 x 12-mm electrode slots
	Electrode Holder	1 x temperature sensor slot
		1 x overhead stirrer slot
	Display	5.7" graphical color display with backlight
	Languages	English, Portuguese, Spanish
	Power Supply	100-240 Vac, 50/60 Hz
Additional	Power Draw	0.5 Amps
Specifications	Enclosure Material	ABS, PC and Stainless Steel
	Keypad	Polyester
	Dimensions	315 x 205 x 375 mm (12.4 x 8.1 x 14.8 ")
	Weight	approx. 4.3 kg (9.5 lbs.) with 1 pump, stirrer and sensors
	Operating Environment	10 to 40 °C (50 to 104 °F); up to 95 % RH
	Storage Environment	-20 to 70 °C (-4 to 158 °F); up to 95 % RH

A2. HI922. TECHNICAL SPECIFICATIONS

	5 x multi-purpose slots (titrant/reagent tubes)
	3 x 12-mm electrodes slots
Electrode Holder	1 x overhead stirrer slot
	1 x temperature sensor slot
	1 x aspiration tube slot
Cu:	magnetic stirrer (built-in)
Stirrer	overhead stirrer (optional)
Temperature Sensor	HI7662-AW (included)
Peristaltic Pumps	Up to three (Slots 1, 2 & 3)
Diaphragm Pumps	One (Slot 4)
Peripheral Units	USB Barcode Reader
Turus	16 beakers x 150 mL with Built-in RFID
Trays	18 beakers x 100 mL with Built-in RFID
	ASTM short-form glass beakers, 100 & 150 mL
Beakers	HI920-060 (150 mL), Plastic beakers
	HI920-053 (100 mL), Plastic beakers
	Buttons for manual operation of tray
Control Panel	Manual operation of peristaltic or diaphragm pumps
	2-line backlight display with status information
Enclosure Material	ABS plastic and steel
Electrode Holder Material	ABS plastic
Tray Material	ABS plastic and acrylic
Keypad Material	ABS plastic and polycarbonate
Weight	approx. 13 kg (29 lbs)
Operating Environment	10 to 40°C, up to 95% relative humidity
Storage Environment	-20 to 70°C, up to 95% relative humidity

APPENDIX 2. ACCESSORIES

A2.1. SOLUTIONS
A2.1.1. pH BUFFERS
A2.1.2. pH BUFFERS IN FDA APPROVED BOTTLE
A2.1.3. pH TECHNICAL BUFFERS
A2.1.4. pH MILLESIMAL BUFFERS
A2.1.5. ELECTRODE CLEANING SOLUTIONS
A2.1.6. ELECTRODE CLEANING SOLUTIONS IN FDA APPROVED BOTTLE
A2.1.7. ELECTRODE STORAGE SOLUTIONS
A2.1.8. ELECTRODE STORAGE SOLUTIONS IN FDA APPROVED BOTTLE
A2.1.9. ELECTRODE REFILL ELECTROLYTE SOLUTIONS
A2.1.10. ELECTRODE REFILL ELECTROLYTE SOLUTIONS IN FDA APPROVED BOTTLE
A2.1.11. ORP PRETREATMENT SOLUTIONS
A2.1.12. TITRATION REAGENTS
A2.1.13. ION SELECTIVE ELECTRODE CALIBRATION STANDARDS
A2.2 SENSORS
A2.1.1. pH ELECTRODES
A2.1.2. ORP ELECTRODES
A2.1.3. HALF-CELL ELECTRODES
A2.1.4. ION SELECTIVE ELECTRODES
A2.1.5. TEMPERATURE SENSOR
A2.3. TITRATOR COMPONENTS
A2.4. AUTOSAMPLER COMPONENTS

ACCESSORIES

A2.1. SOLUTIONS

A2.1.1. pH CALIBRATION BUFFERS

HI7001M	pH 1.68 Buffer Solution, 230 mL
HI7001L	pH 1.68 Buffer Solution, 500 mL
HI7004M	pH 4.01 Buffer Solution, 230 mL
HI7004L	pH 4.01 Buffer Solution, 500 mL
HI7006M	pH 6.86 Buffer Solution, 230 mL
HI7006L	pH 6.86 Buffer Solution, 500 mL
HI7007M	pH 7.01 Buffer Solution, 230 mL
HI7007L	pH 7.01 Buffer Solution, 500 mL
HI7009M	pH 9.18 Buffer Solution, 230 mL
HI7009L	pH 9.18 Buffer Solution, 500 mL
HI7010M	pH 10.01 Buffer Solution, 230 mL
HI7010L	pH 10.01 Buffer Solution, 500 mL

A2.1.2. pH CALIBRATION BUFFERS IN FDA APPROVED BOTTLE

H18004L	pH 4.01 Buffer Solution, 500 mL
H18006L	pH 6.86 Buffer Solution, 500 mL
H18007L	pH 7.01 Buffer Solution, 500 mL
HI8009L	pH 9.18 Buffer Solution, 500 mL
H18010L	pH 10.01 Buffer Solution, 500 mL

A2.1.3. pH TECHNICAL CALIBRATION BUFFERS

HI5016	pH 1.68 Buffer Solution, 500 mL
HI5003	pH 3.00 Buffer Solution, 500 mL
HI5004	pH 4.01 Buffer Solution, 500 mL
HI5068	pH 6.86 Buffer Solution, 500 mL
HI5007	pH 7.01 Buffer Solution, 500 mL
HI5091	pH 9.18 Buffer Solution, 500 mL
HI5010	pH 10.01 Buffer Solution, 500 mL
HI5124	pH 12.45 Buffer Solution, 500 mL

A2.1.4. pH MILLESIMAL CALIBRATION BUFFERS

HI6016	pH 1.679 Buffer Solution, 500 mL
HI6016-01	pH 1.679 Buffer Solution, 1 L
HI6003	pH 3.000 Buffer Solution, 500 mL
HI6003-01	pH 3.000 Buffer Solution, 1 L
HI6004	pH 4.010 Buffer Solution, 500 mL
HI6004-01	pH 4.010 Buffer Solution, 1 L
HI6068	pH 6.862 Buffer Solution, 500 mL
HI6068-01	pH 6.862 Buffer Solution, 1 L

HI6007	pH 7.010 Buffer Solution, 500 mL
HI6007-01	pH 7.010 Buffer Solution, 1 L
HI6091	pH 9.177 Buffer Solution, 500 mL
HI6091-01	pH 9.177 Buffer Solution, 1 L
HI6010	pH 10.010 Buffer Solution, 500 mL
HI6010-01	pH 10.010 Buffer Solution, 1 L
HI6124	pH 12.450 Buffer Solution, 500 mL
HI6124-01	pH 12.450 Buffer Solution, 1 L

A2.1.5. ELECTRODE CLEANING SOLUTIONS

HI7061M	General Purpose Solution, 230 mL
HI7061L	General Purpose Solution, 500 mL
HI7073M	Protein Cleaning Solution, 230 mL
HI7073L	Protein Cleaning Solution, 500 mL
HI7074M	Inorganic Cleaning Solution, 230 mL
H17074L	Inorganic Cleaning Solution, 500 mL
HI7077M	Oil & Fat Cleaning Solution, 230 mL
HI7077L	Oil & Fat Cleaning Solution, 500 mL

A2.1.6. ELECTRODE CLEANING SOLUTIONS IN FDA APPROVED BOTTLE

HI8061M	General Purpose Solution, 230 mL
HI8061L	General Purpose Solution, 500 mL
HI8073M	Protein Cleaning Solution, 230 mL
HI8073L	Protein Cleaning Solution, 500 mL
HI8077M	Oil & Fat Cleaning Solution, 230 mL
HI8077L	Oil & Fat Cleaning Solution, 500 mL

A2.1.7. ELECTRODE STORAGE SOLUTIONS

H170300M	Storage Solution, 230 mL
H170300L	Storage Solution, 500 mL

A2.1.8. ELECTRODE STORAGE SOLUTIONS IN FDA APPROVED BOTTLE

HI80300M	Storage Solution, 230 mL
HI80300L	Storage Solution, 500 mL

A2.1.9. ELECTRODE REFILL ELECTROLYTE SOLUTIONS

HI7071	3.5M KCl + AgCl Electrolyte, 30 mL, for single junction electrodes
HI7072	1 M KNO ₃ Electrolyte, 30 mL
HI7075	KNO ₃ and KCI Electrolyte, 30 mL
HI7076	1M NaCl Electrolyte, 30 mL
HI7078	$(NH_4)_2SO_4$ Electrolyte, 30 mL
HI7082	3.5M KCl Electrolyte, 30 mL, for double junction electrodes

A2.1.10. ELECTRODE REFILL ELECTROLYTE SOLUTIONS IN FDA APPROVED BOTTLE HI8071 3.5M KCl + AgCl Electrolyte, 30 mL, for single junction electrodes HI8072 1M KNO₃ Electrolyte, 30 mL HI8082 3.5M KCl Electrolyte, 30 mL, for double junction electrodes A2.1.11. **ORP PRETREATMENT SOLUTIONS** HI7091M Reducing Pretreatment Solution, 230 mL HI7091L Reducing Pretreatment Solution, 500 mL HI7092M Oxidizing Pretreatment Solution, 230 mL HI7092L Oxidizing Pretreatment Solution, 500 mL **TITRATION REAGENTS** A2.1.12. HI70429 0.05 M AgNO₃ Titration Reagent, 1 L HI70433 0.01 N Stabilized Iodine Titration Reagent, 1 L HI70439 0.1 M Na₂S₂O₂Titration Reagent, 1 L 0.02 N Stabilized Iodine Titration Reagent, 1 L HI70440 HI70441 0.04 N Stabilized Iodine Titration Reagent, 1 L HI70448 0.02 M AgNO₃ Titration Reagent, 1 L HI70449 0.02 M EDTA Titration Reagent, 1 L HI70455 0.01 N NaOH Titration Reagent, 1 L HI70456 0.1 N NaOH Titration Reagent, 1 L HI70457 1 N NaOH Titration Reagent, 1 L HI70458 0.01 M H₂SO₄ Titration Reagent, 1 L HI70459 0.05 M H₂SO, Titration Reagent, 1 L HI70462 0.01 N HCl Titration Reagent, 1 L HI70463 0.1 N HCl Titration Reagent, 1 L HI70464 1 N HCl Titration Reagent, 1 L ION SELECTIVE ELECTRODE CALIBRATION STANDARDS A2.1.13. HI4001-01 0.1 M Ammonia Standard HI4001-02 100 ppm Ammonia Standard (as N) HI4001-03 1000 ppm Ammonia Standard (as N) 0.1 M Bromide Standard HI4002-01 HI4003-01 0.1 M Cadmium Standard HI4004-01 0.1 M Calcium Standard HI4005-01 0.1 M Carbon Dioxide Standard 1000 ppm Carbon Dioxide Standard (as CaCO₃) HI4005-03 HI4007-01 0.1 M Chloride Standard HI4007-02 100 ppm Chloride Standard HI4007-03 1000 ppm Chloride Standard HI4008-01 0.1 M Cupric Standard

HI4010-01	0.1 M Fluoride Standard
HI4010-02	100 ppm Fluoride Standard
HI4010-03	1000 ppm Fluoride Standard
HI4011-01	0.1 M Iodide Standard
HI4012-01	0.1 M Lead Standard
HI4012-21	0.1 M Sulfate Standard
HI4013-01	0.1 M Nitrate Standard
HI4013-02	100 ppm Nitrate Standard
HI4013-03	1000 ppm Nitrate Standard
HI4014-01	0.1 M Potassium Standard
HI4015-01	0.1 M Silver Standard

A2.2. SENSORS

A2.2.1. pH ELECTRODES

HI1043B

Glass-body, double junction, refillable, combination pH electrode.

Use: strong acid and base, paint and solvents

HI1053B

Glass-body, triple ceramic, conic shape, refillable, combination pH electrode. Use: emulsions, fats and creams, soil and semi-solids samples

HI1083B

Glass-body, micro, Viscolene, nonrefillable, combination pH electrode. Use: biotechnology and micro titration

HI1131B

Glass-body, double junction, refillable, combination pH electrode.

Use: general purpose

HI1330B

Glass-body, semimicro, single junction, refillable, combination pH electrode. Use: laboratory, vials, and test tubes

HI1331B

Glass-body, semimicro, single junction, refillable, combination pH electrode.

Use: flasks

HI1230B

 $\label{eq:Plastic-body} \ensuremath{\left(\mathsf{PEI}\right)}\xspace, \ens$

Use: general purpose

HI2031B

Glass-body, conical tip, refillable, combination pH electrode.

Use: dairy and semi-solid products

HI1332B

Plastic-body (PEI), double junction, refillable, combination pH electrode.

Use: chemicals, field applications and quality control testing.

FC100B

Plastic-body (PVDF), double junction, refillable, combination pH electrode.

Use: cheese

FC200B

Plastic-body (PVDF), single junction, conical tip, non-refillable Viscolene electrolyte, combination pH electrode. Use: milk, yogurt, dairy products, and semi-solid foods

FC210B

Glass-body, double junction, conical tip, non-refillable Viscolene electrolyte, combination pH electrode.

Use: milk, yogurt, and cream

FC220B

Glass-body, single junction, refillable, combination pH electrode.

Use: milk, yogurt, cream, sauce, and fruit juices

FC911B

Plastic-body (PVDF), double junction, refillable, combination pH electrode.

Use: sauce, juices, dairy products and other liquid or slurry forms of food

HI1413B

Glass-body, single junction, flat tip, non-refillable Viscolene electrolyte, combination pH electrode. Use: surfaces, skin, leather, paper, and emulsions

A2.2.2. ORP ELECTRODES

HI3131B

Glass-body, refillable, combination platinum ORP electrode. Use: laboratories and general purpose

HI3230B

Plastic-body (PEI), gel-filled, combination platinum ORP electrode. Use: municipal water and quality control

HI4430B

Plastic-body (PEI), gel-filled, combination gold ORP electrode. Use: oxidants and ozone

A2.2.3. HALF-CELL ELECTRODES

HI2110B

Glass-body, single half-cell pH electrode.

Use: general purpose

HI5311

Glass-body, Ag/AgCl reference half-cell electrode, double junction, refillable with 4mm banana plug with 1m (3.3') cable. Use: general purpose with wide temperature range

HI5315

Plastic-body (PEI), double junction, Ag/AgCl reference half-cell electrode, refillable with 4mm plug with 1 m (3.3') cable. Use: Ion Selective Electrodes

HI5412

Glass-body, single Calomel reference half-cell electrode, refillable with 4mm plug with 1m (3.3') cable.

Use: general purpose with constant temperature range

A2.2.4. ION SELECTIVE ELECTRODES

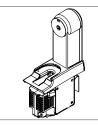
HI4101 Ammonia ISE HI4002 / HI4102 Bromide ISE HI4003 / HI4103 Cadmium ISE HI4004 / HI4104 Chloride ISE HI4005 Carbon Dioxide ISE HI4007 / HI4107 Chloride ISE HI4008 / HI4108 Cupric ISE HI4009 / HI4109 Cyanide ISE HI4010 / HI4110 Fluoride ISE HI4011 / HI4111 Iodide ISE HI4012 / HI4112 Lead ISE HI4013 / HI4113 Nitrate ISE HI4014 / HI4114 Potassium ISE HI4015 / HI4115 Silver / Sulfide ISE FC300B Sodium

A2.2.5. TEMPERATURE SENSOR

HI7662-TW

Temperature probe with 1 m (3.3') paneled cable.

A2.3. TITRATOR COMPONENTS





3 Way Valve HI900260

Burette with:

5 mL Syringe

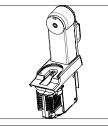
10 mL Syringe

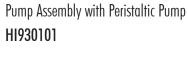
25 mL Syringe

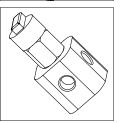
HI900225

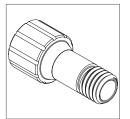
HI900210

HI900205



















50 mL Syringe HI900250

Titrator Peristaltic Pump Complete **Tubing Set** HI930202

Aspiration Tube with fitting and protection tube HI900270

Dispensing Tube with dispensing tip, fitting, protection tube and tube guide HI930280

Overhead Stirrer + 3 propellers HI930301

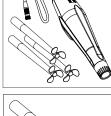
Replacement Propellers (3 pcs.) HI930302

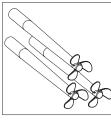
High Chemical Resistance Propellers (3 pcs.) HI930303

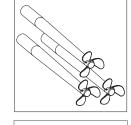
Stirrer Support HI930320

Tool for burette cap removal HI900942



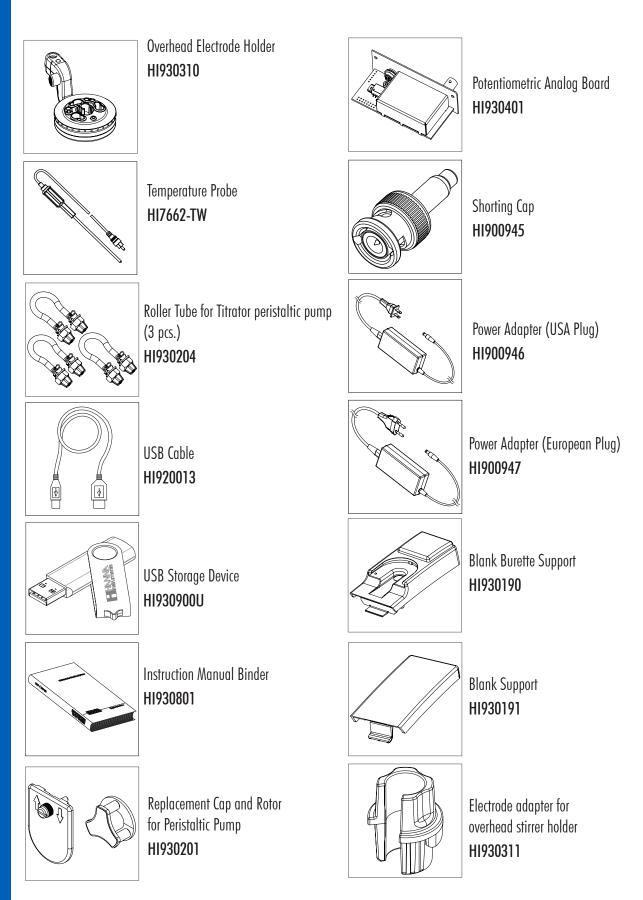








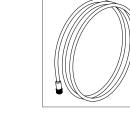
A2-9



A2.4. AUTOSAMPLER COMPONENTS



Autosampler HI922 - XYZ



Communication Cable HI920-933 (HI932 to HI921/HI922)

BNC Extension Cable (1 m)

HI920-931





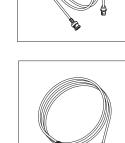




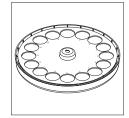
Electrode Holder HI920-310

Tray Locking Screw

HI920-960



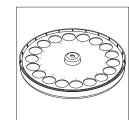
Reference Extension Cable (1 m) HI920-932



16 Beaker Tray, 60 mm dia. Single Row with RFID HI920-11660W



Temperature Sensor HI7662-AW



18 Beaker Tray, 53 mm dia. Single Row with RFID HI920-11853W



USB Memory Stick HI920-901



Plastic Beaker for HI920-11660 (20 pcs.) HI920-060



Titrant Dispensing Tube (1.5 m) HI920-281



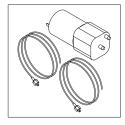
Plastic Beaker for HI920-11853 (20 pcs.) HI920-053



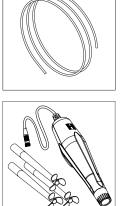
Peristaltic Pump with dispensing tubing HI920-103



Peristaltic pump with aspiration tubing HI920-104



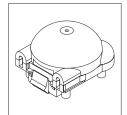
Membrane Pump with tubing HI920-113



Membrane Pump Complete Tubing Set HI920-212

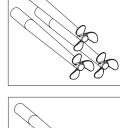
TYGON Tube (5 m) HI920-290





Replacement Cap and Rotor for Peristaltic Pump HI920-201

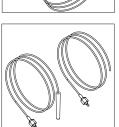
Tubing Set with plastic dispensing tube guide for peristaltic pump HI920-208



Replacement Propellers (3 pcs.) HI930302

High Chemical Resistance Propellers

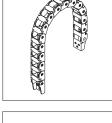




Tubing Set with stainless steel aspiration tube for peristaltic pump HI920-203



Roller Tube for Autosampler peristaltic pump (3 pcs.) HI920-204



Cable Chain HI920-320

(3 pcs.)

HI930303



25 mm x 7 mm Stir Bars (10 pcs.) HI731319

Certification

All Hanna Instruments conform to the CE European Directives.



RoHS compliant

Disposal of Electrical & Electronic Equipment. The product should not be treated as household waste. Instead hand it over to the appropriate collection point for the recycling of electrical and electronic equipment which will conserve natural resources.

Ensuring proper product and battery disposal prevents potential negative consequences for the environment and human health. For more information, contact your city, your local household waste disposal service, the place of purchase or go to www.hannainst.com.



Recommendations for Users

Before using this product, make sure it is entirely suitable for your specific application and for the environment in which it is used. Any variation introduced by the user to the supplied equipment may degrade the meters' performance. For yours and the meter's safety do not use or store the meter in hazardous environments.

Warranty The H1932 is warranted for two years against defects in workmanship and materials when used for its intended purpose and maintained according to instructions. Damage due to accidents, misuse, tampering or lack of prescribed maintenance is not covered.

If service is required, contact your local Hanna Instruments Office. If under warranty, report the model number, date of purchase, serial number and the nature of the problem. If the repair is not covered by the warranty, you will be notified of the charges incurred. If the instrument is to be returned to Hanna Instruments, first obtain a Returned Goods Authorization (RGA) number from the Technical Service department and then send it with shipping costs prepaid. When shipping any instrument, make sure it is properly packed for complete protection.

Hanna Instruments reserves the right to modify the design, construction or appearance of its products without advance notice.

HI932

AUTOMATIC POTENTIOMETRIC TITRATOR





0.1N SODIUM HYDROXIDE TITRANT CONCENTRATION

DESCRIPTION

Method for the standardization (titer determination) of 0.1N Sodium Hvdroxide (NaOH) titrant solution against Potassium Hvdrogen Phthalate (KHP). The results are expressed in N (eq/L).

REFERENCE

AOAC Official Methods of Analysis, Official Method 936.16

ELECTRODE

- HI1131B Combination pH Electrode
- HI7662-T **Temperature Probe**

REAGENTS

- HI70456 0.1N Sodium Hydroxide (1 L)
- HI70401 Potassium Hydrogen Phthalate (20 g)
- HI70436 Deionized Water (1 aal)

ACCESSORIES

- HI70300L Storage Solution (500 mL)
- HI7071 Electrode Fill Solution (30 mL x 4)
- HI7004L pH 4.01 Buffer Solution (500 mL)
- HI7007L pH 7.01 Buffer Solution (500 mL)
- HI7010L pH 10.01 Buffer Solution (500 mL)
- HI740036P 100 mL Plastic Beaker (10 pcs)
- Analytical Balance with 0.0001 g resolution

DEVICE PREPARATION

- Connect the pH electrode and temperature probe to the titrator.
- Install a 25 mL burette filled with 0.1N sodium hydroxide (HI70456) on pump one and verify that no air bubbles are present in the burette or tubing. If necessary prime the burette until all the air has been removed completely.
- Press Select from the main screen. Use the arrow keys to highlight HIOOO1EN 0.1N Sodium Hydroxide and press Select

ELECTRODE PREPARATION

- Press Mode from the main screen, if necessary select the analog board and press
- Calibrate the electrode using pH 4.01, 7.01 and 10.01 buffers. Refer to the instruction manual for calibration procedure.

SAMPLE PREPARATION

- Crush approximately 3 grams of potassium hydrogen phthalate (HI70401) and dry it for 2 hours at 120°C. Cool to room temperature in a desiccator.
- Place a clean 100 mL plastic beaker on the analytical balance.
- Zero the balance.
- Carefully weigh approximately 0.20 grams of dried potassium hydroaen phthalate into the beaker. Ensure that all of the potassium hydrogen phthalate is on the bottom of the beaker.

- Record the exact weight of the sample once the balance has • stabilized with an accuracy of 0.0001 grams.
- Remove the beaker from the balance and add deionized water to the 50 mL mark on the beaker.

ANALYSIS

Place the beaker under the stirrer assembly and lower it to immerse the pH electrode, temperature probe and stirrer. Ensure that the reference junction of the pH electrode is 5 to 6 mm below the surface. If necessary add extra deionized water. *Note:* The dispensing tip should be slightly submerged in the sample.

Press start stop . You will be prompted to enter the weight of the analyte (weight of potassium hydrogen phthalate). Use the numeric keypad to enter the exact weight and press start the analysis.

Note: Ensure that the potassium hydrogen phthalate dissolves completely during the pre-titration stir time. Erroneous results may occur if the sample does not dissolve completely prior to titration. If necessary the pre-titration stir time can be increased.

- At the end of the titration, after detection of the equivalence point, "Titration Completed" will appear with the result. The result is expressed in N (eq/L) of sodium hydroxide.
- Remove the pH electrode, temperature probe and stirrer from the sample and rinse them thoroughly with deionized water.
- Record the result.

Note: For improved accuracy, repeat this procedure a minimum of three times and calculate the average value.

For methods utilizing 0.1N sodium hydroxide titrant solution, follow the steps below to enter the titer/standardized value.

- Select the method utilizing 0.1N sodium hydroxide.
- Press Method Options from the main screen. •
- Using the arrow keys, highlight Titrant Conc. and press Select
- Use the numeric keypad to enter the standardized (titer) value of the titrant then press Accept. • Press Escape to exit the View/Modify Method screen. Use
- the arrow keys to highlight *Save Method* and press Select

0.1N SODIUM HYDROXIDE TITRANT CONCENTRATION

METHOD PARAMETERS

Name:	0.1N Sodium Hydroxide
Method Revision:	3.0
Analysis Type:	Standard Titration
Analog Board:	Analog 1
Stirrer Configurati	lon:
Stirrer:	Stirrer 1
Stirring Speed:	: 1400 RPM
Pump Configuration:	:
Titrant Pump:	Pump 1
Reagent Addition 1	
Reagent Addition 2	2: Disabled
Dosing Type:	Dynamic
Min Vol:	0.030 mL
Max Vol:	0.500 mL
delta E:	4.500 mV
	pH 1EQ point, 1st Der
Recognition Optior	ns:
Threshold:	500 mV/mL
Range:	NO
Filtered Deriva	atives: NO
Pre-Titration Volu	ame: 5.000 mL
Pre-Titration Stin	Time: 60 sec
Measurement Mode:	
delta E:	0.3 mV
delta t:	2 sec
Min wait:	3 sec
Max wait:	30 sec
Electrode Type:	рH
Blank Option:	No Blank
	tdz. Titrant by Weight
Dilution Option:	Disabled
Titrant Name:	0.1N NaOH
Analyte Size:	0.20000 g
Analyte Entry:	Manual
Maximum Titrant Vo	
	-2000.0 to 2000.0 mV
Volume/Flow Rate:	
Signal Averaging:	1 Reading
Significant Figures	S: XXXXX

CALCULATIONS

CALCOLATIONS		
Calculations: Stdz.	Titrant by Weight	
Titrant units:	N (eq/L)	
Titrant volume dosed:	V (L)	
Standard weight:	0.200 g	
mw of standard:	204.23 g/mol	
Titrant/Standard:	1.000 eq/mol	
	-	
$\frac{\text{eq}}{\text{L}} \text{NaOH} = \frac{0.200 * 1.000}{204.23 * \text{V(L)}}$		
L 204.	23 * V(L)	
RESULTS		
Titration R	eport	
Method Name: 0.1N	Sodium Hydroxide	
Time & Date: 1	7:03 Jun 07, 2018	
Report ID:	Ti 00053	
-	—	
Titration Re	esults	
Method Name: 0.1N	Sodium Hydroxide	
Time & Date: 1	7:03 Jun 07, 2018	
Analyte Size:	0.20920 g	
End Point Volume:	10.215 mL	
pH Equivalence Point:	8.394	
Result:	0.10027 N(eq/L)	
Initial & Final pH:	4.173 to 9.570	
Titration Duration:	6:25 [mm:ss]	
IICIACION DUIACION.	0.20 [1000.55]	

Analyst Signature:_____

Titration went to Completion

0.1N HYDROCHLORIC ACID TITRANT CONCENTRATION

DESCRIPTION

Method for the standardization (titer determination) of 0.1N Hydrochloric Acid (HCl) titrant solution against Sodium Hydroxide (NaOH). The results are expressed in N (eq/L).

REFERENCE

AOAC Official Methods of Analysis, Official Method 936.15

ELECTRODE

- HI1131B Combination pH Electrode
- HI7662-T Temperature Probe

REAGENTS

- HI70463 0.1N Hydrochloric Acid (1 L)
- HI70456 0.1N Sodium Hydroxide (1 L)
- HI70436 Deionized Water (1 gal)

ACCESSORIES

- HI70300L Storage Solution (500 mL)
- HI7071 Electrode Fill Solution (30 mL x 4)
- HI7004L pH 4.01 Buffer Solution (500 mL)
- HI7007L pH 7.01 Buffer Solution (500 mL)
- HI7010L pH 10.01 Buffer Solution (500 mL)
- HI740036P 100 mL Plastic Beakers (10 pcs)
- 10 mL Class A Volumetric Pipette

DEVICE PREPARATION

- Connect the pH electrode and temperature probe to the titrator.
- Install a 25 mL burette filled with 0.1N hydrochloric acid (HI70463) on pump one and verify that no air bubbles are present in the burette or tubing. If necessary prime the burette until all the air has been removed completely.
- Press Select Method from the main screen. Use the arrow keys to highlight HI0002EN 0.1N Hydrochloric Acid and press Select

ELECTRODE PREPARATION

- Press Mode from the main screen and press PH
- Calibrate the electrode using pH 4.01, 7.01 and 10.01 buffers. Refer to the instruction manual for calibration procedure.

SAMPLE PREPARATION

- Use a Class A volumetric pipette to transfer exactly 10.00 mL of 0.1N sodium hydroxide (HI70456) to a clean 100 mL beaker.
- Add deionized water to the 50 mL mark on the beaker.

ANALYSIS

 Place the beaker under the stirrer assembly and lower it to immerse the pH electrode, temperature probe and stirrer. Ensure that the reference junction of the pH electrode is 5 to 6 mm below the surface. If necessary add extra deionized water.

Note: The dispensing tip should be slightly submerged in the sample.

- Press stop , the titrator start the analysis.
- At the end of the titration, after detection of the equivalence point, "Titration Completed" will appear with the result. The result is expressed in N (eq/L) of hydrochloric acid.
- Remove the pH electrode, temperature probe and stirrer from the sample and rinse them thoroughly with deionized water.
 - Record the result. *Note:* For improved accuracy, repeat this procedure a minimum of three times and calculate the average value.

For methods utilizing 0.1N hydrochloric acid titrant solution, follow the steps below to enter the titer/standardized value.

- Select the method utilizing 0.1N hydrochloric acid.
- Press Method Options from the main screen.
- Using the arrow keys, highlight *Titrant Conc.* and press
 Select
- Use the numeric keypad to enter the standardized (titer) value of the titrant then press Accept .
- Press Escape to exit the View/Modify Method screen. Use the arrow keys to highlight *Save Method* and press Select

0.1N HYDROCHLORIC ACID TITRANT CONCENTRATION

METHOD PARAMETERS

METTODIARAMETERS	
Name:	0.1N Hydrochloric Acid
Method Revision:	3.0
Analysis Type:	Standard Titration
Analog Board:	Analog 1
Stirrer Configurat	ion:
Stirrer:	Stirrer 1
Stirring Speed	
Pump Configuration	:
Titrant Pump:	Pump 1
Reagent Addition	
Reagent Addition	2: Disabled
Dosing Type:	Dynamic
Min Vol:	0.030 mL
Max Vol:	0.500 mL
delta E:	6.000 mV
End Point Mode:	pH 1EQ point, 1st Der
Recognition Optio	ns:
Threshold:	500 mV/mL
Range:	NO
Filtered Deriv	atives: NO
Pre-Titration Vol	ume: 5.000 mL
Pre-Titration Sti	r Time: 0 sec
Measurement Mode:	Signal Stability
delta E:	1.0 mV
delta t:	2 sec
Min wait:	3 sec
Max wait:	15 sec
Electrode Type:	рH
Blank Option:	No Blank
Calculations:	Stdz. Titrant by Volume
Dilution Option:	Disabled
Titrant Name:	0.1N HCl
Analyte Size:	10.0000 mL
Analyte Entry:	Fixed
Maximum Titrant V	olume: 15.000 mL
Potential Range:	-2000.0 to 2000.0 mV
	25 mL/50.0 mL/min
Signal Averaging:	1 Reading
Significant Figure	2
2	

CALCULATIONS

Calculations: Stdz. Titran	t by Volume	
Titrant units:	N (eq/L)	
Titrant volume dosed:	V (L)	
Standard volume:	10.000 mL	
Standard conc.:	0.100 eq/L	
$\frac{\text{eq}}{2}$ HCl = $\frac{10.000 * 0.10}{2}$	0	
L ICI – V(L) * 1000	_	
RESULTS		
Titration Report		
Method Name: 0.1N Hydroch	nloric Acid	
Time & Date: 14:55 Ju.	ly 30, 2018	
Report ID:	Ti_00002	

Titration Results

Method Name: 0	.1N Hydrochloric Acid
Time & Date:	14:55 July 30, 2018
Analyte Size:	10.000 mL
End Point Volume:	9.979 mL
pH Equivalence Poi	nt: 5.059
Result:	0.10020 N(eq/L)
Initial & Final pH	: 12.135 to 4.989
Titration Duration	: 2:45 [mm:ss]
Titration went to	Completion

Analyst Signature:_____

METHOD ID: HI0003EN

0.1M SODIUM THIOSULFATE TITRANT CONCENTRATION

DESCRIPTION

Method for the standardization (titer determination) of 0.1M Sodium Thiosulfate $(Na_2S_2O_3)$ titrant solution against Potassium lodate (KIO₂). The results are expressed in M (mol/L).

REFERENCE

Standard Methods for the Examination of Water and Wastewater 19th Edition, Method 4500-Cl B

ELECTRODE

• HI3131B Combination ORP Electrode

REAGENTS

- HI70439 0.1M Sodium Thiosulfate (1 L)
- HI70407 Potassium lodate (20 g)
- HI70425 16% Sulfuric Acid (500 mL)
- HI70468 Potassium lodide (35 g)
- HI70436 Deionized Water (1 gal)

ACCESSORIES

- HI70300L Storage Solution (500 mL)
- HI7071 Electrode Fill Solution (30 mL x 4)
- HI740036P 100 mL Plastic Beakers (10 pcs)
- Analytical Balance 0.0001 g
- 100 mL Class A Volumetric Flask
- 10 mL Class A Volumetric Pipette

DEVICE PREPARATION

- Connect the ORP electrode to the titrator.
- Install a 25 mL burette filled with 0.1M sodium thiosulfate (HI70439) on pump one and verify that no air bubbles are present in the burette or tubing. If necessary prime the burette until all the air has been removed completely.
- Press Select Method
 from the main screen. Use the arrow keys to highlight HI0003EN 0.1M Sodium Thiosulfate and press Select

ELECTRODE PREPARATION

• Prepare the ORP electrode according to the procedure in the manual.

SAMPLE PREPARATION

- Crush approximately 2 grams of potassium iodate (HI70407) and dry it for 2 hours at 120°C. Cool to room temperature in a desiccator.
- Carefully weigh approximately 0.35 grams of dried potassium iodate.
- Record the exact weight of the sample once the balance has stabilized with an accuracy of 0.0001 grams.
- Carefully transfer the salt to a 100 mL Class A volumetric flask. Add approximately 80 mL of deionized water, and mix to dissolve. Once the salt is completely dissolved bring the flask to volume with deionized water, mix well.

- Use a Class A volumetric pipette to transfer exactly 10.00 mL of the solution to a clean 100 mL plastic beaker.
- Add deionized water to the 50 mL mark on the beaker.
- Add 5.00 mL of 16% sulfuric acid (HI70425) and 1.5 grams of potassium iodide (HI70468) to the beaker.

ANALYSIS

 Place the beaker under the stirrer assembly and lower it to immerse the ORP electrode and stirrer. Ensure that the reference junction of the ORP electrode is 5 to 6 mm below the surface. If necessary add extra deionized water.

Note: The dispensing tip should be slightly submerged in the sample.

- Press stop . You will be prompted to enter the weight of the analyte (weight of potassium iodate). Use the numeric keypad to enter the exact weight and press enter to start the analysis.
- At the end of the titration, after detection of the equivalence point, "Titration Completed" will appear with the result. The result is expressed in M (mol/L) of sodium thiosulfate.
- Remove the ORP electrode and stirrer from the sample and rinse them thoroughly with deionized water.
- Record the result.

Note: For improved accuracy, repeat this procedure a minimum of three times and calculate the average value.

For methods utilizing 0.1M sodium thiosulfate titrant solution, follow the steps below to enter the titer/standardized value.

- Select the method utilizing 0.1M sodium thiosulfate.
- Press Method Options from the main screen.
- Using the arrow keys, highlight *Titrant Conc.* and press
 Select
- Use the numeric keypad to enter the standardized (titer) value of the titrant then press Accept
- Press Escape to exit the View/Modify Method screen. Use the arrow keys to highlight *Save Method* and press Select

0.1M SODIUM THIOSULFATE TITRANT CONCENTRATION

METHOD PARAMETERS

METHOD FARAMETERS	
Name:	0.1M Sodium Thiosulfate
Method Revision:	3.0
Analysis Type:	Standard Titration
Analog Board:	Analog 1
Stirrer Configurat	cion:
Stirrer:	Stirrer 1
Stirring Speed	
Pump Configuration	1:
Titrant Pump:	Pump 1
Reagent Addition	
Reagent Addition	
Dosing Type:	Dynamic
Min Vol:	0.030 mL
Max Vol:	0.600 mL
delta E:	6.500 mV
	mV 1EQ point, 1st Der
Recognition Optic	
Threshold:	50 mV/mL
Range:	NO
Filtered Deriv	
Pre-Titration Vol	
Pre-Titration Sti	
Measurement Mode:	2 1
delta E:	0.3 mV
delta t:	2 sec
Min wait:	2 sec
Max wait:	20 sec
Electrode Type:	ORP
Blank Option:	No Blank
	Stdz. Titrant by Weight
Dilution Option:	Enabled
Final Dilution	
Aliquot Volume	
Titrant Name:	0.1M Na2S2O3
Analyte Size:	0.35000 g
Analyte Entry: Maximum Titrant V	Manual Volume: 15.000 mL
Volume/Flow Rate:	-2000.0 to 2000.0 mV 25 mL/50.0 mL/min
Signal Averaging:	
Significant Figure	2
Significant rigule	

CALCULATIONS

Calculations: Stdz. Tit	rant by Weight
Titrant units:	M (mol/L)
Titrant volume dosed:	V (L)
Standard weight:	0.350 g
Dilution Factor:	0.100
Final Dilution volume:	100.000 mL
Aliquot Volume:	10.000 mL
mw of standard:	214.00 g/mol
Titrant/Standard:	6.000 mol/mol
mol Na S 0 - 0.350 * 0	.10 * 6.0
$\frac{\text{mol}}{\text{L}} \text{Na}_2 \text{S}_2 \text{O}_3 = \frac{0.350 * 0}{214.00}$	

RESULTS

	Titration	n Report
Method	Name: 0.1M	1 Sodium Thiosulfate
Time &	Date:	17:10 Jun 22, 2018
Report	ID:	Ti_00073

Titration Results		
Method Name: 0.1M	Sodium Thiosulfate	
Time & Date:	17:10 Jun 22, 2018	
Analyte Size:	0.35020 g	
End Point Volume:	9.635 mL	
mV Equivalence Point:	233.0	
Result:	0.10191 M (mol/L)	
Initial & Final mV:	361.8 to 173.4	
Titration Duration:	2:51 [mm:ss]	
Titration went to Com	pletion	

Analyst Signature:

METHOD ID: HI0010EN

0.1M FERROUS AMMONIUM SULFATE TITRANT CONCENTRATION

DESCRIPTION

Method for the standardization (titer determination) of 0.1MFerrous Ammonium Sulfate (FAS) titrant solution against Potassium Dichromate (K₂Cr₂O₂). The results are expressed in M (mol/L).

REFERENCE

Standard Methods for the Examination of Water and Wastewater 21st Edition, Method 5220B

ELECTRODE

• HI3131B Combination ORP Electrode

REAGENTS

- HI70444 25% Sulfuric Acid
- HI70436 Deionized Water (1 gal)
- Ferrous Ammonium Sulfate (ACS Grade)
- Potassium Dichromate (ACS Grade)

ACCESSORIES

- HI70300L Storage Solution (500 mL)
- HI7071 Electrode Fill Solution (30 mL x 4)
- HI740036P 100 mL Plastic Beakers (10 pcs)
- Analytical Balance with 0.0001 g resolution
- 100 mL Class A Volumetric Flask
- 500 mL Class A Volumetric Flask
- 10 mL Class A Volumetric Pipette

TITRANT PREPARATION

- Carefully weigh 19.607 grams of ferrous ammonium sulfate.
- Carefully transfer the salt to a 500 mL Class A volumetric flask. Add approximately 300 mL of deionized water, and mix to dissolve.
- Add 40.00 mL of 25% sulfuric acid (HI70444) to the flask. Invert the solution to mix.
- Allow the flask to return to room temperature.
- Bring the flask to volume with deionized water, mix well.

DEVICE PREPARATION

- Connect the ORP electrode to the titrator.
- Install a 25 mL burette filled with 0.1M ferrous ammonium sulfate on pump one and verify that no air bubbles are present in the burette or tubing. If necessary prime the burette until all the air has been removed completely.
- Press Select Method from the main screen. Use the arrow keys to highlight HI0010EN 0.1M FAS and press Select.

ELECTRODE PREPARATION

• Prepare the ORP electrode according to the procedure in the manual.

SAMPLE PREPARATION

 Crush approximately 2 grams of potassium dichromate and dry it for 2 hours at 150°C. Cool to room temperature in a desiccator.

- Carefully weigh approximately 0.49 grams of dried potassium dichromate.
- Record the exact weight of the sample once the balance has stabilized with an accuracy of 0.0001 grams.
- Carefully transfer the salt to a 100 mL Class A volumetric flask. Add approximately 80 mL of deionized water, and mix to dissolve. Once the salt is completely dissolved bring the flask to volume with deionized water, mix well.
- Use a Class A volumetric pipette to transfer exactly 10.00 mL of the solution to a clean 100 mL plastic beaker.
- Add 25.00 mL of 25% sulfuric acid (HI70444) to the beaker.
- Add deionized water to the 50 mL mark on the beaker.

ANALYSIS

 Place the beaker under the stirrer assembly and lower it to immerse the electrode and stirrer. Ensure that the reference junction of the ORP electrode is 5 to 6 mm below the surface. If necessary add extra deionized water.

Note: The dispensing tip should be slightly submerged in the sample.

- Press start stop
 You will be prompted to enter the weight of the analyte (weight of potassium dichromate). Use the numeric keypad to enter the exact weight and press enter to start the analysis.
- At the end of the titration, after detection of the equivalence point, "Titration Completed" will appear with the result. The result is expressed in M (mol/L) of ferrous ammonium sulfate.
- Remove the ORP electrode and stirrer from the sample and rinse them thoroughly with deionized water.
- Record the result.

Note: For improved accuracy, repeat this procedure a minimum of three times and calculate the average value.

For methods utilizing 0.1M ferrous ammonium sulfate titrant solution, follow the steps below to enter the titer/standardized value.

- Select the method utilizing 0.1M ferrous ammonium sulfate
- Press Method from the main screen.
- Using the arrow keys, highlight *Titrant Conc.* and press
 Select
- Use the numeric keypad to enter the standardized (titer) value of the titrant then press Accept .
- Press Escape to exit the View/Modify Method screen and select Save Method and press Select

0.1M FERROUS AMMONIUM SULFATE TITRANT CONCENTRATION

METHOD PARAMETERS

Name:	0.1M FAS
Method Revision:	3.0
Analysis Type:	Standard Titration
Analog Board:	Analog 1
Stirrer Configuration:	:
Stirrer:	Stirrer 1
Stirring Speed:	1400 RPM
Pump Configuration:	
Titrant Pump:	Pump 1
Reagent Addition 1:	Disabled
Reagent Addition 2:	Disabled
Dosing Type:	Dynamic
Min Vol:	0.030 mL
Max Vol:	0.500 mL
delta E:	4.500 mV
End Point Mode: mV	1EQ point, 1st Der
Recognition Options:	
Threshold:	35 mV/mL
Range:	NO
Filtered Derivativ	ves: NO
Pre-Titration Volume:	: 5.000 mL
Pre-Titration Stir Ti	ime: 0 sec
Measurement Mode:	Signal Stability
delta E:	0.5 mV
delta t:	3 sec
Min wait:	2 sec
Max wait:	20 sec
Electrode Type:	ORP
Blank Option:	No Blank
Calculations: Stdz	. Titrant by Weight
Dilution Option:	Enabled
Final Dilution Vol	ume: 100.000 mL
Aliquot Volume:	10.000 mL
Titrant Name:	0.1M FAS
Analyte Size:	0.49000 g
Analyte Entry:	Manual
Maximum Titrant Volum	ne: 15.000 mL
Potential Range: -	2000.0 to 2000.0 mV
Volume/Flow Rate:	25 mL/50.0 mL/min
Signal Averaging:	1 Reading
Significant Figures:	XXXXX

CALCULATIONS

Calculations: Stdz. Tit	rant by Weight
Titrant units:	M (mol/L)
Titrant volume dosed:	V (L)
Standard weight:	0.490 g
Dilution Factor:	0.100
Final Dilution volume:	100.000 mL
Aliquot Volume:	10.000 mL
mw of standard:	294.18 g/mol
Titrant/Standard:	6.000 mol/mol
$\frac{\text{mol}}{\text{FAS}} = \frac{0.490 \times 0.12}{1000}$	
L 294.18 *	* V(L)

RESULTS

	Titrati	on Report
Method	Name:	0.1M FAS
Time &	Date:	15:59 August 1, 2018
Report	ID:	Ti_00015

Titration ResultsMethod Name:0.1M FASTime & Date:15:59 August 1, 2018Analyte Size:0.491 gEnd Point Volume:9.879 mLmV Equivalence Point:667.4Result:0.10137 M (mol/L)Initial & Final mV:791.3 to 598.0Titration Duration:3:05 [mm:ss]

Analyst Signature:____

Titration went to Completion

0.02M SILVER NITRATE TITRANT CONCENTRATION

DESCRIPTION

Method for the standardization (titer determination) of 0.02M Silver Nitrate (AgNO₃) titrant solution against Sodium Chloride (NaCl). The results are expressed in M (mol/L).

REFERENCE

AOAC Official Methods of Analysis, Official Method 941.18

ELECTRODE

HI4115 Silver/Sulfide Combination ISE

REAGENTS

- HI70448 0.02M Silver Nitrate (1 L)
- HI70406 Sodium Chloride (20 g)
- HI70427 1.5M Nitric Acid Solution (500 mL)
- HI70436 Deionized Water (1 gal)

ACCESSORIES

- HI7072 Electrode Fill Solution (4 x 30 mL)
- Analytical Balance with 0.0001 g resolution
- 150 mL Glass Beaker
- 100 mL Class A Volumetric Flask
- 5 mL Class A Volumetric Pipette

DEVICE PREPARATION

- Connect the Silver/Sulfide electrode to the titrator.
- Install a 25 mL burette filled with 0.02M silver nitrate (HI70448) on pump one and verify that no air bubbles are present in the burette or tubing. If necessary prime the burette until all the air has been removed completely.
- Press Select Method from the main screen. Use the arrow keys to highlight HI0200EN 0.02M Silver Nitrate and press Select

ELECTRODE PREPARATION

• Prepare the Silver/Sulfide electrode according to the procedure in the manual.

SAMPLE PREPARATION

- Crush approximately 2 grams of sodium chloride (HI70406) and dry it for 2 hours at 140°C. Cool to room temperature in a desiccator.
- Weigh 0.20 g of dried sodium chloride with an accuracy of 0.0001 g. Transfer the salt to a 100 mL volumetric flask. Add approximately 80 mL of distilled water and mix. Dissolve completely before bringing to volume.
- Use a Class A volumetric pipette to transfer exactly 5.00 mL of prepared standard solution to a 150 mL glass beaker and add distilled water to the 100 mL mark on the beaker.
- Add 10.00 mL of 1.5M nitric acid (HI70427) to the beaker.

ANALYSIS

 Place the beaker under the stirrer assembly and lower it to immerse the Silver/Sulfide electrode and stirrer. Ensure that the reference junction of the electrode is 5 to 6 mm below the surface. If necessary add extra deionized water.

Note: The dispensing tip should be slightly submerged in the sample.

- Press start stop
 You will be prompted to enter the weight of the analyte (weight of sodium chloride). Use the numeric keypad to enter the exact weight and press enter to start the analysis.
- At the end of the titration, after detection of the equivalence point, "Titration Completed" will appear with the result. The result is expressed in M (mol/L) of silver nitrate.
- Remove the electrode and stirrer from the sample and rinse them thoroughly with deionized water.
- Record the result.

Note: For improved accuracy, repeat this procedure a minimum of three times and calculate the average value.

For methods utilizing 0.02M silver nitrate titrant solution, follow the steps below to enter the titer/standardized value.

- Select the method utilizing 0.02M silver nitrate.
- Press Method Options from the main screen.
- Using the arrow keys, highlight *Titrant Conc.* and press Select
- Use the numeric keypad to enter the standardized (titer) value of the titrant then press Accept.
- Press Escape to exit the View/Modify Method screen. Use the arrow keys to highlight Save Method and press Select

0.02M SILVER NITRATE TITRANT CONCENTRATION

METHOD PARAMETERS

METHOD PAKAMETERS	
Name:	0.02M Silver Nitrate
Method Revision:	3.0
Analysis Type:	Standard Titration
Analog Board:	Analog 1
Stirrer Configuratio	on:
Stirrer:	Stirrer 1
Stirring Speed:	1400 RPM
Pump Configuration:	
Titrant Pump:	Pump 1
Reagent Addition 1:	
Reagent Addition 2:	
Dosing Type:	Dynamic
Min Vol:	0.030 mL
Max Vol:	0.500 mL
delta E:	8.000 mV
	mV 1EQ point, 1st Der
Recognition Options	
Threshold:	100 mV/mL
Range:	NO
Filtered Derivat	
Pre-Titration Volum Pre-Titration Stir	
Measurement Mode:	Time: 0 sec Signal Stability
delta E:	1.0 mV
delta t:	2 sec
Min wait:	2 sec 2 sec
Max wait:	20 sec
Electrode Type:	Silver/Sulfide
Blank Option:	No Blank
	dz. Titrant by Weight
Dilution Option:	Enabled
Final Dilution N	
Aliquot Volume:	5.000 mL
Titrant Name:	0.02M AqNO3
Analyte Size:	0.20000 g
Analyte Entry:	Manual
Maximum Titrant Vol	Lume: 15.000 mL
Potential Range:	-2000.0 to 2000.0 mV
Volume/Flow Rate:	25 mL/50.0 mL/min
Signal Averaging:	1 Reading
Significant Figures:	: XXXXX

CALCULATIONS

Calculations: Stdz. Tit	rant by Weight
Titrant units:	M (mol/L)
Titrant volume dosed:	V (L)
Standard weight:	0.200 g
Dilution Factor:	0.05
Final Dilution volume:	100.000 mL
Aliquot Volume:	5.000 mL
mw of standard:	58.440 g/mol
Titrant/Standard:	1.000 mol/mol
$\frac{\text{mol}}{\text{L}} \text{AgNO}_{3} = \frac{0.200 * 0}{58.440}$	
Ц 58.440	^ ∨(⊥)

RESULTS

	Titrati	on Report
Method	Name:	0.02M Silver Nitrate
Time &	Date:	15:52 August 1, 2018
Report	ID:	Ti 00037

Titration ResultsMethod Name:0.02M Silver NitrateTime & Date:15:52 August 1, 2018Analyte Size:0.1923 gEnd Point Volume:9.065 mLmV Equivalence Point:273.1Result:0.01815 M (mol/L)Initial & Final mV:146.9 to 291.0Titration Duration:2:21 [mm:ss]Titration went to Completion

Analyst Signature:____

ALKALINITY OF WATER 0 to 2500 mg/L CaCO₃, pH 4.5 Endpoint

DESCRIPTION

Method for the determination of total (methyl red) alkalinity in water by titration of a sample to pH 4.5. The results are expressed in mg/L (ppm) as calcium carbonate.

For the determination of phenolphthalein alkalinity, set the endpoint to pH 8.3.

REFERENCE

Standard Methods for the Examination of Water and Wastewater $21^{\,\rm st}$ edition, Method 2320B

ELECTRODE

- HI1131B Combination pH Electrode
- HI7662-T Temperature Probe

REAGENTS

- HI70463 0.1N Hydrochloric Acid (1 L)
- HI70436 Deionized Water (1 gal)

ACCESSORIES

- HI70300L Storage Solution (500 mL)
- HI7082 Electrode Fill Solution (4 x 30 mL)
- HI7004L pH 4.01 Buffer Solution (500 mL)
- HI7007L pH 7.01 Buffer Solution (500 mL)
- HI7010L pH 10.01 Buffer Solution (500 mL)
- HI740036P 100 mL Plastic Beaker (10 pcs)
- 50 mL Class A Volumetric Pipette

DEVICE PREPARATION

- Connect the pH electrode and temperature probe to the titrator.
- Install a 25 mL burette filled with 0.1N hydrochloric acid (HI70463) on pump one and verify that no air bubbles are present in the burette or tubing. If necessary prime the burette until all the air has been removed completely.
- For the determination of the exact concentration of the 0.1N hydrochloric acid, follow *HIOOO2EN 0.1N Hydrochloric Acid* Titrant Concentration.
- Press Select Method from the main screen. Use the arrow keys to highlight Alkalinity of Water and press Select .

ELECTRODE PREPARATION

- Press Mode from the main screen, if necessary select the analog board and press PH.
- Calibrate the electrode using pH 4.01, 7.01 and 10.01 buffers. Refer to the instruction manual for calibration procedure.

SAMPLE PREPARATION

 Use a Class A volumetric pipette to transfer exactly 50.00 mL of sample to a clean 100 mL plastic beaker.

ANALYSIS

- Place the beaker under the stirrer assembly and lower it to immerse the pH electrode, temperature sensor and stirrer. Ensure that the reference junction of the pH electrode is 5 to 6 mm below the surface. If necessary add extra deionized water. *Note:* The dispensing tip should be slightly submerged in the sample.
- Press start stop , the titrator will start the analysis.
- At the end of the titration, when pH 4.50 is reached, "Titration Completed" will appear with the result. The result is expressed in mg/L as calcium carbonate.
- Remove the pH electrode, temperature probe and stirrer from the sample and rinse them thoroughly with deionized water.
- Record the result.

ALKALINITY OF WATER 0 to 2500 mg/L CaCO₃, pH 4.5 Endpoint

METHOD PARAMETERS

METTODTAKAMETEKS	
Name:	Alkalinity of Water
Method Revision:	3.0
Analysis Type:	Standard Titration
Analog Board:	Analog 1
Stirrer Configuration	n:
Stirrer:	Stirrer 1
Stirring Speed:	1400 RPM
Pump Configuration:	
Titrant Pump:	Pump 1
Reagent Addition 1:	Disabled
Reagent Addition 2:	Disabled
Dosing Type:	Dynamic
Min Vol:	0.050 mL
Max Vol:	0.500 mL
delta E:	5.000 mV
End Point Mode:	Fixed 4.500 pH
Pre-Titration Volume	e: 0.000 mL
Pre-Titration Stir 7	Time: 0 sec
Measurement Mode:	Signal Stability
delta E:	1.0 mV
delta t:	2 sec
Min wait:	2 sec
Max wait:	20 sec
Electrode Type:	pH
Blank Option:	No Blank
Calculations: Sa	mple Calc. by Volume
Dilution Option:	Disabled
Titrant Name:	0.1N HCl
Titrant Conc.:	0.1000 N(eq/L)
Analyte Size:	50.000 mL
Analyte Entry:	Fixed
Maximum Titrant Volu	
Potential Range:	-2000.0 to 2000.0 mV
Volume/Flow Rate:	25 mL/50.0 mL/min
Signal Averaging:	1 Reading
Significant Figures:	XXXXX
2	

CALCULATIONS

Calculations:	Sample	Calc.	by	Volume
Titrant units:			Ν	(eq/L)
Titrant volume do	sed:			V (L)
Final result unit	s:			mg/L
Titrant Conc.:		0.10	1 00	l(eq/L)
Sample/Titrant:		0.5	500	mol/eq
mw of sample:		100	0.0	9 g/mol
Sample Volume:				.000 mL
$\frac{\text{mg}}{\text{T}} \text{CaCO}_3 = \frac{\text{V(L)} * 1000}{\text{V(L)}}$) * 0.10 *	0.5 * 1	00.0)9 * 1000
L = -	50	.00		

RESULTS

	Titrat	ion Report
Method	Name:	Alkalinity of Water
Time &	Date:	14:36 August 1, 2018
Report	ID:	Ti_00036

Titratio	n Results
Method Name:	Alkalinity of Water
Time & Date:	14:36 August 1, 2018
Analyte Size:	50.000 mL
End Point Volume:	9.336 mL
pH Fixed End Point:	4.500
Result:	934.44 mg/L
Initial & Final pH:	10.232 to 4.419
Titration Duration:	3:23 [mm:ss]
Titration went to C	ompletion

Analyst Signature:_____

ACIDITY OF WATER

ACIDITY OF WATER 0 to 2500 mg/L, pH 8.3 Endpoint

DESCRIPTION

Method for the determination of total (phenolphthalein) acidity in water by titration of a sample to pH 8.3. The results are expressed in mg/L (ppm) as calcium carbonate.

For the determination of methyl orange acidity, set the endpoint to pH 3.7.

REFERENCE

Standard Methods for the Examination of Water and Wastewater 21st edition, Method 2310B

ELECTRODE

- Combination pH Electrode HI1131B
- HI7662-T **Temperature Probe**

REAGENTS

- HI70456 0.1N Sodium Hydroxide (1 L)
- HI70436 Deionized Water (1 gal)

ACCESSORIES

- HI70300L Storage Solution (500 mL)
- HI7082 Electrode Fill Solution (4 x 30 mL)
- HI7004L pH 4.01 Buffer Solution (500 mL)
- HI7007L pH 7.01 Buffer Solution (500 mL)
- HI7010L pH 10.01 Buffer Solution (500 mL) •
- HI740036P 100 mL Plastic Beaker (10 pcs)
- 50 mL Class A Volumetric Pipette

DEVICE PREPARATION

- Connect the pH electrode and temperature probe to the titrator.
- Install a 25 mL burette filled with 0.1N sodium hydroxide (HI70456) on pump one and verify that no air bubbles are present in the burette or tubing. If necessary prime the burette until all the air has been removed completely.
- For the determination of the exact concentration of the 0.1N sodium hydroxide, follow HIOOO1EN 0.1N Sodium Hydroxide Titrant Concentration.
- Press Select Method from the main screen. Use the arrow keys to highlight Acidity of Water and press

ELECTRODE PREPARATION

- Press Mode from the main screen, if necessary select the analog board and press [PH]. Calibrate the electrode using pH 4.01, 7.01 and 10.01 buffers.
- Refer to the instruction manual for calibration procedure.

SAMPLE PREPARATION

Use a Class A volumetric pipette to transfer exactly 50.00 mL of • sample to a clean 100 mL plastic beaker.

ANALYSIS

- Place the beaker under the stirrer assembly and lower it to immerse the pH electrode, temperature sensor and stirrer. Ensure that the reference junction of the pH electrode is 5 to 6 mm below the surface. If necessary add extra deionized water. *Note:* The dispensing tip should be slightly submerged in the sample.
- start stop , the titrator will start the analysis. Press
- At the end of the titration, when pH 8.30 is reached, "Titration Completed" will appear with the result. The result is expressed in mg/L as calcium carbonate.
- Remove the pH electrode, temperature probe and stirrer from the sample and rinse them thoroughly with deionized water.
- Record the result.

ACIDITY OF WATER 0 to 2500 mg/L, pH 8.3 Endpoint

METHOD PARAMETERS

METHOD FARAMETERS	
Name:	Acidity of Water
Method Revision:	3.0
Analysis Type:	Standard Titration
Analog Board:	Analog 1
Stirrer Configuration:	:
Stirrer:	Stirrer 1
Stirring Speed:	1400 RPM
Pump Configuration:	
Titrant Pump:	Pump 1
Reagent Addition 1:	Disabled
Reagent Addition 2:	Disabled
Dosing Type:	Dynamic
Min Vol:	0.050 mL
Max Vol:	0.500 mL
delta E:	5.000 mV
End Point Mode:	Fixed 8.300 pH
Pre-Titration Volume:	: 0.000 mL
Pre-Titration Stir Ti	ime: 0 sec
Measurement Mode:	Signal Stability
delta E:	1.0 mV
delta t:	2 sec
Min wait:	2 sec
Max wait:	20 sec
Electrode Type:	рH
Blank Option:	No Blank
Calculations: Sam	ple Calc. by Volume
Dilution Option:	Disabled
Titrant Name:	0.1N NaOH
Titrant Conc.:	0.1000 N(eq/L)
Analyte Size:	50.000 mL
Analyte Entry:	Fixed
Maximum Titrant Volum	
2	2000.0 to 2000.0 mV
Volume/Flow Rate:	25 mL/50.0 mL/min
Signal Averaging:	1 Reading
Significant Figures:	XXXXX

CALCULATIONS

Calculations:	Sample	Calc.	by	Volume
Titrant units:			Ν	(eq/L)
Titrant volume do	sed:			V (L)
Final result unit	s:			(mg/L)
Titrant Conc.:		0.100	1 00	l(eq/L)
Sample/Titrant:		0.5	500	mol/eq
mw of sample:		100	0.09	9 g/mol
Sample Volume:				.000 mL
$\frac{\text{mg}}{\text{r}} \text{CaCO}_3 = \frac{\text{V(L)} * 1000}{1000}$	0 * 0.10 *	0.5 * 1	00.0	9 * 1000
$\underline{-}$ L L L L L L L L L L	5(0.0		

RESULTS

	Т	itration Report
Method	Name:	Acidity of Water
Time &	Date:	14:54 August 1, 2018
Report	ID:	Ti_00023

Titratio	n Results
Method Name:	Acidity of Water
Time & Date:	14:54 August 1, 2018
Analyte Size:	50.000 mL
End Point Volume:	5.879 mL
pH Fixed End Point:	8.300
Result:	588.43 (mg/L)
Initial & Final pH:	2.465 to 8.398
Titration Duration:	3:42 [mm:ss]
Titration went to C	ompletion

Analyst Signature:_____

CHLORIDE IN WATER 0 to 150 ppm (mg/L)

DESCRIPTION

Method for the determination of chloride in water. The results are expressed as **ppm (mg/L) as Chloride**.

REFERENCE

Standard Methods for the Examination of Water and Wastewater 21st edition, Method 4500-Cl

ELECTRODE

• HI4115 Silver/Sulfide Combination ISE

REAGENTS

- HI70448 0.02M Silver Nitrate (1 L)
- HI70427 1.5M Nitric Acid Solution (500 mL)
- HI70436 Deionized Water (1 gal)

ACCESSORIES

- HI7072 Electrode Fill Solution (4 x 30 mL)
- 150 mL Glass Beaker
- 100 mL Class A Volumetric Pipette
- 10 mL Class A Volumetric Pipette

DEVICE PREPARATION

- Connect the Silver/Sulfide electrode to the titrator.
- Install a 25 mL burette filled with 0.02M silver nitrate (HI70448) on pump one and verify that no air bubbles are present in the burette or tubing. If necessary prime the burette until all the air has been removed completely.
- For the determination of the exact concentration of the 0.02M Silver Nitrate, follow *HIO200EN 0.02M Silver Nitrate* Titrant Concentration
- Press Select Method from the main screen. Use the arrow keys to highlight H11007EN Chloride in Water and press Select

ELECTRODE PREPARATION

• Prepare the Silver/Sulfide electrode according to the procedure in the manual.

SAMPLE PREPARATION

- Use a class A volumetric pipette to transfer exactly 100.00 mL of sample to a clean 150 mL beaker.
- Add 10.00 mL of 1.5M nitric acid (HI70427) to the beaker.

ANALYSIS

 Place the beaker under the stirrer assembly and lower it to immerse the electrode and stirrer. Ensure that the reference junction of the electrode is 5 to 6 mm below the surface. If necessary add extra deionized water.

Note: The dispensing tip should be slightly submerged in the sample.

- Press start stop , the titrator will start the analysis.
- At the end of the titration, after detection of the equivalence point, "Titration Completed" will appear with the result. The result is expressed in ppm (mg/L) of chloride.
- Remove the electrode and stirrer from the sample and rinse them thoroughly with deionized water.
- Record the result.

CHLORIDE IN WATER 0 to 150 ppm (mg/L)

METHOD PARAMETERS

METHUD PAKAMETEKS	
Name:	Chloride in Water
Method Revision:	3.0
Analysis Type:	Standard Titration
Analog Board:	Analog 1
Stirrer Configuration:	:
Stirrer:	Stirrer 1
Stirring Speed:	1400 RPM
Pump Configuration:	
Titrant Pump:	Pump 1
Reagent Addition 1:	Disabled
Reagent Addition 2:	Disabled
Dosing Type:	Dynamic
Min Vol:	0.030 mL
Max Vol:	0.500 mL
delta E:	5.000 mV
End Point Mode: mv	
Recognition Options:	
Threshold:	100 mV/mL
Range:	NO
Filtered Derivativ	ves: NO
Pre-Titration Volume:	: 0.000 mL
Pre-Titration Stir T	ime: 0 sec
Measurement Mode:	Signal Stability
delta E:	1.0 mV
delta t:	2 sec
Min wait:	2 sec
Max wait:	20 sec
Electrode Type:	Silver/Sulfide
Blank Option:	No Blank
Calculations: Sam	ple Calc. by Volume
Dilution Option:	Disabled
Titrant Name:	0.02M AgNO3
Titrant Conc.:	2.0000E-2 M (mol/L)
Analyte Size:	100.0000 mL
Analyte Entry:	Manual
Maximum Titrant Volur	ne: 25.000 mL
Potential Range: -	2000.0 to 2000.0 mV
Volume/Flow Rate:	25 mL/50.0 mL/min
Signal Averaging:	1 Reading
Significant Figures:	XXXXX

CALCULATIONS

Calculations:	Sample Calc. by Volume
Titrant units:	M (mol/L)
Titrant volume d	losed: V (L)
Titrant Conc.:	2.0000E-2 M (mol/L)
Sample/Titrant:	1.000 mol/mol
mw of sample:	35.453 g/mol
Sample Volume:	100.000 mL
mg _ V(L) * 1000	* 0.02 * 1.0 * 35.45 * 1000
L	100.0
RESULTS	

RESULTS

	Т	itration Report
Method	Name:	Chloride in Water
Time &	Date:	15:11 August 1, 2018
Report	ID:	Ti_00052

Titration Results

Method Name:	Chloride in Water
Time & Date: 1	5:11 August 1, 2018
Analyte Size:	100.000 mL
End Point Volume:	4.781 mL
mV Equivalence Point	: 280.3
Result:	33.897 ppm (mg/L)
Initial & Final mV:	194.8 to 298.5
Titration Duration:	1:24 [mm:ss]
Titration went to Cor	mpletion

Analyst Signature:_____

NEUTRALIZATION WITH SULFURIC ACID 0 to 200 meg/L

DESCRIPTION

Method for the determination of strong or weak base concentration by titration of a sample to the equivalence point with sulfuric acid. The results are expressed as **meq/L**.

ELECTRODE

- HI1131B Combination pH Electrode
- HI7662-T Temperature Probe

REAGENTS

- HI70459 0.05M Sulfuric Acid (1 L)
- HI70436 Deionized Water (1 gal)

ACCESSORIES

- HI70300L Storage Solution (500 mL)
- HI7082 Electrode Fill Solution (4 x 30 mL)
- HI7004L pH 4.01 Buffer Solution
- HI7007L pH 7.01 Buffer Solution
- HI7010L pH 10.01 Buffer Solution
- HI740036P 100 mL Plastic Beaker (10 pcs)
- 10 mL Class A Volumetric Pipette

DEVICE PREPARATION

- Connect the pH electrode and temperature probe to the titrator.
- Install a 25 mL burette filled with 0.05M sulfuric acid (HI70459) on pump one and verify that no air bubbles are present in the burette or tubing. If necessary prime the burette until all the air has been removed completely.
- For the determination of the exact concentration of the 0.05M sulfuric acid, follow *HI0103EN 0.05M Sulfuric Acid* Titrant Concentration.
- Press Select from the main screen. Use the arrow keys to highlight H11008EN Neutralization w/H2SO4 and press Select

ELECTRODE PREPARATION

- Press Mode from the main screen, if necessary select the analog board and press PH
- Calibrate the electrode using pH 4.01, 7.01 and 10.01 buffers. Refer to the instruction manual for calibration procedure.

SAMPLE PREPARATION

- Use a class A volumetric pipette to transfer exactly 10.00 mL of sample to a clean 100 mL plastic beaker.
- Add deionized water to the 50 mL mark on the beaker.

ANALYSIS

• Place the beaker under the stirrer assembly and lower it to immerse the pH electrode, temperature probe and stirrer. Ensure that the reference junction of the pH electrode is 5 to 6 mm below the surface.

Note: The dispensing tip should be slightly submerged in the sample.____

- Press start stop the titrator will start the analysis.
- At the end of the titration, after detection of the equivalence point, "Titration Completed" will appear with the result. The result is expressed in meq/L.
- Remove the pH electrode, temperature probe and stirrer from the sample and rinse them thoroughly with deionized water.
- Record the result.

NEUTRALIZATION WITH SULFURIC ACID 0 to 200 meq/L

METHOD PARAMETERS

METHOD PARAMETERS	
Name:	Neutralization w/ H2SO4
Method Revision:	3.0
Analysis Type:	Standard Titration
Analog Board:	Analog 1
Stirrer Configura	tion:
Stirrer:	Stirrer 1
Stirring Spee	d: 1400 RPM
Pump Configuratio	n:
Titrant Pump:	Pump 1
Reagent Addition	1: Disabled
Reagent Addition	2: Disabled
Dosing Type:	Dynamic
Min Vol:	0.050 mL
Max Vol:	0.500 mL
delta E:	20.000 mV
End Point Mode:	pH 1EQ point, 1st Der
Recognition Opti	
Threshold:	50 mV/mL
Range:	NO
Filtered Deri	vatives: NO
Pre-Titration Vo	lume: 0.000 mL
Pre-Titration Vo Pre-Titration St	
	ir Time: 0 sec
Pre-Titration St	ir Time: 0 sec
Pre-Titration St Measurement Mode	ir Time: 0 sec : Signal Stability 1.0 mV
Pre-Titration St Measurement Mode delta E:	ir Time: 0 sec : Signal Stability 1.0 mV 2 sec
Pre-Titration St Measurement Mode delta E: delta t: Min wait:	ir Time: 0 sec : Signal Stability 1.0 mV 2 sec 2 sec
Pre-Titration St Measurement Mode delta E: delta t: Min wait: Max wait:	ir Time: 0 sec : Signal Stability 1.0 mV 2 sec 2 sec 15 sec
Pre-Titration St Measurement Mode delta E: delta t: Min wait: Max wait: Electrode Type:	ir Time: 0 sec : Signal Stability 1.0 mV 2 sec 2 sec 15 sec pH
Pre-Titration St Measurement Mode delta E: delta t: Min wait: Max wait: Electrode Type: Blank Option:	ir Time: 0 sec : Signal Stability 1.0 mV 2 sec 2 sec 15 sec pH No Blank
Pre-Titration St Measurement Mode delta E: delta t: Min wait: Max wait: Electrode Type: Blank Option: Calculations:	ir Time: 0 sec : Signal Stability 1.0 mV 2 sec 2 sec 15 sec pH
Pre-Titration St Measurement Mode delta E: delta t: Min wait: Max wait: Electrode Type: Blank Option:	ir Time: 0 sec : Signal Stability 1.0 mV 2 sec 2 sec 15 sec pH No Blank Sample Calc. by Volume
Pre-Titration St Measurement Mode delta E: delta t: Min wait: Max wait: Electrode Type: Blank Option: Calculations: Dilution Option:	ir Time: 0 sec : Signal Stability 1.0 mV 2 sec 2 sec 15 sec pH No Blank Sample Calc. by Volume Disabled 0.05M H2S04
Pre-Titration St Measurement Mode delta E: delta t: Min wait: Max wait: Electrode Type: Blank Option: Calculations: Dilution Option: Titrant Name: Titrant Conc.:	ir Time: 0 sec : Signal Stability 1.0 mV 2 sec 2 sec 15 sec pH No Blank Sample Calc. by Volume Disabled
Pre-Titration St Measurement Mode delta E: delta t: Min wait: Electrode Type: Blank Option: Calculations: Dilution Option: Titrant Name: Titrant Conc.: Analyte Size:	ir Time: 0 sec : Signal Stability 1.0 mV 2 sec 2 sec 15 sec pH No Blank Sample Calc. by Volume Disabled 0.05M H2S04 5.0000E-2 M (mol/L)
Pre-Titration St Measurement Mode delta E: delta t: Min wait: Max wait: Electrode Type: Blank Option: Calculations: Dilution Option: Titrant Name: Titrant Conc.:	ir Time: 0 sec : Signal Stability 1.0 mV 2 sec 2 sec 15 sec pH No Blank Sample Calc. by Volume Disabled 0.05M H2S04 5.0000E-2 M (mol/L) 10.000 mL Fixed
Pre-Titration St Measurement Mode delta E: delta t: Min wait: Electrode Type: Blank Option: Calculations: Dilution Option: Titrant Name: Titrant Conc.: Analyte Size: Analyte Entry: Maximum Titrant	ir Time: 0 sec : Signal Stability 1.0 mV 2 sec 2 sec 15 sec pH No Blank Sample Calc. by Volume Disabled 0.05M H2S04 5.0000E-2 M (mol/L) 10.000 mL Fixed Volume: 20.000 mL
Pre-Titration St Measurement Mode delta E: delta t: Min wait: Electrode Type: Blank Option: Calculations: Dilution Option: Titrant Name: Titrant Conc.: Analyte Size: Analyte Entry: Maximum Titrant	ir Time: 0 sec : Signal Stability 1.0 mV 2 sec 2 sec 15 sec pH No Blank Sample Calc. by Volume Disabled 0.05M H2S04 5.0000E-2 M (mol/L) 10.000 mL Fixed Volume: 20.000 mL -2000.0 to 2000.0 mV
Pre-Titration St Measurement Mode delta E: delta t: Min wait: Electrode Type: Blank Option: Calculations: Dilution Option: Titrant Name: Titrant Conc.: Analyte Size: Analyte Entry: Maximum Titrant Potential Range: Volume/Flow Rate	ir Time: 0 sec : Signal Stability 1.0 mV 2 sec 2 sec 15 sec pH No Blank Sample Calc. by Volume Disabled 0.05M H2S04 5.0000E-2 M (mol/L) 10.000 mL Fixed Volume: 20.000 mL -2000.0 to 2000.0 mV : 25 mL/50.0 mL/min
Pre-Titration St Measurement Mode delta E: delta t: Min wait: Electrode Type: Blank Option: Calculations: Dilution Option: Titrant Name: Titrant Conc.: Analyte Size: Analyte Entry: Maximum Titrant Potential Range:	ir Time: 0 sec : Signal Stability 1.0 mV 2 sec 2 sec 15 sec pH No Blank Sample Calc. by Volume Disabled 0.05M H2S04 5.0000E-2 M (mol/L) 10.000 mL Fixed Volume: 20.000 mL -2000.0 to 2000.0 mV : 25 mL/50.0 mL/min : 1 Reading

CALCULATIONS

Calculations:Sample Calc. by VoluTitrant units:M (mol/Titrant volume dosed:VFinal result units:medTitrant Conc.:5.0000E-2 M (mol/Sample/Titrant:2.000 eq/nSample Volume:10.000	/L) (L) g/L /L) nol
$\frac{\text{meq}}{\text{meq}} = \frac{V(\text{L}) * 1000 * 0.05 * 2.0 * 1000}{0.05 * 2.0 * 1000}$	
L 10.0	
RESULTS	
Titration Report	
Method Name: Neutralization w/ H2S Time & Date: 09:46 August 1, 20 Report ID: Ti_000)18
Titration Results	

Method Name:	Neutralization w/ H2SO4
Time & Date:	09:46 August 1, 2018
Analyte Size:	10.000 mL
End Point Volum	ne: 9.562 mL
pH Equivalence	Point: 7.966
Result:	95.620 meq/L
Initial & Final	pH: 11.655 to 6.248
Titration Durat	ion: 3:26 [mm:ss]
Titration went	to Completion

Analyst Signature:_____

NEUTRALIZATION WITH SODIUM HYDROXIDE 0 to 200 meq/L

DESCRIPTION

Method for the determination of strong or weak acid concentration by titration of a sample to the equivalence point with sodium hydroxide. The results are expressed as **meq/L**.

ELECTRODE

- HI1131B Combination pH Electrode
- HI7662-T Temperature Probe

REAGENTS

- HI70456 0.1N Sodium Hydroxide (1 L)
- HI70436 Deionized Water (1 gal)

ACCESSORIES

- HI70300L Storage Solution (500 mL)
- HI7082 Electrode Fill Solution (4 x 30 mL)
- HI7004L pH 4.01 Buffer Solution
- HI7007L pH 7.01 Buffer Solution
- HI7010L pH 10.01 Buffer Solution
- HI740036P 100 mL Plastic Beakers (10 pcs)
- 10 mL Class A Volumetric Pipette

DEVICE PREPARATION

- Connect the pH electrode and temperature probe to the titrator.
- Install a 25 mL burette filled with 0.1N sodium hydroxide (HI70456) on pump one and verify that no air bubbles are present in the burette or tubing. If necessary prime the burette until all the air has been removed completely.
- For the determination of the exact concentration of the 0.1N sodium hydroxide, follow *HI0001EN 0.1N Sodium Hydroxide* Titrant Concentration.
- Press Select Method from the main screen. Use the arrow keys to highlight H11009EN Neutralization w/NaOH and press Select

ELECTRODE PREPARATION

- Press Mode from the main screen, if necessary select the analog board and press PH
- Calibrate the electrode using pH 4.01, 7.01 and 10.01 buffers. Refer to the instruction manual for calibration procedure.

SAMPLE PREPARATION

- Use a class A volumetric pipette to transfer exactly 10.00 mL of sample to a clean 100 mL plastic beaker.
- Add deionized water to the 50 mL mark on the beaker.

ANALYSIS

• Place the beaker under the stirrer assembly and lower it to immerse the pH electrode, temperature probe and stirrer. Ensure that the reference junction of the pH electrode is 5 to 6 mm below the surface.

Note: The dispensing tip should be slightly submerged in the sample.

- Press start stop , the titrator will start the analysis.
- At the end of the titration, after detection of the equivalence point, "Titration Completed" will appear with the result. The result is expressed in meq/L.
- Remove the pH electrode, temperature sensor and stirrer from the sample and rinse them thoroughly with deionized water.
- Record the result.

NEUTRALIZATION WITH SODIUM HYDROXIDE 0 to 200 meq/L

METHOD PARAMETERS

METHUD FARAMETERS	
Name:	Neutralization w/ NaOH
Method Revision:	3.0
Analysis Type:	Standard Titration
Analog Board:	Analog 1
Stirrer Configurat	ion:
Stirrer:	Stirrer 1
Stirring Speed	1: 1400 RPM
Pump Configuration	
Titrant Pump:	Pump 1
Reagent Addition	1: Disabled
Reagent Addition	
Dosing Type:	Dynamic
Min Vol:	0.050 mL
Max Vol:	0.500 mL
delta E:	20.000 mV
End Point Mode:	pH 1EQ point, 1st Der
Recognition Optic	
Threshold:	50 mV/mL
Range:	NO
Filtered Deriv	vatives: NO
Pre-Titration Vol	
Pre-Titration Sti	
Measurement Mode:	
delta E:	1.0 mV
delta t:	2 sec
Min wait:	2 sec
Max wait:	15 sec
Electrode Type:	Hq
Blank Option:	No Blank
Calculations:	Sample Calc. by Volume
Dilution Option:	Disabled
Titrant Name:	0.1N NaOH
Titrant Conc.:	0.1000 N(eq/L)
Analyte Size:	10.000 mL
Analyte Entry:	Fixed
Maximum Titrant V	
	-2000.0 to 2000.0 mV
Volume/Flow Rate:	
Signal Averaging:	
Significant Figure	_

CALCULATIONS

Sample	Calc.	by	Volume
		Ν	(eq/L)
osed:			V (L)
cs:			meq/L
	0.10	1 00	l(eq/L)
		10.	.000 mL
.000 * 0.1	* 1.0 *	100	00
10.0	сС		
	osed: cs: .000 * 0.1	osed: cs: 0.10	osed: cs: 0.1000 M 10. .000 * 0.1 * 1.0 * 100

RESULTS

	Titra	ation Report
Method	Name:	Neutralization w/ NaOH
Time &	Date:	10:29 August 2, 2018
Report	ID:	Ti_00017

Titration Results

Time & Date: 10:29 August 2, 2018
Analyte Size: 10.000 mL
End Point Volume: 15.970 mL
pH Equivalence Point: 8.431
Result: 159.70 meq/L
Initial & Final pH: 2.675 to 10.316
Titration Duration: 3:20 [mm:ss]
Titration went to Completion

Analyst Signature:_____

DESCRIPTION

Method for verifying the dosing and potentiometric signal accuracy of the titrator. This method should be used to troubleshoot a titrator equipped with a 25 mL burette. The titrator dispenses a 20.00 mL pre-titration volume, waits 20 seconds and dispenses an additional 20.00 mL dose, bringing the total volume to 40.00 mL. This procedure can also be used to check the stability of the mV and temperature channels.

The specifications of the dosing accuracy are \pm 0.1% of the full burette volume (\pm 0.025 mL for a 25 mL burette). For the accuracy of other burette volumes, see the instruction manual.

If the results are not correct, check all fittings for leakage, and burette and tubing for air bubbles. Repeat the measurement.

REFERENCE

ISO/TC 48/SC1N 380E and 383E: "Piston and/or Plunger Operated Volumetric Apparatus"

ACCESSORIES

- HI762000C 0°C Temperature Key
- HI762070C 70°C Temperature Key
- HI70436 Deionized Water (1 gal)
- HI7662-T Temperature Probe
- Shorting Cap
- Narrow Neck Beaker
- Analytical Balance with 0.0001g resolution

DEVICE PREPARATION

- Connect the shorting cap to the BNC socket on Analog Board 1
- Install a 25 mL burette filled with room temperature deionized water (HI70436) on pump one and verify that no air bubbles are present in the burette or tubing. If necessary prime the burette until all the air has been removed completely.
- Press Select Method
 from the main screen. Use the arrow keys to highlight H11011EN Troubleshooting 1 and press

LARGE DOSE DISPENSING PROCEDURE

- Add a small amount of deionized water to a narrow neck beaker. By doing this the air space in the beaker will be vapor-saturated minimizing evaporation.
- Place the narrow neck beaker on an analytical balance.
- Zero the balance.
- Place the dosing tip through the neck of the beaker. Take care
 not to immerse it in the liquid during dispensing and not to
 touch the beaker walls.
- Press start stop
- Write down the exact weight displaced on the balance after each dose.

- The following information is needed to verify the accuracy of the dosing system:
 - The temperature of the dispensed water
 - The atmospheric air pressure
 - The density of the weight used to calibrate the balance
- This procedure can be repeated on pump 2.

Other burette sizes can be checked using the following settings:

Burette Volume	Pre-titration Volume	Max. Titrant Volume
5 mL	4.000 mL	8.000 mL
10 mL	8.000 mL	16.000 mL

METHOD PARAMETERS

Name:	Troubleshooting 1
Method Revision:	3.0
Analysis Type:	Standard Titration
Analog Board:	Analog 1
Stirrer Configuration	n:
Stirrer:	Stirrer 1
Stirring Speed:	0 RPM
Pump Configuration:	
Titrant Pump:	Pump 1
Reagent Addition 1:	Disabled
Reagent Addition 2:	Disabled
Dosing Type:	Linear - 20.000 mL
End Point Mode:	Fixed 10.0 mV
Pre-Titration Volume	e: 20.000 mL
Pre-Titration Stir 5	Time: 0 sec
Measurement Mode:	Timed Increment
Time interval:	20 sec
Electrode Type:	Shorting Cap
Blank Option:	No Blank
Calculations: N	o Formula (mL only)
Titrant Name:	DI Water
Maximum Titrant Volu	
Potential Range: -2	2000.0 to 2000.0 mV
Volume/Flow Rate:	25 mL/50.0 mL/min
Signal Averaging:	1 Reading
Significant Figures:	XXXXX

CALCULATIONS

The measured volume of the dispensed liquid is calculated from the measure mass using the following equation:

$$V = m \star \frac{1}{\rho} \star \left(1 + \frac{\rho_{\text{air}}}{\rho_{\text{L}}} - \frac{\rho_{\text{air}}}{\rho_{\text{std}}} \right)$$

- V Volume of measure mass of water (mL)
- m Measure mass of water (g)
- ρ_1 Density of dispensed water (g/mL)
- $\rho_{\rm our}$ Density of ambient air (g/mL)
- $\rho_{\mbox{\tiny std}}^{\mbox{\tiny std}}$ Density of calibration standard weight (g/mL)

ALTERNATIVE CALCULATIONS

If the actual values of the above parameters are not accessible the following equation can be used:

 $V = M^*F$

V Volume of measured mass of water (mL)

F Transformation factor

The transformation factor takes into account the air buoyancy, the water density and temperature dependence. Standard values can be used to obtain the transformation factor.

The values from the table below have been calculated by correcting the air and water density with temperature, assuming the density of dry air $\rho_{\text{air}}=0.0012$ g/mL and density of calibration steel standard weigh $\rho_{\text{STD}}=8$ g/mL.

Temperature (°C)	Factor
17.0	1.002290
18.0	1.002467
19.0	1.002654
20.0	1.002853
21.0	1.003061
22.0	1.003282
23.0	1.003512
24.0	1.003752
25.0	1.004002
26.0	1.004261
27.0	1.004531
28.0	1.004809
29.0	1.005097
30.0	1.005395

TEMPERATURE CHANNEL FAST CHECK PROCEDURE

- Connect the shorting cap to the BNC socket on Analog Board 1.
- Connect the HI762000C 0°C temperature key to the RCA socket (temperature sensor input) on Analog Board 1.
- On the main screen select Mode, if necessary select the analog board and press mv.
- The titrator should display ATC 0.0 \pm 0.4 $^{\circ}\text{C}$ with no fluctuations or drift.
- Connect the HI762070C 70°C temperature key to the RCA socket (temperature sensor input) on Analog Board 1.
- The titrator should display ATC 70.0 \pm 0.4°C with no fluctuations or drift.
- This procedure can be repeated on analog board 2.

TEMPERATURE & MV CHANNEL LOGGING PROCEDURE

- Connect the shorting cap to the BNC socket on Analog Board 1.
- Connect the HI762000C 0°C temperature key to the RCA socket (temperature sensor input) on Analog Board 1.
- On the main screen select Mode if necessary select the analog board and press mv.
- Press mV/setup and use the arrow keys to highlight Logging Interval. Set the logging interval to 15 seconds and press Accept. Press Escape to return to the main screen.
- Press the results key and use the arrow keys to highlight *Setup* pH/mV/ISE Report, press Select .
- Select *Potential and Temperature and Units* (the selected fields are marked with an *). All other fields should be unselected.
- Press Save Report to return to the Data Parameters screen.
- Press Escape to return to the main screen.
- Once on the main screen press <u>Start</u> Log to start the automatic log.
- Let the log run for about 10 minutes. Press Log to stop the automatic log.
- Press results, use the arrow keys to highlight *Review Last* Analysis Report, and press select
- The mV column should display 0.0 \pm 0.1 mV and the temperature column should display 0.0°C \pm 0.4°C.
- This procedure can be repeated using the HI762070C 70°C temperature key and on analog board 2.

DESCRIPTION

Method for verifying the dosing of the titrator. This method should be used to troubleshoot a titrator equipped with a 25 mL burette. The titrator dispenses a 10.00 mL pre-titration volume, waits 20 seconds and dispenses an additional 0.5 mL dose twenty times, waiting 20 seconds between each dose, bringing the total volume to 20 mL. This procedure can also be used to check the stirrer functionality.

The specifications of the dosing accuracy are \pm 0.1% of the full burette volume (\pm 0.025 mL for a 25 mL burette). For the accuracy of other burette volumes, see the instruction manual.

If the results are not correct, check all fittings for leakage, and burette and tubing for air bubbles. Repeat the measurement.

REFERENCE

ISO/TC 48/SC1N 380E and 383E: "Piston and/or Plunger Operated Volumetric Apparatus"

ACCESSORIES

- HI762000C 0°C Temperature Key
- HI70436 Deionized Water (1 gal)
- HI7662-T Temperature Probe
- Shorting Cap
- Narrow Neck Beaker
- Analytical Balance with a resolution of 0.0001g

DEVICE PREPARATION

- Connect the shorting cap to the BNC socket on Analog Board 1
- Install a 25 mL burette filled with room temperature deionized water (HI70436) on pump one and verify that no air bubbles are present in the burette or tubing. If necessary prime the burette until all the air has been removed completely.
- Press Select Method from the main screen. Use the arrow keys to highlight *H11012EN Troubleshooting 2* and press Select .

SMALL DOSE DISPENSING PROCEDURE

- Add a small amount of deionized water to a narrow neck beaker. By doing this the air space in the beaker will be vapor-saturated minimizing evaporation.
- Place the narrow neck beaker on an analytical balance.
- Zero the balance.
- Place the dosing tip through the neck of the beaker. Take care
 not to immerse it in the liquid during dispensing and not to
 touch the beaker walls.
- Press
 start
 stop
- Write down the exact weight displaced on the balance after each dose.

- The following information is needed to verify the accuracy of the dosing system:
 - The temperature of the dispensed water
 - The atmospheric air pressure
 - The density of the weight used to calibrate the balance
- This procedure can be repeated on pump 2.

Other burette sizes can be checked using the following settings:

Burette Volume	Pre-titration Volume	Max. Titrant Volume
5 mL	4.000 mL	8.000 mL
10 mL	8.000 mL	16.000 mL

METHOD PARAMETERS

Name:	Troubleshooting 2
Method Revision:	3.0
Analysis Type:	Standard Titration
Analog Board:	Analog 1
Stirrer Configuration	n:
Stirrer:	Stirrer 1
Stirring Speed:	0 RPM
Pump Configuration:	
Titrant Pump:	Pump 1
Reagent Addition 1:	Disabled
Reagent Addition 2:	Disabled
Dosing Type:	Linear - 0.500 mL
End Point Mode:	Fixed 10.0 mV
Pre-Titration Volume	e: 10.000 mL
Pre-Titration Stir 7	Time: 0 sec
Measurement Mode:	Timed Increment
Time interval	10 sec
Electrode Type:	Shorting Cap
Blank Option:	No Blank
Calculations: N	o Formula (mL only)
Titrant Name:	DI Water
Maximum Titrant Volu	ume: 20.000 mL
Potential Range: -2	2000.0 to 2000.0 mV
Volume/Flow Rate:	25 mL/50.0 mL/min
Signal Averaging:	1 Reading
Significant Figures:	XXXXX

CALCULATIONS

The measured volume of the dispensed liquid is calculated from the measure mass using the following equation:

$$V = m * \frac{1}{\rho} * \left(1 + \frac{\rho_{\text{air}}}{\rho_{\text{L}}} - \frac{\rho_{\text{air}}}{\rho_{\text{std}}} \right)$$

- V Volume of measure mass of water (mL)
- m Measure mass of water (g)
- $\rho_{\rm L}$ Density of dispensed water (g/mL)
- $\rho_{\rm our}$ Density of ambient air (g/mL)
- $\rho_{\text{std}}^{\text{\tiny ST}}$ Density of calibration standard weight (g/mL)

ALTERNATIVE CALCULATIONS

If the actual values of the above parameters are not accessible the following equation can be used:

 $V = M^*F$

Volume of measured mass of water (mL) V

F Transformation factor

The transformation factor takes into account the air buoyancy, the water density and temperature dependence. Standard values can be used to obtain the transformation factor.

The values from the table below have been calculated by correcting the air and water density with temperature, assuming the density of dry air $\rho_{\mbox{\tiny nir}}=$ 0.0012 g/mL and density of calibration steel standard weigh $\rho_{\text{STD}} = 8$ g/mL.

Temperature (°C)	Factor		
17.0	1.002290		
18.0	1.002467		
19.0	1.002654		
20.0	1.002853		
21.0	1.003061		
22.0	1.003282		
23.0	1.003512		
24.0	1.003752		
25.0	1.004002		
26.0	1.004261		
27.0	1.004531		
28.0	1.004809		
29.0	1.005097		
30.0	1.005395		

STIRRING SPEED FAST CHECK PROCEDURE

- On the main screen select Mode , if necessary select the analog board and press mv
- Press <u>mv</u> and use the arrow keys to highlight *Stirrer Configuration*. Use the arrow keys to highlight *Stirrer* 1. Press Accept
- Use the arrow keys to highlight *Strring Speed*. Use the numeric • keypad to enter 200 rpms then press
- Press Escape to exit the mV Setup screen. From the main screen, press stir , use the up arrow key to • increase the stir speed slowly to 2500 rpms.
- Check that the propeller continues to increase speed, following the commands.
- This procedure can be repeated on stirrer 2. •

CONCENTRATION OF PHOSPHORIC ACID 0.00 to 0.01 M (mol/L)

DESCRIPTION

Method for the determination of phosphoric acid (H_3PO_4) , by titration of a sample to the point of inflection with sodium hydroxide.

The first inflection point corresponds to the H_3PO_4 content and the difference between the first and second corresponds to $H_2PO_4^-$. The results are express as **M (mol/L) phosphoric acid**.

If only phosphoric acid and no other acids or bases are present in the sample, then $H_3PO_4 = H_2PO_4$. If H_3PO_4 is greater than $H_2PO_4^-$ this means other weak acids or bases are present (e.g. citric acid / citrate or ascorbic acid / ascorbate).

ELECTRODE

- HI1131B Combination pH Electrode
- HI7662-T Temperature Probe

REAGENTS

- HI70456 0.1N Sodium Hydroxide (1 L)
- HI70436 Deionized Water (1 gal)

ACCESSORIES

- HI70300L Storage Solution (500 mL)
- HI7082 Electrode Fill Solution (4 x 30 mL)
- HI7004L pH 4.01 Buffer Solution (500 mL)
- HI7007L pH 7.01 Buffer Solution (500 mL)
- HI7010L pH 10.01 Buffer Solution (500 mL)
- 100 mL Class A Volumetric Pipette
- 150 mL Glass Beaker

DEVICE PREPARATION

- Connect the pH electrode and temperature probe to the titrator.
- Install a 25 mL burette filled with 0.1N sodium hydroxide (H170456) on pump one and verify that no air bubbles are present in the burette or tubing. If necessary prime the burette until all the air has been removed completely.
- For the determination of the exact concentration of the 0.1N sodium hydroxide, follow *HI0001EN 0.1N Sodium Hydroxide* Titrant <u>Concentration</u>.
- Press Select Method
 from the main screen. Use the arrow keys to highlight H11014EN Concentration of H3P04 and press Select

ELECTRODE PREPARATION

- Press Mode from the main screen, if necessary select the analog board and press PH
- Calibrate the electrode using pH 4.01, 7.01 and 10.01 buffers. Refer to the instruction manual for calibration procedure.

SAMPLE PREPARATION

• Use a Class A volumetric pipette to transfer exactly 100.00 mL of sample to a clean 150 mL glass beaker.

ANALYSIS

• Place the beaker under the stirrer assembly and lower it to immerse the pH electrode, temperature probe and stirrer. Ensure that the reference junction of the pH electrode is 5 to 6 mm below the surface.

Note: The dispensing tip should be slightly submerged in the sample.

- Press start stop , the titrator will start the analysis.
- At the end of the titration, after the detection of the second equivalence point, "Titration Completed" will appear with the phosphoric acid concentration. The result is expressed in M (mol/L) of phosphoric acid.
- Remove the pH electrode, temperature probe and stirrer from the sample and rinse them thoroughly with deionized water.
- Record the result.

CONCENTRATION OF PHOSPHORIC ACID 0.00 to 0.01 M (mol/L)

METHOD PARAMETERS

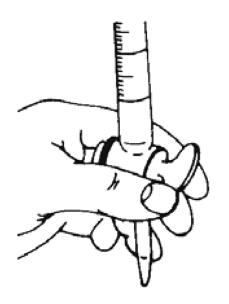
Name:	Concentration of H3PO4	
Method Revision:	3.0	
Analysis Type:	Standard Titration	
Analog Board:	Analog 1	
Stirrer Configurat	zion:	
Stirrer:	Stirrer 1	
Stirring Speed	d: 1400 RPM	
Pump Configuration	1:	
Titrant Pump:	Pump 1	
Reagent Addition	1: Disabled	
Reagent Addition	2: Disabled	
Dosing Type:	Dynamic	
Min Vol:	0.030 mL	
Max Vol:	0.500 mL	
delta E:	8.000 mV	
End Point Mode:	pH 2EQ points, 1st Der	
Recognition Optic		
Threshold:	50 mV/mL	
Range:	NO	
Filtered Deriv	vatives: NO	
Pre-Titration Vol	ume: 0.000 mL	
Pre-Titration Sti	r Time: 10 sec	
Measurement Mode:		
Measurement Mode: delta E:		
	Signal Stability	
delta E:	Signal Stability 0.8 mV	
delta E: delta t:	Signal Stability 0.8 mV 2 sec	
delta E: delta t: Min wait: Max wait:	Signal Stability 0.8 mV 2 sec 2 sec	
delta E: delta t: Min wait: Max wait: Electrode Type:	Signal Stability 0.8 mV 2 sec 2 sec 20 sec	
delta E: delta t: Min wait: Max wait:	Signal Stability 0.8 mV 2 sec 2 sec 20 sec pH	
delta E: delta t: Min wait: Max wait: Electrode Type: Blank Option:	Signal Stability 0.8 mV 2 sec 2 sec 20 sec pH No Blank	
delta E: delta t: Min wait: Max wait: Electrode Type: Blank Option: Calculations:	Signal Stability 0.8 mV 2 sec 2 sec 20 sec pH No Blank Sample Calc. by Volume	
delta E: delta t: Min wait: Max wait: Electrode Type: Blank Option: Calculations: Dilution Option:	Signal Stability 0.8 mV 2 sec 2 sec 20 sec pH No Blank Sample Calc. by Volume Disabled	
delta E: delta t: Min wait: Max wait: Electrode Type: Blank Option: Calculations: Dilution Option: Titrant Name: Titrant Conc.:	Signal Stability 0.8 mV 2 sec 2 sec 20 sec pH No Blank Sample Calc. by Volume Disabled 0.1N NaOH	
delta E: delta t: Min wait: Max wait: Electrode Type: Blank Option: Calculations: Dilution Option: Titrant Name:	Signal Stability 0.8 mV 2 sec 2 sec 20 sec pH No Blank Sample Calc. by Volume Disabled 0.1N NaOH 0.1000 N(eq/L)	
delta E: delta t: Min wait: Max wait: Electrode Type: Blank Option: Calculations: Dilution Option: Titrant Name: Titrant Conc.: Analyte Size:	Signal Stability 0.8 mV 2 sec 2 sec 20 sec pH No Blank Sample Calc. by Volume Disabled 0.1N NaOH 0.1000 N(eq/L) 100.000 mL Fixed	
delta E: delta t: Min wait: Max wait: Electrode Type: Blank Option: Calculations: Dilution Option: Titrant Name: Titrant Name: Titrant Conc.: Analyte Size: Analyte Entry: Maximum Titrant W	Signal Stability 0.8 mV 2 sec 2 sec 20 sec pH No Blank Sample Calc. by Volume Disabled 0.1N NaOH 0.1000 N(eq/L) 100.000 mL Fixed	
delta E: delta t: Min wait: Max wait: Electrode Type: Blank Option: Calculations: Dilution Option: Titrant Name: Titrant Name: Titrant Conc.: Analyte Size: Analyte Entry: Maximum Titrant W	Signal Stability 0.8 mV 2 sec 2 sec 20 sec pH No Blank Sample Calc. by Volume Disabled 0.1N NaOH 0.1000 N(eq/L) 100.000 mL Fixed Volume: 20.000 mL -2000.0 to 2000.0 mV	
delta E: delta t: Min wait: Max wait: Electrode Type: Blank Option: Calculations: Dilution Option: Titrant Name: Titrant Conc.: Analyte Size: Analyte Entry: Maximum Titrant W Potential Range:	Signal Stability 0.8 mV 2 sec 2 sec 20 sec pH No Blank Sample Calc. by Volume Disabled 0.1N NaOH 0.1000 N(eq/L) 100.000 mL Fixed Volume: 20.000 mL -2000.0 to 2000.0 mV 25 mL/50.0 mL/min	
delta E: delta t: Min wait: Max wait: Electrode Type: Blank Option: Calculations: Dilution Option: Titrant Name: Titrant Conc.: Analyte Size: Analyte Entry: Maximum Titrant W Potential Range: Volume/Flow Rate:	Signal Stability 0.8 mV 2 sec 2 sec 20 sec pH No Blank Sample Calc. by Volume Disabled 0.1N NaOH 0.1000 N(eq/L) 100.000 mL Fixed Volume: 20.000 mL -2000.0 to 2000.0 mV 25 mL/50.0 mL/min 1 Reading	

CALCULATIONS

Calculations: Sample Calc. by Volume Titrant units: N (eq/L) Titrant volume dosed: V (L) Final result units: M (mol/L) Titrant Conc.: 0.1000 N(eq/L) Sample/Titrant: 1.000 mol/eq Sample Volume: 100.000 mL			
$\frac{\text{mol}}{\text{L}} = \frac{\text{V(L)} * 100 * 0.1 * 1.0}{100.00}$			
L 100.00			
RESULTS			
Titration Report			
Method Name: Concentration of H3PO4			
Time & Date: 11:56 August 2, 2018			
Report ID: Ti_00034			
Titration Results			
Method Name: Concentration of H3P04			
Time & Date: 11:56 August 2, 2018			
Analyte Size: 100.000 mL			
Equivalence point 1:			
DH			
pH: 4.677			
Volume: 4.397 mL			
Volume: 4.397 mL Result: 4.3972E-03 M (mol/L)			
Volume: 4.397 mL Result: 4.3972E-03 M (mol/L) Equivalence point 2:			
Volume: 4.397 mL Result: 4.3972E-03 M (mol/L) Equivalence point 2: pH: 8.916			
Volume: 4.397 mL Result: 4.3972E-03 M (mol/L) Equivalence point 2: pH: 8.916			
Volume: 4.397 mL Result: 4.3972E-03 M (mol/L) Equivalence point 2: pH: 8.916 Volume: 4.429 mL			
Volume: 4.397 mL Result: 4.3972E-03 M (mol/L) Equivalence point 2: pH: 8.916 Volume: 4.429 mL Result: 4.4293E-03 M (mol/L)			

HI932/HI931

AUTOMATIC POTENTIOMETRIC TITRATOR







1. GENERAL REVIEW OF TITRATION THEORY	
1.1. INTRODUCTION TO TITRATIONS	5
1.2 USES OF TITRATIONS	5
1.3. ADVANTAGES AND DISADVANTAGES OF TITRATIONS	5
2. TYPES OF TITRATION	6
2.1. TITRATIONS ACCORDING TO THE MEASUREMENT METHOD	6
2.1.1. AMPEROMETRIC TITRATIONS	
2.1.2. POTENTIOMETRIC TITRATIONS	6
2.1.3. SPECTROPHOTOMETRIC TITRATIONS	7
2.2. TITRATIONS ACCORDING TO THE REACTION TYPE	8
2.2.1. ACID-BASE TITRATIONS	
2.2.2. ARGENTOMETRIC TITRATIONS	9
2.2.3. COMPLEXOMETRIC TITRATIONS	9
2.2.4. ION SELECTIVE TITRATIONS	
2.2.5. NON-AQUEOUS SOLVENT ACID-BASE TITRATIONS	
2.2.6. PRECIPITATION TITRATIONS	
2.2.7. REDOX TITRATIONS	11
2.2.8. KARL FISCHER TITRATION	
2.3. TITRATIONS ACCORDING TO THE TITRATION SEQUENCE	
2.3.1. BACK TITRATIONS	
2.3.2. MULTIPLE ENDPOINT TITRATIONS	
3. INTRODUCTION TO TITRATION APPARATUS AND TYPICAL TITRATION PROCEDURE	13
3.1. MANUAL TITRATION	
3.2. AUTOMATIC TITRATION	
4. TITRATION RESULTS	
4.1. ACCURACY	14
4.2. REPEATABILITY	
4.3. SOURCES OF ERROR	14
4.3.1. SAMPLING ERRORS	
4.3.2. ERRORS WITH TITRANT AND STANDARD	
4.3.2.1. PREPARATION ERRORS	
4.3.2.2. DISPENSING ERRORS	
4.3.3. CHEMICAL REACTION ERRORS	
4.3.4. ENDPOINT DETERMINATION ERRORS	15
5. CALCULATIONS	
5.1. SAMPLE CALCULATION	16
5.2. STANDARDIZE TITRANT	
5.3. BLANK TITRATION	
5.4. MULTIPLE ENDPOINT TITRATION	
5.5. BACK TITRATION	18
6. GLOSSARY	18

TITRATION THEORY

1. GENERAL REVIEW OF TITRATION THEORY

1.1. INTRODUCTION TO TITRATIONS

A titration is a quantitative, volumetric procedure used in analytical chemistry to determine the concentration of an analyte (the species being measured) in solution. The concentration of the analyte is determined by slowly adding a titrant (reagent) to the solution. As the titrant is added, a chemical reaction occurs between the titrant and the analyte.

Titration reactions are relatively fast, simple reactions that can be expressed using a chemical equation. The titration reaction continues as the titrant is added until all of the analyte is consumed and the analyte reacts completely and quantitatively with the titrant.

The point at which all of the analyte has been reacted is called the equivalence point, also known as the theoretical or stoichiometric endpoint. This point is accompanied by an abrupt physical change in the solution, which sharply defines the endpoint of the reaction. The physical change associated with the titration endpoint can be produced by the titrant or an indicator and can be detected either visually or by some other physical measurement.

Titrations cannot be used to determine the quantity of all analytes. The chemical reaction between the titrant and analyte must fulfill four requirements:

- The reaction must be fast and occur within approximately one second after the titrant is added
- The reaction must go to completion
- The reaction must have well-known stoichiometry (reaction ratios)
- A convenient endpoint or inflection point

Titrations are highly precise and can provide many advantages over alternative methods. Titrations are quickly performed and require relatively simple apparatus and instrumentation.

1.2. USES OF TITRATIONS

Titrations can be used in many applications, including:

- Acid content of plant effluents, food (e.g.: cheese and wine), plating and etching baths, petroleum products, drugs
- Base content of fertilizer (containing ammonia), bleach, minerals
- Hardness in water
- Metal content of alloys, minerals, ores, clays, waters, plating baths, paints, paper, plant materials, biological fluids, petroleum products
- Moisture content in foodstuffs, petrochemicals, pharmaceutical products, and plastics

• Redox reagent concentrations such as available chlorine in potable water, peroxide, traces of oxidants and reductants in food, reductants in high temperature or high pressure boiler water, vitamin analysis

1.3. ADVANTAGES AND DISADVANTAGES OF TITRATIONS

Some advantages of titrations as an analytical technique are:

- More precise results than many instrumental methods, such as measurement by electrode, the accuracy of the measurement is up to 0.1%
- Simple methods, reasonable capital costs, and easy training
- Suitability to measure major components of a mixture or product
- Automation can reduce time and labor spent on each analysis

Some disadvantages of titrations are:

- Time it takes to prepare standards and titrants
- Good technique is required to achieve precise results (training and practice required)
- Not suitable for determining trace or minor components of a mixture or product
- Limited dynamic range, it may require additional sample preparations (dilution) and repeat analyses

2. TYPES OF TITRATIONS

2.1. TITRATIONS ACCORDING TO THE MEASUREMENT METHOD

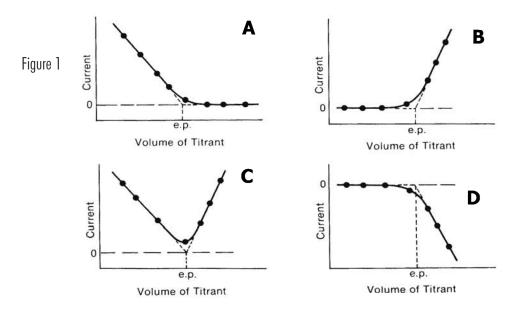
2.1.1. AMPEROMETRIC TITRATIONS

An amperometric titration is performed by placing two electrodes (often a metal ISE and a reference electrode) into the sample solution and holding the potential of the metal electrode at a selected voltage. The current that flows, due to the oxidation or reduction of a reactant or product, is plotted vs. volume of titrant to provide the titration curve and locate the equivalence point. Changes in the current are due to changes in the concentration of a particular species (being oxidized or reduced at the electrode).

Generally the reaction between the analyte and titrant forms a new species. Depending on the titration, the reactants are electroactive and the products are not, or vice-versa. Amperometric titration curves look like two straight lines intersecting at the equivalence point, this is due to the change in the electroactivity of the solution.

Many metal ions can be amperometrically titrated using a precipitation, complexation or redox reaction. Some metal ions and species that can be determined in this manner include silver, barium, halides, potassium, magnesium, palladium, molybdate, sulfate, tungstate, zinc, bismuth, cadmium, fluoride, indium, thallium, iodine, and gold.

Figure 1 shows four amperometric titrations and their endpoints. In graph "A" the analyte is electroactive and gives current but the reacted species does not. In "B" the reactant is not active but the titrant is. In "C" both the analyte and titrant are active and both give current flow. Graph "D" shows the same situation as "B"; however, the current has an opposite sign (the titrant is reduced).



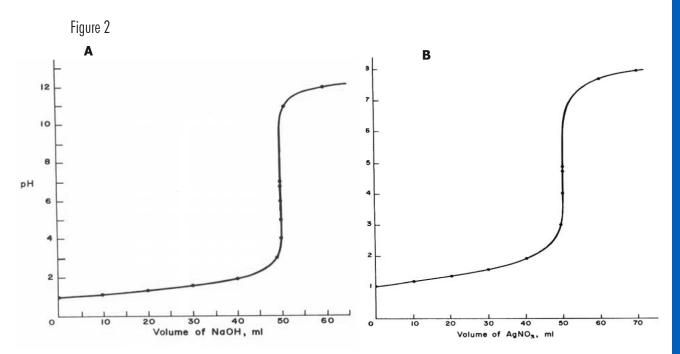
2.1.2. POTENTIOMETRIC TITRATIONS

Potentiometric titrations are done by measuring the voltage across the solution using an electrode system. An electrode system consists of an indicator electrode and a reference electrode. As titrant is added the variations in the potential of the indicator electrode, with respect to the reference electrode, are monitored to show the progress of the titration.

Potentiometry is the measurement of a potential under conditions of zero current flow. The measured potential can then be used to determine the analytical quantity of interest, generally a component concentration of the analyte solution. The potential that develops in the electrochemical cell is the result of the free energy change that would occur if the chemical phenomena were to proceed until the equilibrium condition has been satisfied.

There are many types of titrations where potentiometry can be used,e.g., pH electrodes for acid-base titrations, platinum ORP electrodes in redox titrations, ion selective electrodes, such as chloride or fluoride for a specific ion titration, and silver electrodes for argentometric (silver-based) titrations.

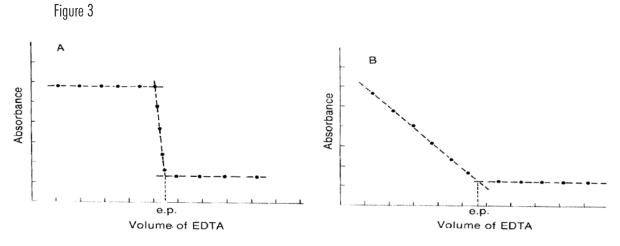
An example of potetiometric titrations are shown below. Figure 2 "A" is the pH of a solution vs. the volume of titrant and "B" is the potential from a chloride electrode vs. the volume of $AgNO_3$.



2.1.3. SPECTROPHOTOMETRIC TITRATIONS

The name comes from the method used to detect the endpoint of the titration, not its chemistry. Highly colored indicators that change color during the course of the titration are available for many titrations. More accurate data on the titration curve can be obtained if the light absorption is monitored instrumentally using a light source, a simple monochromator and a photodetector, rather than visually determining the color or light absorption change. Light absorption by either an indicator or by one of the reactants or products can be used to monitor the titration.

In the first titration curve, Figure 3 "A", the absorption of a metal-indicator complex is being monitored. The absorption is constant while the metal is complexed by the EDTA titrant. The metal indicator complex was stripped, causing a sharp break in the titration curve. The point where all the metal is complexed and stripped from the indicator is the equivalence point. This point is marked by "e.p." on the graph. In the second titration curve, Figure 3 "B", the metal complex is being measured while being titrated with EDTA. The new complex being formed is not colored and does not absorb light. The extrapolated intersection of the two lines determines the equivalence point.



7

TITRATION THEORY

2.2. TITRATIONS ACCORDING TO THE REACTION TYPE

2.2.1. ACID-BASE TITRATIONS

Acid—base titrations are the most common type of titrations. They are based upon a reaction between an acid and a base, a stoichiometric neutralization, or the exchange of protons. Virtually all acid-base titrations are carried out using a strong acid or a strong base as the titrant. The endpoint of a titration carried out with a weak acid or a weak base would be difficult to detect due to a small change in pH at the equivalence point.

Chemical indicators can be used to determine the endpoint. The indicator will change color to signify that the end of the titration has been reached. The color of the indicator is dependent upon the concentration of ions in the solution. An acid-base indicator is composed of a conjugate weak acid-weak base pair, where the two forms exhibit different colors depending on the pH of the solution. For an indicator, the acid ionization constant K_n is usually written as:

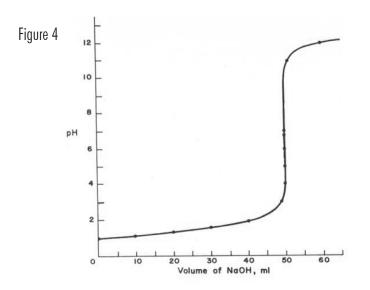
$$K_{a} = \frac{[H_{3}O^{+}][In^{-}]}{[HIn]}$$

HIn is the acid form of the indicator and In⁻ is the base form. At the center of the change region, the ratio of [In⁻] to [HIn] is one, $[H_3O^+] = K_a$ and $pH = pK_a$. The color change region is usually ± 1 pH unit around this point. Table 1 contains a list of some aqueous acid-base chemical indicators, as well as the pH range, the pK_a and the expected color (acid and base form). When choosing the proper indicator you should select one that has a pK_a as close to the endpoint of the titration.

When chemical indicators are not suitable, a potentiometric pH titration can also be used. The pH of the solution is plotted versus the volume of titrant added. Figure 4 shows a traditional strong acid-strong base titration curve. The graph shows the volume of NaOH added to an acidic solution and the resulting pH of the solution. Note the abrupt change in the pH at the equivalence point.

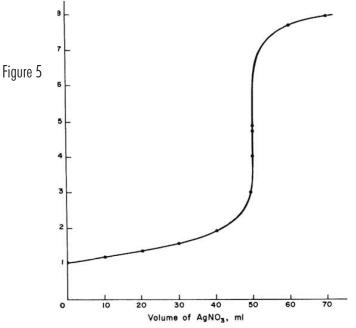
pH Range	Indicator	рКа	Acid Form	Base Form
0.0 - 1.6	Methyl Violet		Yellow	Blue
1.2 - 2.8	Thymol Blue	1.65	Red	Yellow
3.2 - 4.4	Methyl Orange	3.46	Red	Yellow
3.8 - 5.4	Bromocresol Green	4.90	Yellow	Blue
4.8 - 6.0	Methyl Red	5.00	Red	Yellow
5.2 - 6.8	Chlorophenol Blue	6.25	Yellow	Red
6.0 -7.6	Bromothymol Blue	7.30	Yellow	Blue
6.6 - 8.0	Phenol Red	8.00	Yellow	Red
7.4 -9.0	Metacresol Purple	8.30	Yellow	Purple
8.0 - 9.6	Thymol Blue	9.20	Yellow	Blue
8.2 - 10.0	Phenolphthalein	9.50	Clear	Pink
9.4 -10.6	Thymolphthalein		Clear	Blue
10.1 - 12.0	Alizarin Yellow R		Yellow	Red
11.4 - 12.6	Indigo Carmine		Blue	Yellow

Table 1



2.2.2 ARGENTOMETRIC TITRATIONS

Argentometric titrations use silver (nitrate) as the titrant and are generally precipitation titrations, as many silver salts are insoluble. These titrations are commonly used to titrate and determine the concentration of bromide, chloride, cyanide, iodide, and sulfide. Argentometric titrations can be done with Mohr's indicator (when all of the chloride has reacted, a red silver chromate precipitate is formed) or the titration can be easily followed with a silver ISE (or chloride ISE for chloride titrations) and a reference electrode. Figure 5 shows the titration of 50 mL of 0.1N NaCl with 0.1N AgNO₃. The potentiometric signal is from a chloride ISE and is plotted as pCl (- log [Cl-]).

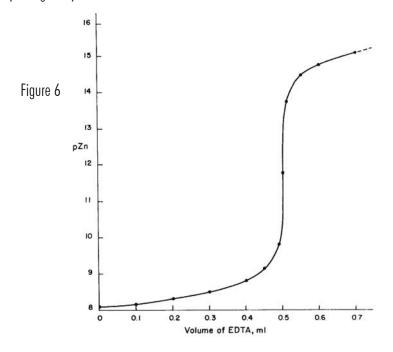


2.2.3. COMPLEXOMETRIC TITRATIONS

A complex is a species where a central metal ion is covalently bonded to one or more electron donating groups called ligands. In a complexometric titration, metal ions are titrated using a titrant that binds strongly to it. Often these titrants contain EDTA or CDTA, polydentate ligands that form very stable coordination compounds with metal ions. The complexation reaction must be fast in order to be useful for direct titration. Some metal ions react too slowly with EDTA for a direct titration.

IITRATION THEORY

An indicator electrode that responds to the metal ion can be used to monitor the titration progress. The titration curve will appear similar to a usual potentiometric titration. Complexation indicators change color at the endpoint as all metal ions are "consumed", or complexed, by the titrant. The titration curve will appear similar to a potentiometric titration when using an indicator electrode that responds to the metal ion (see Figure 6).



2.2.4. ION SELECTIVE TITRATIONS

The most popular ion selective titration is an acid-base titration. The hydrogen ion concentration is specifically measured and monitored during the titration process to locate the equivalence point. Using an ion selective electrode (ISE) as the indicator electrode, the potentiometric signal (in mV) is used to directly follow a specific ion's concentration (or activity).

Examples of ISE titrations include titrating fluoride with an aluminum titrant using a fluoride ISE, chloride with silver nitrate using a chloride ISE, sodium with a sodium ISE, etc. The equivalence point can be determined by plotting the mV value vs. the amount of titrant added.

2.2.5. NON-AQUEOUS SOLVENT ACID-BASE TITRATIONS

Non-aqueous solvents must be used to titrate very weak acids and bases due to the inherent leveling effect water has on all acids and based dissolved in it. A wide variety of weak acids and bases can be titrated using non-aqueous solvents. Mixtures of acids or bases can often be individually analyzed in a single sequential titration.

TITRATION OF ACIDS

Weak acids with pK_a's up to about 11 can be titrated in non-aqueous solvents. These include carboxylic acids, enols, phenols, imides, sulfonic acids, and inorganic acids. Water or lower alcohols are suitable for titrating medium to strong acids (pK_a less than 5). Titrating a weaker acid with a strong base titrant requires a solvent less acidic than water or ethanol/methanol. Solvents such as acetone, acetonitrile, t-butyl alcohol, dimethylformamide, isopropanol and pyridine have been found to work well for acid-base titrations of strong, medium and weak acids/bases. Titrants include alcoholic potassium hydroxide and various sodium or potassium alkoxides in a 10:1 mixture of benzene/methanol. The best titrants are quaternary ammonium hydroxides (such as tetrabutylammonium hydroxide) due to good solubility of tetraalkylammonium salts of the titrated acids and the clean potentiometric titration curve obtained (see Figure 7). Figure 7 -200 -300 -400 -400 -500 -600 -600 -700 -800 -900 -1 2 3 4 5 ml of 0.1*M* tributylmethylammonium hydroxide

potential, mv

TITRATION OF BASES

Weak bases with pK's up to about 11, which do not ionize with water, can be titrated in non-aqueous solvents. These bases include aliphatic and aromatic amines, basic nitrogen heterocycles, alkali metal and amine salts of acids, and many other organic basic compounds. Titrating a weak base with a strong acid titrant requires a basic solvent that is as weak as possible. Water and alcohols allow the titration of medium strength bases such as aliphatic amines (pK $_{\rm b} = 4$ to 5), but not the titration of weaker bases such as pyridine ($pK_{b} = 8.8$). Glacial acetic acid works well for weak bases and has been used extensively. Less basic solvents such as acetone, acetonitrile, and nitromethane extend the range of titrable compounds.

The endpoint for non-aqueous titrations are usually determined potentiometrically using a pH glass electrode, a modified calomel or double junction reference electrode with a low-flow rate reference junction. Good potentiometric titration curves are obtained in most solvents, except those with very low dielectric constants such as benzene, chloroform and others, when high electrical resistance of the solvent causes unstable potentials.

2.2.6. PRECIPITATION TITRATIONS

Precipitation titrations allow for faster analysis compared to the old gravimetric analysis, where a precipitate is formed, filtered, dried and weighed to analyze a compound. Typically silver halides, silver thiocyanate and a few mercury, lead, and zinc salts are titrated using this method. The chemical reactions must form an insoluble salt and precipitate out quickly in order to be analyzed by this method. When the reaction is not quick, a back titration can be used. A measured excess of the precipitating reagent (titrant) is added to force the reaction to occur, and then unreacted titrant is then titrated with a standard solution of another reagent.

2.2.7. REDOX TITRATIONS

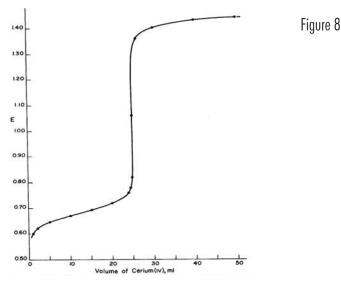
There are a number of oxidation-reduction reactions that can be used to determine unknown concentration by titration. If the reaction goes to completion, is fast and has an analytical signal available to follow it, a titration can be performed. The term "fast" means that each addition of titrant is reacted completely and the sensing electrode is able to detect the change in solution in less than one second.

Redox titrations are potentiometric titrations where the mV signal from a combination ORP (redox) electrode (usually with a platinum indicator electrode) is used to follow the reaction of oxidant/reductant. The electrode potential is determined by the Nernst equation and is controlled by the oxidant reductant ratio.

Visual indicators such as Ferrion are also available. The oxidized and reduced form of the indicator will have different colors and can be used to determine the end point.

Various reductants can be determined by titrants with oxidants such as potassium permanganate, potassium chromate or iodine. Commonly used reductants that are used as titrants include sodium thiosulfate, and ferrous ammonium sulfate.

As with Acid-Base titrations the potential changes dramatically at the equivalence point.



2.2.8. KARL FISCHER TITRATIONS

This method is based on a well-defined chemical reaction between water and the Karl Fischer reagent. The chemistry provides excellent specificity for water determination. The method can be used to determine free and bound water in a sample matrix. The Karl Fischer method is widely considered to produce the most rapid, accurate and reproducible results and has the largest detectable concentration range spanning 1 ppm to 100%.

The determination of water content is one of the most commonly practiced methods in laboratories around the world. Knowledge of water content is critical to understanding chemical and physical properties of materials and ascertaining product quality. Water content determination is conducted on many sample types including pharmaceuticals and cosmetics, foods and natural products, organic and inorganic compounds, chemicals, solvents and gases, petroleum and plastic products as well as paints and adhesives. The KF method is verifiable and can be fully documented. As a result, Karl Fischer titration is the standard method for analysis of water in a multitude of samples as specified by numerous organizations including the Association of Official Analytical Chemists, the United States and European Pharmacopoeia, ASTM, American Petroleum Institute, British Standards and DIN.

2.3. TITRATIONS ACCORDING TO THE TITRATION SEQUENCE

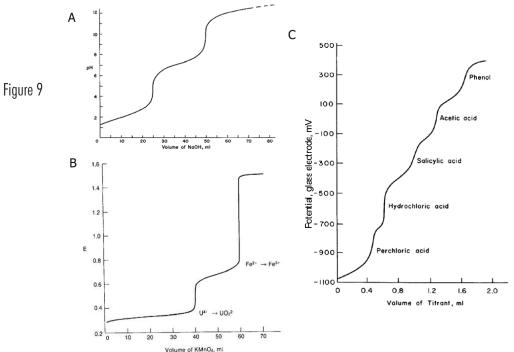
2.3.1. BACK TITRATIONS

Back titrations are generally used when a reaction is too slow to be directly accomplished using a "direct" titration, where the reaction goes to completion within a few seconds. In a back titration, a large excess of a reagent is added to the sample solution, helping a slow reaction to go to completion. The unreacted, excess reagent is then titrated. The difference in the total volume of the first reagent added and amount determined from the second titration is the quantity of reagent required to complete the first reaction.

2.3.2. MULTIPLE ENDPOINT TITRATIONS

Under certain conditions, some titrations can exhibit more than one equivalence point and be titratable to the individual endpoints to determine the concentration of each individual component. Examples of these types of titrations include acid-base (where different strength acid or bases are in a mixture), redox (where each species has a different reduction potential), complexometric (where different species are separately titratable), and acid-base using polyprotic acids (the pK_a of the different protons varies enough to separate them).

Figure 9 shows three different types of multiple endpoint titrations. "A" shows the titration of a polyprotic acid. The different acid strengths of the first and second proton can be determined. "B" illustrates a mixture of two different metal redox species, where the different redox potentials allow the species to be separated. "C" is the titration of a solution containing strong, weak, and very weak acids.



3. INTRODUCTION TO TITRATION APPARATUS AND TYPICAL TITRATION PROCEDURE

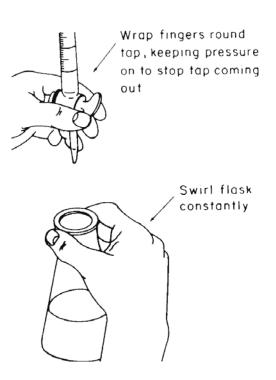
3.1. MANUAL TITRATION

Apparatus required for manual titration include:

- Volumetric Burette, for precisely controlled delivery of titrant to the reaction vessel
- An Erlenmeyer, or similar flask, that facilitates constant mixing or swirling required to ensure solution homogeneity
- Volumetric pipettes for the precise addition of samples and indicator solutions
- Titrant solutions of known concentration
- A visual or instrumental indicator for detecting the completion of the reaction

A typical manual titration consists of the following steps:

- 1. A volumetric pipette is typically used to add a known volume of sample to the flask
- 2. An indicator solution or instrument probe is added to the flask
- 3. A burette is used to measure the addition of titrant to the flask and dispense titrant in a controlled manner
- 4. Titrant is added via the burette until the method indication signals the reaction endpoint
- 5. The concentration of analyte is calculated based on the concentration and volume of titrant required to reach the endpoint



3.2. AUTOMATIC TITRATION

Automatic titrators are high-precision analytical instruments that deliver the titrant, monitor the physical change associated with the titration reaction, automatically stop at the endpoint and calculates the concentration of the analyte. Automatic titrators are best for repetitive titrations and high-accuracy analyses.

An automatic titrator must have an accurate liquid dispensing system. In high accuracy systems like the H1900-series titrators, the liquid dispensing system consists of a stepper-motor driven piston syringe burette capable of accurately and precisely dispensing very small volumes of titrant, a valve system to switch between titrant intake and outlet and a dispensing tip. These three main subsystem components must be as accurate as possible, with very low gear backlash in the burette pump, minimal piston seal flexing, precision ground inner diameter of the glass syringe, a low dead volume valve, minimal evaporation/permeation, and chemically resistant tubing.

Apparatus required for automatic titration include:

- An automatic titrator, equipped with a burette
- A beaker
- An electronic stirring system, either a propeller stirrer or a magnetic stir bar and stir plate
- Volumetric pipettes for the precise addition of samples
- Standard titrant solutions of known concentration
- An electrode system that can be used to determine the endpoint of the titration

A typical automatic titration consists of the following steps:

- 1. Set up the automatic titrator according to the manufacturer's instructions
- 2. A volumetric pipette is typically used to add a known volume of sample to the beaker
- 3. Submerge the propeller stirrer or add the stir bar to the beaker, and turn on
- 4. Start the titration, the titrator will automatically stop at the endpoint and determine the concentration of the analyte

4. TITRATION RESULTS

4.1. ACCURACY

The factors most critical to achieving accurate results with the H1932 titration systems are the concentration of the sample, size of the sample and having an optimized set of method parameters.

4.2. **REPEATABILITY**

Repeatability, or the agreement between replicate determinations, is expressed quantitatively as the relative standard deviation (RSD).

4.3. SOURCES OF ERROR

One of the advantages of volumetric analysis is excellent accuracy and precision. The sources of error can be grouped into sampling, titrant and standards, chemical reactions, endpoint determination and calculations.

4.3.1. SAMPLING ERRORS

- Selection of a non-homogeneous or non-representative sample
- Sample changed or was contaminated during collection, storage or transfers
- Poor technique when transferring sample to beaker or flask
- Errors in the balance, calibrate and check balance regularly

4.3.2. ERRORS WITH TITRANT AND STANDARD

4.3.2.1. PREPARATION ERRORS

Incorrect preparation due to:

- Poor technique in weighing the salt or when transferring to volumetric glassware
- Low-purity of salts or water used to make titrant and standard
- Dirty or wet glassware
- Improper storage of titrant or standard which allows water gain, evaporation or deterioration
- Failure to standardize frequently to adjust for change in titrant
- Failure to flush titrator tubing with a volume of titrant before standardizing
- Volume errors from pipettes and volumetric flasks, grade A glassware is required
- Balance errors when weighing out salts, calibrate and check balance regularly

4.3.2.2. DISPENSING ERRORS

Incorrect dispensing due to:

- Dead valve volume and leaking valve
- Inaccuracy in motor drive and gear lash/ backlash
- Poor burette/ piston seal
- Non-uniform diameter of burette glass cylinder
- Chemical incompatibility with tubing or bubble generation
- Density/ temperature changes in titrant

4.3.3. CHEMICAL REACTION ERRORS

- Inappropriate solvent or sample resulting in side reactions
- Poor mixing of the titrant and solvent or sample in the titration vessel
- Reaction between titrant and sample is not rapid
- Reaction does not go to completion
- Reaction has side reactions

4.3.4. ENDPOINT DETERMINATION ERRORS

Most manual titrations use a visual indicator to indicate when the endpoint is reached and the titration should be stopped. Automatic titrators use instrumental methods to determine the end of a titration and the equivalence point. There are two predominant methods used to determine the equivalence point, first derivative and second derivative.

The inflection point of the titration curve (mV vs. Volume) is normally assumed to be the equivalence point. The first derivative is often used to determine the inflection point. The maximum value of the first derivative (dmV vs. dV) corresponds to the theoretical equivalence point. During a titration it is rare to have a data point exactly at the first derivative maximum, the maximum value is determined by interpolating the first derivative data points.

The second derivative ($d^2 \text{ mV}$ vs. dV^2) can also be used to determine the equivalence point, and can offer advantages over the first derivative method. Second derivatives have increased sensitivity to smaller inflection points and easier numerical evaluation of the actual equivalence point. The value where the second derivative is equal to zero is the equivalence point. The second derivative requires fewer points located near the equivalence point, where data is often not obtained or not as reliable.

Errors in determining the endpoint can result from:

- Incorrect signals from the sensor
- Sensor drift
- Sensor or instrument has slow response, keep sensors in good condition
- Inappropriate setting on the titrator

5. CALCULATIONS

The main variables used in calculating a result from a titration are the sample volume, the concentration of the titrant, and the volume of titrant required to reach the equivalence point. At the equivalence point, an equal number of equivalents of the analyte and titrant has been added.

5.1. SAMPLE CALCULATION

By Mass

$C sample = \frac{V titrant \times C titrant \times Ratio \times FW analyte}{m sample} \times 100$

C sample	Sample Concentration (g/100g)
V titrant	Volume of titrant (L)
C titrant	Titrant Concentration (eq/L)
Ratio	Equivalence ratio of analyte/ titrant (mol analyte/ eq titrant)
FW analyte	Formula Weight of the Analyte (g/mol)
m sample	Mass of sample (g)

By Volume

 $C sample = \frac{V \ titrant \times C \ titrant \times Ratio \times FW \ analyte}{V \ sample} \times 100$

Sample Concentration (g/100mL)
Volume of titrant (L)
Titrant Concentration (eq/L)
Equivalence ratio of analyte/ titrant (mol analyte/ eq titrant)
Formula Weight of the Analyte (g/mol)
Volume of Sample (mL)

5.2. STANDARDIZE TITRANT

Titrant standardization is the second most important calculation in titrations. A primary standard is titrated in order to determine the concentration of the titrant. This is essentially a typical titration calculated in "reverse", where the concentration of the solution is known and the titrant is the unknown.

By Mass

C titrant =	m standard × Ratio
C ilirani –	$\overline{FW \text{ standard} \times V \text{ titrant}}$

C titrant	Titrant Concentration (N)
m standard	Mass of Standard (g)
Ratio	Equivalence ratio of titrant/standard (eq titrant/ mol standard)
FW standard	Formula Weight of the Standard (g/mol)
V titrant	Volume of Titrant (L)

 $C titrant = \frac{V standard \times (1 L/1000 mL) \times C standard}{V titrant}$

C titrant	Concentration of titrant (N)
V standard	Volume of Standard (mL)
C standard	Concentration of standard (eq/L)
V titrant	Volume of Titrant (L)

5.3. BLANK TITRATION

In a blank titration a pre-titration is performed, often times on the solvent to be used for the sample titration, and the titrant volume required to reach the endpoint is noted. This blank value nullifies error due to titrant required to react with the components of the titration solution matrix. The basic titration equation can be used for a blank titration, with the single modification that the volume of titrant used in the blank titration should be subtracted from the regular titration titrant volume.

 $C sample = \frac{C \ titrant \times (V \ sample - V \ blank) \times Ratio \times FW \ analyte}{m \ sample} \times 100$

Sample Concentration (g/100g)
Titrant Concentration (eq/L)
Volume of Titrant required for the sample (L)
Volume of Titrant required for the blank (L)
Equivalence ratio of analyte/ titrant (mol analyte/ eq titrant)
Formula Weight of the Analyte (g/mol)
Mass of sample (g)

5.4. MULTIPLE ENDPOINT TITRATION

Some titrations have two or more endpoints, each corresponding to the equivalence point for a specific reaction. Multiple endpoint titrations are similar to a blank titration in that the volume of titrant required to reach the first endpoint is subtracted from the titrant volume used to reach the next sequential endpoint.

 $C sample 1 = \frac{V \ titrant \ 1 \times C \ titrant \ \times Ratio \ \times FW \ analyte \ 1}{m \ sample} \times 100$

 $C sample 2 = \frac{(V \ titrant \ 2 - V \ titrant \ 1) \times C \ titrant \times Ratio \times FW \ analyte \ 2}{m \ sample} \times 100$

 $C sample 3 = \frac{(V \ titrant \ 3 - V \ titrant \ 2) \times C \ titrant \times Ratio \times FW \ analyte \ 3}{m \ sample} \times 100$

C sample1	Sample 1 Concentration (g/100g)
C sample2	Sample 2 Concentration (g/100g)
C sample3	Sample 3 Concentration (g/100g)
V titrant 1	Volume of titrant required to reach the first end point (L)
V titrant 2	Volume of titrant required to reach the second end point (L)
V titrant 3	Volume of titrant required to reach the third end point (L)
C titrant	Concentration of titrant (N)

Ratio	Equivalence ratio of analyte/ titrant (mol analyte/ eq titrant
FW analyte 1	Formula Weight of the Analyte 1 (g/mol)
FW analyte 2	Formula Weight of the Analyte 2 (g/mol)
FW analyte 3	Formula Weight of the Analyte 3 (g/mol)
m sample	Weight of Sample (mL)

5.5. BACK TITRATION

The equation used in back titration calculations is also similar to the equation for a blank titration. Instead of subtracting the initial amount of titrant needed to react with the blank, the amount of second titrant needed to react with the excess titrant added in the first titration is subtracted from the amount of the first titrant added. The difference between the two amounts is the amount of titrant necessary to reach the first equivalence point.

 $C sample = \frac{(C \ titrant \ 1 \times V \ titrant \ 1 - C \ titrant \ 2 \times V \ titrant \ 2) \times Ratio \times FW \ analyte}{V \ sample} \times 100$

C sample	Sample Concentration (g/100mL)
C titrant 1	Concentration of titrant 1 (N)
V titrant 1	Volume of titrant 1 (L)
C titrant 2	Concentration of titrant 2 (N)
V titrant 2	Volume of titrant 2 (L)
Ratio	Equivalence ratio of analyte/ titrant (mol analyte/ eq titrant)
FW analyte	Formula Weight of the analyte (g/mol)
V sample	Volume of sample (mL)

6. GLOSSARY

Acid

A chemical species that can donate one or more protons (hydrogen ions).

Acid-Base Titration

Stoichiometric neutralization titrations, based upon the reaction that occurs between an acid and base.

Activity

A physical property corresponding to the concentration of all ions in a solution. Electrodes respond to activity.

Amperometric Titration

Titrations where the current flow between two electrodes (often a metal electrode and a reference electrode) are used to monitor the titration progress.

Analyte

The chemical species being measured in a titration.

Argentometric Titration

Titrations that use silver (nitrate) as the titrant. These titrations are typically precipitation titrations.

Automatic Titrator

An instrument designed to automatically carry out a titration. It will add the appropriate amount of titrant, determine the endpoint and calculate the results.

Back Titration

A type of titration where an excess amount of titrant is added to a sample, forcing a sluggish reaction to go to completion. The excess reagent is then "back" titrated with a second titrant.

Base

A chemical species that can accept one or more protons (hydrogen ions).

Biamperometric Indication

Uses a double platinum pin electrode to measure the current flow through a titration solution.

Bivoltametric Indication

Uses a double platinum pin electrode to measure the voltage required to maintain a constant current flow through a titration solution while constant voltage is applied across the platinum elements of the electrode.

Burette

A graduated cylindrical piece of laboratory glassware that is used to dispense precise amounts of solution.

Complex Ion

A species where a central metal ion is covalently bonded to one or more electron donating groups called ligands.

Complexometric Titrations

Metal ions are titrated using a titrant that binds strongly to it. The titrants often contain Ethylenediaminetetraacetic Acid (EDTA) or Cyclohexylenedinitrilotetraacetic Acid (CDTA).

Endpoint

The point were a titration is stopped because a physical change in the solution has indicated a completed titration. Titration endpoints typically coincide with the equivalence point. A fixed value endpoint (pH or mV) can be used as well. The titration will stop at the desired point regardless if the titration is complete.

Equivalence point

The point where the quantity of titrant is stoichiometrically equal to the quantity of analyte.

Formal

The theoretical number of equivalents per liter of the solution. It is used in solutions where the exact concentration of a species may be affected by the other ions present, therefore the stated concentration may not be exactly correct.

Gravimetric Analysis

A quantitative determination of an analyte based on the mass of the solid.

Indicator Electrode

An electrode that responds to the species of interest. The electrode potential is proportional to the concentration or activity of that ion in the solution being measured.

Indicators

Chemical indicators are typically organic dyes that change form under different physical conditions, causing a color change that can be seen by an analyst. Typically used in manual titrations, chemical indicators have been replaced with electrometric indicators, which are used with automatic titrators.

Inflection Point

The point on a titration curve were the second derivative curve changes signs.

Ion Selective Electrode (ISE)

An electrode that responds to a specific ion. The electrode potential is proportional to the concentration or activity of that ion in the solution being measured.

Karl Fischer Titration

A titration that uses a chemical reaction that is specific for determining water.

Manual Titration

A titration that is carried out by hand. The analyst must add the appropriate amount of titrant, determine the endpoint and calculate the results.

Molar

The concentration of a solute in a solution.

Mole (mol)

A quantity of a chemical species. The molecular weight of a substance in grams is equal to the mass of one mole of the substance. One mole is equal to 6.022 x 1023 atoms or molecules.

Monochromator

A device that allows only a narrow range of wavelengths to pass though it by separating the light into different wavelengths.

Multiple Endpoint Titration

A titration that reacts multiple species in solution sequentially using the same titrant. The concentration of each analyte can be determined from their respective endpoints.

Nernst Equation

The fundamental equation relating cell voltage to the concentration of a solution.

Neutralization

A chemical reaction where an acid and a base react to form a neutral salt and water.

Non-aqueous

A solution that does not contain water.

Non-aqueous Titration

A titration that is preformed in non-aqueous solutions, typically used to titrate very weak acids and bases to eliminate the leveling effect water has on all acids and bases dissolved in it.

Normal

The concentration of a solution which accounts for any stoichiometric difference between the various species in a solution.

Oxidation / Reduction Potential (ORP)

The measurement describing whether a species wants to donate or accept electrons from other species in a redox reaction. If a solutions reduction potential is higher than the species it is reacting with, it will typically gain electrons or be reduced. If the potential is lower than the species it is reacting with, it will typically lose electrons or be oxidized.

Oxidant

The species that is accepting electrons in a redox reaction.

Pipette

Scientific apparatus that is used to deliver precise volumes of liquids.

Polyprotic Acid

Acids that are capable of donating more than one proton per acid molecule.

Potentiometric Titration

A titration in which the endpoint is determined by monitoring the voltage of the solution using an electrode.

Precipitation Titration

A titration in which the analyte reacts with the titrant to form an insoluble compound. The endpoint is typically detected with an ISE sensitive to either the analyte or titrant.

Reagent

The chemical added in a titration that causes the given reaction to occur.

Reduction-Oxidation Reaction (redox)

A chemical reaction in which the atoms involved in the reaction have their oxidation numbers changed. Reduction is the gain of electrons, which decreases the oxidation number. Oxidation is the loss of electrons, which increases the oxidation number.

Reductants

The electron donor in a redox reaction.

Reference Electrode

An electrode that supplies a constant electrode potential. It is used in combination with an "indicator" electrode, allowing for the "indicator" electrode potential to be measured.

Relative Standard Deviation (RSD)

A measure of the amount of relative variation in a set of data. It is calculated by dividing the standard deviation by the mean: RSD = (Standard Deviation of X) * 100 / (Mean of X)

Repeatability

The variation in sample measurements taken by a single person or instrument under the same conditions.

Spectrophotometric Titration

A titration in which the endpoint is marked by a change in the color and/or color intensity.

Stoichiometry

The quantitative relationship of the reactants and products in a chemical reaction.

Titrant

The chemical added in a titration that causes the given reaction to occur.

Titration

A quantitative, volumetric procedure used in analytical chemistry to determine the concentration of an analyte in solution. The concentration of the analyte is determined by slowly adding a titrant to the solution. As the titrant is added, a chemical reaction between the titrant and the analyte occurs.

Titration Curve

A graph containing the physical data obtained for a titration. The data plotted is often an independent variable (volume of titrant) vs. a dependent variable (pH of the solution). From the titration curve, the equivalence point or endpoint can be determined.

World Headquarters

Hanna Instruments Inc. Highland Industrial Park 584 Park East Drive Woonsocket, RI 02895 USA www.hannainst.com

Local Office

Hanna Instruments Inc. Highland Industrial Park 584 Park East Drive Woonsocket, RI 02895 USA Phone: 800.426.6287 Fax: 401.765.7575 e-mail: tech@hannainst.com

