



Letter to the Editor

TR2/TR4 overexpression in a humanized sickle cell disease mouse model decreases RBC adhesion to VCAM-1


To the Editor:

Vaso-occlusion is a hallmark of sickle cell disease (SCD) accounting for much of its morbidity and mortality. The initiation and propagation of vaso-occlusion results from processes that impair blood flow through the microvasculature. In particular, sickle erythrocytes (SSRBCs) have a greater propensity to adhere to the vessel wall than non-sickle erythrocytes [1]. SSRBC adhesion may directly occlude microvessels or indirectly by delaying the transit time of SSRBCs through the capillary and propagating intra-capillary sickling. Very late antigen-4 (VLA-4) or $\alpha 4\beta 1$ integrin is one of the most characterized RBC adhesion molecules and supports adhesion between SSRBCs and endothelial vascular cell adhesion molecule 1 (VCAM-1) [2,3]. VLA-4 is expressed on immature RBCs/reticulocytes, and these cells are found in increased numbers in the peripheral blood of SCD patients [2,3].

Hydroxyurea (HU), a known inducer of fetal hemoglobin (HbF), is the only FDA-approved therapy for SCD and has been shown to improve morbidity and mortality in patients with SCD. There is increasing evidence suggesting that HU achieves this by decreasing the adhesive properties of SSRBCs to the endothelium and sub-endothelial matrix [4]. Not all patients respond to HU therapy, and HU is not well tolerated amongst all the responders. Thus, there is a need for alternative approaches to increase HbF levels to achieve similar or better clinical efficacy. Previous studies have shown that forced transgenic expression of TR2/TR4, a nuclear orphan hormone receptor that binds to the γ -globin gene promoter, in sickle cell mice leads to induction of the γ -globin gene [5]. Subsequent analysis revealed that elevated TR2/TR4 levels enhanced HbF production and improved the disease phenotype within SCD mice [5]. These findings suggest that TR2/TR4 may be a suitable therapeutic target to induce persistent HbF accumulation in patients with SCD.

The current study was designed to determine whether alleviation of the disease phenotype was due, in part, to a reduction in SSRBC adhesion to VCAM-1. A microfluidic-based flow system was utilized to measure erythrocyte adhesion to VCAM-1 during physiologic flow. Flow adhesion assays were performed with a flow system (Bioflux 1000Z; Fluxion, San Francisco, CA) utilizing the 48-well plate format. RBCs were perfused through VCAM-1 coated channels, as previously described [6]. SSRBCs from SCD:Tg^{TR2/TR4} mice demonstrated significantly lower adhesion to VCAM-1 when compared to SCD mice (69 ± 26 vs. 383 ± 80 RBCs/mm², respectively, $p = 0.02$).

The normal reticulocyte fraction in blood is generally low, 0.5–1%. Chronic hemolytic anemia, a consequence of SCD, results in elevated numbers of circulating reticulocytes. VLA-4 is expressed on reticulocytes in high numbers. VLA-4 expression is usually lost from mature

erythrocytes prior to release into the peripheral circulation. In SCD, “stress” reticulocytes are thought to leave the bone marrow prematurely in response to erythropoietic stress leading to reticulocyte-enriched RBCs in the circulation. Reticulocyte counts in WT, SCD, and SCD:Tg^{TR2/TR4} mice were measured by flow cytometry using thioazole orange for reticulocyte staining in whole blood. As previously reported [5], TR2/TR4 overexpression in SCD mice exhibited significantly lower reticulocyte levels when compared to SCD mice (23 to 50.7%, respectively, $p < 0.01$). Independent of SCD, reticulocyte-enriched RBCs from non-sickle cell patients are more adherent than mature erythrocytes from the same individual [7]. This is due to high levels of VLA-4 present on reticulocytes, which mediate adhesion to VCAM-1 on the endothelium and fibronectin within the sub-endothelial matrix. It is plausible that a reduction in circulating reticulocytes improves the disease phenotype in SCD:Tg^{TR2/TR4} mice by minimizing VLA-4/VCAM-1 interactions thus preventing subsequent vaso-occlusion. Erythrocyte adhesion levels positively correlated with % reticulocytes ($r = 0.911$, $p < 0.01$). Additionally, the positive correlation between erythrocyte adhesion and an indicator of vaso-occlusive end organ injury, spleen % body weight ($r = 0.718$, $p = 0.01$), suggests that adhesion may directly or indirectly mediate vaso-occlusive pathology. Interestingly the reticulocyte % also correlated with spleen % body weight ($r = 0.764$, $p = 0.01$).

The protective effect of TR2/TR4 overexpression may also be attributed, in part, to a reduction in RBC sickling. Previous findings demonstrate that increased levels of HbF reduces erythrocyte sickling which over time reduces phosphatidyl serine (PS) exposure and further lowers the adhesive potential of SSRBCs to the endothelium [8]. Our data demonstrates that higher HbF levels correlate with lower SSRBC adhesion to VCAM-1 ($r = -0.857$, $p = 0.01$) although future analysis aimed at directly evaluating the role that forced TR2/TR4 expression has on PS exposure is needed.

Our data demonstrate that forced transgenic TR2 and TR4 expression reduces SSRBC adhesion to VCAM-1, possibly by reducing the burden of VLA-4-containing reticulocytes. Targeting of non-globin proteins or transcription factors may ameliorate VLA-4 mediated adhesion and vasculopathy, suggesting that this may be a promising therapeutic approach to induce persistent HbF accumulation and for treatment of SCD vasculopathy.

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References

- [1] R.P. Hebbel, M.A. Boogaerts, S. Koresawa, H.S. Jacob, J.W. Eaton, M.H. Steinberg, Erythrocyte adherence to endothelium as a determinant of vasocclusive severity in sickle cell disease, *Trans. Assoc. Am. Phys.* 93 (1980) 94–99.
- [2] C.C. Joneckis, R.L. Ackley, E.P. Orringer, E.A. Wayner, L.V. Parise, Integrin alpha 4 beta 1 and glycoprotein IV (CD36) are expressed on circulating reticulocytes in sickle cell anemia, *Blood* 82 (12) (1993) 3548–3555.
- [3] R.A. Swerlick, J.R. Eckman, A. Kumar, M. Jeitler, T.M. Wick, Alpha 4 beta 1-integrin expression on sickle reticulocytes: vascular cell adhesion molecule-1-dependent binding to endothelium, *Blood* 82 (6) (1993) 1891–1899.
- [4] C.A. Hillery, M.C. Du, W.C. Wang, J.P. Scott, Hydroxyurea therapy decreases the in vitro adhesion of sickle erythrocytes to thrombospondin and laminin, *Br. J. Haematol.* 109 (2) (May 2000) 322–327.
- [5] A.D. Campbell, S. Cui, L. Shi, et al., Forced TR2/TR4 expression in sickle cell disease mice confers enhanced fetal hemoglobin synthesis and alleviated disease phenotypes, *Proc. Natl. Acad. Sci. U. S. A.* 108 (46) (2011) 18808–18813.
- [6] J. White, M. Lancelot, S. Sarnaik, P. Hines, Increased erythrocyte adhesion to VCAM-1 during pulsatile flow: application of a microfluidic flow adhesion bioassay, *Clin. Hemorheol. Microcirc.* 60 (2014) 201–213.
- [7] G.A. Barabino, L.V. McIntire, S.G. Eskin, D.A. Sears, M. Udden, Endothelial cell interactions with sickle cell, sickle trait, mechanically injured, and normal erythrocytes under controlled flow, *Blood* 70 (1) (Jul 1987) 152–157.
- [8] B.N. Setty, S. Kulkarni, A.K. Rao, M.J. Stuart, Fetal hemoglobin in sickle cell disease: relationship to erythrocyte phosphatidylserine exposure and coagulation activation, *Blood* 96 (3) (2000) 1119–1124.

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